

1 **Ancient DNA reveals differences in behaviour and sociality between brown**
2 **bears and extinct cave bears.**

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33 Running title: ***Investigating bear behaviour using ancient DNA***

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44 **ABSTRACT**

45 Ancient DNA studies have revolutionised the study of extinct species and populations, providing
46 insights on phylogeny, phylogeography, admixture and demographic history. However, inferences
47 on behaviour and sociality have been far less frequent. Here, we investigate the complete
48 mitochondrial genomes of extinct Late Pleistocene cave bears and middle Holocene brown bears
49 that each inhabited multiple geographically proximate caves in northern Spain. In cave bears, we
50 find that, although most caves were occupied simultaneously, each cave almost exclusively
51 contains a unique lineage of closely related haplotypes. This remarkable pattern suggests extreme
52 fidelity to their birth site in cave bears, best described as homing behaviour, and that cave bears
53 formed stable maternal social groups at least for hibernation. In contrast, brown bears do not
54 show any strong association of mitochondrial lineage and cave, suggesting that these two closely
55 related species differed in aspects of their behaviour and sociality. This difference is likely to
56 have contributed to cave bear extinction, which occurred at a time in which competition for caves
57 between bears and humans was likely intense and the ability to rapidly colonise new hibernation
58 sites would have been crucial for the survival of a species so dependent on caves for hibernation
59 as cave bears. Our study demonstrates the potential of ancient DNA to uncover patterns of
60 behaviour and sociality in ancient species and populations, even those that went extinct many tens
61 of thousands of years ago.

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66 **INTRODUCTION**

67 Behaviour and sociality represent key mechanisms allowing populations to rapidly adapt to
68 changing environments, to better exploit available resources, and also to resist pressures such as
69 predation or climatic extremes that may negatively affect survival probability. Conversely, some
70 behaviours could be maladaptive in certain contexts, particularly when populations are exposed to
71 new and/or rapidly changing selective pressures, and may ultimately lead to population or even
72 species extinction. Ancient animal remains can hold information on their behaviour and sociality.
73 Spatial and temporal patterns of association among individuals can be investigated using standard
74 paleontological and isotopic methods, and their relatedness can – at least in principle – be
75 determined using ancient DNA approaches. The later, however, may represent a considerable
76 technical challenge, as advanced DNA degradation will complicate recovery of suitable data that
77 allows fine-scale resolution of genetic relationships among sufficient numbers of individuals to
78 achieve statistical power.

79

80 Bears that lived in Eurasia during the Pleistocene represent a group that may be amenable to
81 behavioural investigations using ancient DNA. Two major species (or species complexes) were
82 widespread and sympatric in Pleistocene Eurasia: brown bears (*Ursus arctos*), that survived
83 through the last glacial maximum (LGM) and are currently widespread across the entire Holarctic
84 region; and the cave bear (*Ursus spelaeus* complex), an iconic representative of the Pleistocene
85 megafauna, that went extinct prior to the LGM (Pacher & Stuart 2009; Stiller *et al.* 2010; 2014).
86 For cave bears in particular, their habit to hibernate in caves has resulted in assemblages
87 consisting of the bones of thousands of individuals at some sites, providing the opportunity to

88 investigate uniquely well-defined fossil populations, deposited within an environment that
89 enhances DNA preservation (Hofreiter *et al.* 2015). Although ancient brown bear remains
90 typically occur at a much lower frequency in caves in comparison to cave bears, comprehensive
91 palaeontological surveys of some caves have produced sufficient samples for population-level
92 analysis (e.g. in Kurten 1968).

93

94 The factors that drove the cave bear to extinction have been subject to considerable study and
95 discussion (Kurten 1968, Grayson & Delpech 2003; Pacher & Stuart 2009; Stiller *et al.* 2010). In
96 agreement with palaeontological data, genetic studies of cave bears have found high genetic
97 diversity and a large and constant population size until 50,000 yBP, followed by a decrease until
98 its ultimate extinction around 24,000 yBP (Pacher & Stuart 2009; Stiller *et al.* 2010; 2014). Thus,
99 the onset of decline of cave bear populations would have started around 25,000 years before the
100 LGM, and is therefore not associated with any periods of substantial climatic change in Europe
101 (Stiller *et al.* 2010; 2014). Brown bears, in contrast, show no evidence of population size changes
102 coinciding with the cave bear population decline (Stiller *et al.* 2010). It has been argued that
103 human activities played a major role in cave bear extinction (Grayson & Delpech 2003; Knapp *et*
104 *al.* 2009; Münzel & Conard 2004; Bon *et al.* 2011; Stiller *et al.* 2014). However, explanations of
105 why human activities could have so profoundly affected cave bear populations and not brown
106 bear populations remain elusive. Differences in behaviour between the two species may have
107 played a role, but identifying such differences is challenging because many aspects of cave bear
108 behaviour remain uncertain. For example, paleontological studies of some cave bear caves have
109 identified multiple depressions (hibernation beds or *bauges*, as described by Koby in 1953) in the

110 cave floor that are thought to have been formed by hibernating bears. While this suggests
111 communal hibernation, it is uncertain whether these were social or even family groups, or rather
112 random assemblages of individuals forced together through competition for hibernation sites.
113 Although genetic data could allow testing of such hypotheses, only a few studies have examined
114 the population structure of cave bears at a local – i.e. individual cave – scale (Orlando *et al.* 2002;
115 Richards *et al.* 2008; Hofreiter *et al.* 2004; Bon *et al.* 2011). Moreover, these studies were all
116 based on short mtDNA fragments, which does not allow fine scale resolution of the genetic
117 relationship between individuals.

118

119 In this study, we investigate complete mitochondrial genome sequences generated from the
120 subfossil remains of multiple cave bears and brown bears from several caves in the North of
121 Spain (Fig. 1). Four of the cave bear caves are located in close proximity (within a radius of
122 10km) within the Serra do Courel mountains (NW Spain), while the fifth one is located 450 km
123 away in Navarra (NE Spain). The brown bear caves are also in close proximity (within a radius of
124 50km). In all cases, there are no apparent topographic barriers separating caves from one another.
125 Thus, for such large bodied and presumably highly mobile mammals as cave bears and brown
126 bears, movement between these caves would, in general, not have represented any significant
127 challenge. In cave bears, we find that, even though caves were occupied simultaneously, each
128 cave almost exclusively contained a unique clade of closely related haplotypes. This remarkable
129 pattern suggests that cave bears returned to the cave where they were born and formed stable
130 maternal social groups for hibernation. In brown bears, however, no such pattern is found
131 suggesting greater flexibility with regard to hibernation site in this closely related species. We

132 discuss the implications of these behavioural differences for the extinction of the cave bear, in
133 addition to the wider potential of ancient DNA for the study of behavioural ecology, sociality, and
134 extinction.

135

136

137 **MATERIALS AND METHODS**

138

139 *Methods overview*

140 We generated mitogenome sequences of cave bears and brown bears from their skeletal remains
141 found in the caves shown in Figure 1. These sequences were used alongside published sequences
142 obtained from GenBank to compare the maternal relatedness of individuals occurring within
143 caves with that occurring among caves using haplotype network analysis, phylogenetic analysis
144 and trait-phylogeny association tests. Finally, the ages of individuals were estimated using a
145 combination of ^{14}C and molecular dating. In particular, we investigated whether the occupation of
146 caves was likely simultaneous, or instead temporally separated.

147

148 All but one of the novel Spanish bear mitogenome sequences reported here were obtained in a
149 single experiment (we refer to as Experiment 1) that used hybridisation capture to enrich
150 sequencing libraries for mtDNA prior to high-throughput sequencing. The details of Experiment 1
151 are reported below. A single Spanish cave bear sequence (sample E-VD-1838), in addition to
152 sequences from seven bears from elsewhere in Europe, were obtained in separate experiments
153 that are described in Section 1 of the Supporting Information.

154

155 ***Sampling locations***

156 The focal specimens used in this study were excavated in caves within karstic systems in the
157 north-west of Spain, and were identified morphologically as either *U. spelaeus* or *U. arctos*. All
158 of these sites represent natural accumulations and none of the remains are in archaeological
159 context. Individual samples originated from different individual animals, identified based on age,
160 sex or spatial distribution of the remains. Initially, specimens from 19 cave sites were investigated.
161 These comprised 85 individuals from nine caves containing cave bear remains, and 24 individuals
162 from ten caves containing brown bear remains. Many of these failed initial screening to identify
163 samples that were likely permit recovery of the complete mitogenome sequence (see below),
164 which limited sampling to five brown bear caves and five cave bear caves (shown in Fig. 1). Full
165 details of the caves and samples investigated are provided in Section 2, Tables S1 & S2, and Fig.
166 S1 of the Supporting Information.

167

168 ***DNA extraction and sample screening***

169 All pre-amplification aDNA analyses were performed in dedicated aDNA laboratories at the
170 University of York (UK) or at the University of Potsdam (Germany). The compact part of bones,
171 either femur, tibia, ribs, skull fragments or teeth, were utilised for DNA extraction. Prior to
172 extraction, samples were UV irradiated for 10 minutes on each side and disposable cutting disks
173 attached to a rotating electric drill were used to remove the outermost bone surface. For each
174 sample, around 250 mg of cleaned bone was ground to powder using ceramic mortar and pestles.
175 DNA extraction followed the protocol of Rohland *et al.* (2010).

176

177 DNA extracts were screened for likely presence and quality of endogenous DNA by attempting to
178 PCR amplify 104bp and 126bp fragments of the mitochondrial control regions of cave bears and
179 brown bears, respectively, using the primers described in Hofreiter *et al.* (2004) and a novel
180 brown bear primer, UaF7 (5'-TCGTGCATTAATGGCGTG-3'). Amplification was assessed using
181 agarose gel electrophoresis and the authenticity of amplification products verified by Sanger
182 sequencing, carried out in both directions using an ABI 3130XL at the Sequencing Service SAI
183 (Servicios Centrais de Investigacion, University of A Coruña, Spain), followed by BLAST
184 alignment of the consensus sequences.

185

186 ***Sequencing library generation and hybridisation capture***

187 We generated individually barcoded Illumina sequencing libraries using 20µl of those extracts for
188 which short-amplicon PCR had previously been successful, following the protocol described in
189 Meyer & Kircher (2010) with the following modifications. First, the filtration step between the
190 blunt end repair and the adapter ligation was substituted by heat inactivation of the enzymes
191 (Bollongino *et al.* 2013; Fortes and Paijmans 2015), in order to reduce the loss of short DNA
192 fragments. Second, we used a double index barcoding system in which both the P5 and P7
193 adapters include a molecular barcode specific for each sample (Kircher *et al.* 2011; Fortes and
194 Paijmans 2015). This facilitates the identification of chimeric molecules that could be formed
195 during PCR amplification of the captured products. Library indexing and amplification involved
196 4 replicate parallel PCRs, each using 15 cycles, which were then pooled and purified using silica
197 columns (Qiagen, France). The resulting cave bear and brown bear libraries were quantified using

198 a Nanodrop Spectrophotometer (Thermo Scientific) and pooled, respectively, in equimolar
199 quantities at a final concentration of 2 ng in 520 µl for hybridisation capture.

200

201 Hybridisation capture was carried out using 244k DNA SureSelect™ microarrays (Agilent,
202 Boblingen, Germany) with 2-fold tiling and 60bp probes. Separate arrays were used for the cave
203 bear and brown bear library pools, with probes based on published mitogenome sequences of a
204 Western European cave bear (EU327344, Bon *et al.* 2008) and brown bear (EU497665, Bon *et al.*
205 2008), respectively. Hybridisation capture followed the protocol of Hodges *et al.* (2009) with one
206 modification. After the initial round of capture enrichment, library pools were amplified using
207 primers IS5 and IS6 (Meyer & Kircher 2010) in 12 parallel PCRs and the resulting products were
208 subjected to a second round of capture enrichment, as described in Fortes & Paijmans (2015).

209

210 ***DNA sequencing and data processing***

211 100bp single-end sequencing of mtDNA enriched library pools was carried out on a single lane of
212 an Illumina HiSeq2000 instrument at the Danish National Sequencing Centre in the University of
213 Copenhagen. The resulting BCL files were converted to fastq format using the Illumina base-
214 calling pipeline (Illumina Pipeline v1.4). The program Cutadapt v1.3 (Martin, 2011) was then
215 used to trim any P7 adapter sequences occurring at the 3' ends of reads, and a custom script used
216 to identify and discard any reads that did not contain the appropriate P5 index, and then trim the
217 index sequence from the remaining reads. Following this procedure, any reads < 25 bp were also
218 discarded. The resulting cave bear and brown bear reads were then mapped to their respective
219 reference mitogenome sequences used for capture probe design, using bwa-0.5.9 (Li & Durbin

220 2009) with seeding disabled, as suggested by Schubert *et al.* (2012). The alignment was sorted,
221 filtered for minimum mapping quality (-q 30) and PCR duplicates removed using samtools (Li *et*
222 *al.* 2009). The Mpileup tool in samtools 0.1.19-44428 was used to to call polymorphic positions
223 and generate consensus sequences, using the -s option to specify a haploid genome. In order to
224 prevent miscalling of polymorphic sites resulting from the presence of postmortem molecular
225 damage to the ancient templates, the terminal five nucleotides at both 5' and 3' read ends were
226 excluded from SNP calling. Furthermore, polymorphic sites with very low coverage
227 (two or three reads) were only retained in the consensus if all reads showed
228 the same variant, otherwise, these sites were treated as missing data (marked
229 N). Polymorphic positions covered by only a single mapped read were also
230 treated as missing data. All polymorphic sites identified in the vcf file were further
231 checked by eye on Tablet version 1.13.05.02 (Milne *et al.* 2013). Read depth and coverage were
232 determined using GATK (MacKenna *et al.* 2010). The presence of molecular damage
233 characteristic of aDNA was confirmed using the software MapDamage (Ginolhac *et al.* 2011).

234

235 ***Phylogenetic and network analysis***

236 Only those novel sequences that provided > 70% total coverage of the mitogenome were used in
237 subsequent analyses. Forty-two novel Spanish sequences were aligned along with seven novel
238 sequences from ancient bears found elsewhere in Europe and 174 published mitogenome
239 sequences from cave bears, brown bear and polar bears using the program MUSCLE (Edgar &
240 Robert 2004) with default settings. A repetitive section of the d-loop was removed from the

241 alignment as this was not recovered in many ancient samples and even when present could not be
242 aligned unambiguously. All subsequent analyses used this alignment or subsamples of it.

243

244 To investigate the phylogenetic relation of Spanish cave bear and brown bear haplotypes to those
245 occurring elsewhere in their respective distributions, we conducted phylogenetic analysis of the
246 complete alignment under maximum likelihood (ML) using RAxML-HPC2 8.2.3 (Stamatakis,
247 2014) on the CIPRES Portal (Miller *et al.* 2010) using the American black bear (*U. americanus*)
248 as outgroup. We selected the GTR model with substitution rate heterogeneity as suitable because
249 this model offers greater flexibility in comparison to other time-reversible substitution models,
250 and the variability of our dataset (2,838 variable sites) is sufficient for all six parameters of the
251 GTR substitution matrix to be estimated accurately. Clade support was assessed using 500
252 bootstrap replicates using the CAT model of substitution rate heterogeneity, which approximates
253 the GAMMA model while offering substantial increases in computational speed. The ML tree
254 was then estimated under the full GTR+GAMMA model to provide the most accurate estimate of
255 the ingroup phylogeny.

256

257 Networks of Spanish cave bear and brown bear haplotypes were then generated using the median-
258 joining algorithm implemented in the program NETWORK (fluxus-engineering.com, Bandelt *et*
259 *al.* 1999). To avoid any confounding effects of missing data on haplotype identification, all
260 alignment columns containing missing data and/or alignment gaps were removed for network
261 analysis.

262

263 We then investigated the strength of association of mitochondrial lineage and cave using trait-
264 phylogeny association tests that account for phylogenetic uncertainty in the software BaTS
265 (Parker *et al.* 2008). If mitochondrial phylogeny and cave are strongly associated, then the
266 inferred number of changes in cave occupation across the phylogeny should be fewer than for a
267 random prediction with no such association. We generated a Bayesian posterior sample of trees in
268 BEAST v. 1.8.2 (Drummond *et al.* 2012), and then randomised the assignment of individuals to
269 caves in order to generate a null distribution of the number of changes in cave occupancy when
270 phylogeny and cave show no association. This strength of association was then tested by
271 comparing this null distribution to the observed number of changes occurring across the posterior
272 sample of trees using the parsimony score (PS) statistic (Slatkin & Maddison 1989). PS is a
273 discrete metric and therefore models changes in cave occupation occurring across the phylogeny
274 as discrete events.

275

276 To generate the posterior sample of trees used in trait-phylogeny association tests, the program
277 PartitionFinder (Lanfear *et al.* 2012) was first used to select appropriate partitions and
278 substitution models within each alignment (details in Section 2 of the Supporting Information,
279 results in Tables S5 & S6, Supporting Information). BEAST analyses involved a coalescent
280 Bayesian Skyline population model with unlinked substitution and strict clock models for each
281 partition. Non-zero variation in substitution rates was rejected by preliminary runs using relaxed
282 clock models. No clock calibrations were applied, and instead the substitution rate of the fastest-
283 evolving partition was fixed to 1 and substitution rates for the remaining partitions estimated
284 relative to the latter partition within open uniform priors between 0–2. MCMC chains ran for

285 sufficient length to achieve convergence and sufficient sampling of all parameters (ESS > 200)
286 after removal of burn-in, as verified in the program TRACER (Rambaut *et al.* 2014).
287 LOGCOMBINER was used to remove pre-burn-in trees prior to trait-phylogeny association tests.

288

289 ***Dating of cave lineages***

290 Thirty-nine samples were directly ¹⁴C dated and 2-sigma calibrated using OxCal 4.2 online
291 (accession date: 07/07/2015), based on the IntCal-13 curve (Reimer *et al.* 2013). For samples that
292 lacked ¹⁴C dates, or were beyond the range of ¹⁴C dating, we estimated their ages using a
293 Bayesian phylogenetic approach in BEAST (Shapiro *et al.* 2011). Phylogenetic age estimation
294 was conducted individually for each undated cave bear and brown bear based on ¹⁴C dated
295 representatives of their respective clades. We additionally tested the reliability of this procedure
296 using a crossvalidation method, in which the age of each ¹⁴C dated sample was estimated and
297 compared to its original ¹⁴C age. Due to the large number of individual analyses required, a
298 custom Perl script was used to automate the generation of BEAST input files. In each analysis,
299 the posterior distribution of the tip date of the undated sample was sampled within an open
300 uniform prior between 0 (present day) and one million years, both of which represent implausible
301 extremes for the ages of these samples, while fixing the ages of ¹⁴C dated samples to the mean
302 calibrated date. Substitution rates for all partitions were estimated within open uniform priors
303 between 0–5x10⁻⁷ substitutions site⁻¹ year⁻¹. Other details of the BEAST analyses were as
304 described above. Finally, we generated fully sampled calibrated phylogenies of the cave bear and
305 brown bear clades by fixing tip dates to either mean calibrated ¹⁴C ages or median phylogenetic
306 age estimates.

307

308

309 **RESULTS**

310

311 ***DNA sequences***

312 PCR screening resulted in successful amplification of mitochondrial control region fragments in
313 57 out of 85 cave bear extracts and 23 out of 24 brown bear DNA extracts (details in Table S2,
314 Supporting Information), which were then subjected to hybridisation capture enrichment and
315 high-throughput sequencing. Mapping of sequence reads to their respective reference
316 mitogenome sequences resulted in consensus sequences of 26 cave bears and 15 brown bears that
317 were > 70% complete and used for further analysis (details in Table S4, Supporting Information).
318 All datasets showed molecular damage patterns characteristic of ancient DNA (Figs. S2 & S3,
319 Supporting Information). For cave bears, we added the sequence from an additional shotgun-
320 sequenced individual (Section 1, Supporting Information) and previously published sequences
321 from four other individuals from the focal caves, bringing the total number of Spanish cave bears
322 analysed to 31.

323

324 Phylogenetic analysis supported the inclusion of these Spanish cave bear and brown bear
325 sequences within the Western European *U. spelaeus* cave bear clade and the Western European
326 brown bear clade 1 (Fig. S4, Supporting Information), identified by previous phylogeographic
327 studies (Hirata *et al.* 2013; Stiller *et al.* 2014). Spanish cave bear and brown bear haplotypes were
328 unique compared to all previously published haplotypes of conspecific bears occurring elsewhere

329 in their respective distributions.

330

331 ***Association of mitochondrial DNA and cave***

332 Network analysis of Spanish cave bear haplotypes revealed close relationships between

333 haplotypes found within the same cave (Fig. 2a). Most caves contain multiple unique haplotypes

334 that are separated from each other by single nucleotide mutations. For example, Eirós and

335 Amutxate caves each contain two unique haplotypes differing from one another by a single

336 nucleotide mutation. Similarly, five unique and closely related haplotypes were found in A Ceza

337 cave, but with the addition of a more divergent haplotype found in a single A Ceza individual

338 (sample C7) that is shared with individuals from Arcoia and Liñares. An additional unique

339 haplotype was found in Liñares cave that differs from this shared haplotype by a single nucleotide

340 mutation. Even considering the occurrence of a single haplotype that is shared among three caves,

341 an overall pattern of separation of haplotype clusters into caves is clear and obvious. Trait-

342 phylogeny association tests further confirmed this pattern, showing fewer observed changes in

343 cave occupation than expected by random (observed mean 5.9, null mean 18.0, $p < 0.001$),

344 indicating a strong association of Spanish cave bear mitochondrial lineages with particular caves.

345

346 In contrast, an obvious segregation of mitochondrial haplotypes among different caves was not

347 observed in middle Holocene Spanish brown bears (Fig. 2b). Haplotypes are widely shared

348 among caves, with the exception of Pena Paleira, which contains three unique haplotypes, but

349 these are not closely related. Trait-phylogeny association tests found the observed number of

350 changes in cave occupation to not differ significantly from random (observed mean 6.5, null

351 mean 8.2, $p = 0.08$), indicating a lack of statistically significant association between
352 mitochondrial lineage and cave in these middle Holocene Spanish brown bears.

353

354 The association of mitochondrial haplotype lineage and cave revealed by network analysis for
355 Iberian cave bears, but not for Iberian Holocene brown bears, is also evident from the time-
356 calibrated phylogenies of their respective clades (Figs. 3 & 4). In addition, the broader geographic
357 sampling of cave bear haplotypes in this analysis reveals that Spanish haplotypes as a whole are
358 not monophyletic, with some cave lineages sharing more recent common ancestry with haplotypes
359 found in France and/or Germany.

360

361 ***Dating***

362 ^{14}C ages spanned a range of $> 40,000$ to 28,251 yBP for cave bears and 41,201 to 2,520 yBP for
363 brown bears (Table S3, Supporting Information).

364

365 Crossvalidation testing of the phylogenetic age estimation procedure resulted in 95% highest
366 posterior densities (HPDs) that included the actual ^{14}C age for all brown bears and all but one
367 cave bear. Median estimated ages were also very close to the known age in most cases (Figs. S5
368 & S6, Supporting Information). These results support the reliability of this approach in estimating
369 the ages of samples without ^{14}C dates. Furthermore, age estimation for undated samples produced
370 unimodal posterior estimates that are consistent with other sources of age information, where
371 available, such as samples that were outside the range of ^{14}C dating and those dated by amino
372 acid racemisation (Table S7, Supporting Information).

373

374 Age estimates for cave bears (Fig. 5a) are compatible with the contemporaneous existence of the
375 A Ceza, Amutxate, Arcoia and Liñares mitochondrial lineages. Although phylogenetic age
376 estimates are associated with substantial uncertainty, the 95% HPDs of age estimates for these
377 four caves show considerable overlap and median estimated ages are broadly comparable with
378 each other, and with ^{14}C dated samples. The simultaneous occupation of these caves is also
379 supported by ^{14}C dating of other specimens not included in this study (Pérez-Rama *et al.* 2011).
380 In contrast to these caves, the Eiros mitochondrial lineage appears to have existed more recently
381 and potentially without temporal overlap with those from other caves, although we do find slight
382 overlap of Eiros ^{14}C dates and HPDs from other caves in some cases (Fig. 5b). Generally younger
383 ^{14}C dates of Eirós in comparison to the other caves have also been reported previously, however,
384 a single specimen was dated to more than 40,000 yBP (Pérez-Rama *et al.* 2011), and may
385 therefore have existed contemporaneously with individuals from other caves. Unfortunately, this
386 sample failed to yield any usable DNA and so its phylogenetic relation to more recent Eirós cave
387 bears remains unknown. Caves containing brown bear remains were almost certainly inhabited
388 simultaneously. ^{14}C ages and a single phylogenetic estimate indicate temporal overlap in the
389 habitation of these five caves between approximately 10,000 and 6,500 yBP (Fig. 5b).

390

391

392 **DISCUSSION**

393

394 *Evidence for homing behaviour*

395 Cave bears and brown bears that died in caves in the north of Spain show remarkably contrasting
396 patterns of mitochondrial haplotype segregation. While no significant association of
397 mitochondrial haplotypes and cave is found in middle Holocene brown bears, in the case of Late
398 Pleistocene cave bears each cave contains, almost exclusively, a unique clade of closely related
399 haplotypes. This structure exists despite caves being located in close geographic proximity and
400 being inhabited simultaneously. We therefore interpret this as evidence of homing behaviour in
401 cave bears. This scenario would involve a single intermixing cave bear population within which
402 individuals – both males and females – returned to their native caves annually for hibernation,
403 that is, the cave in which their mother hibernated and also gave birth, as demonstrated by the
404 large amounts of perinatal individuals in the sites (Torres *et al.* 2002; Pérez-Rama *et al.* 2011).
405 Such homing behaviour does not exclude mating between bears from different caves, but would
406 have sorted the mitochondrial lineages by caves. In contrast, the lack of association between
407 mitochondrial haplotype and cave in middle Holocene brown bears rejects this type of homing
408 behaviour in this closely related species. This is further supported by studies of extant brown bear
409 populations which show greater flexibility with regard to hibernation site than inferred here for
410 cave bears (e.g. in Naves & Palomero 1993).

411

412 Evidence suggests that cave bears hibernated communally (e.g. Philippe & Fosse 2003). Homing
413 behaviour would therefore result in non-random groups of close maternal relatives assembled at
414 each cave. Thus, this behaviour can be further considered as a form of sociality. The temporal
415 stability of these social groups is demonstrated by the observation of multiple unique haplotypes
416 within caves that differ from their nearest relative by a single nucleotide substitution (Fig. 2).

417 This suggests that within-cave haplotype variability is the result of nucleotide mutations that
418 occurred during the period of cave occupation, most likely over thousands of years. A stepwise
419 pattern of haplotype variability within caves has previously been reported for short cave bear
420 control region sequences from the Ach valley, south-western Germany (Hofreiter *et al.* 2007),
421 which in light of our finding suggests the potential for similar homing behaviour in that
422 population. The temporal stability of cave occupation by cave bears is further demonstrated by
423 two morphologically distinct cave bear forms that each occupied separate caves located only a
424 few kilometers apart in Austria. These morphotypes sort into respective, genetically divergent
425 mitochondrial clades. Despite their close proximity, a previous study found no evidence of
426 haplotype exchange between caves even though simultaneous occupation over thousands of years,
427 implying both site fidelity and reproductive isolation (Hofreiter *et al.* 2004). In the case of
428 Spanish cave bears, however, we consider reproductive isolation unlikely due to a lack of any
429 obvious morphological separation and relatively low levels of haplotype divergence between
430 caves. Our preferred alternative, a single population with homing behaviour, makes specific
431 predictions about patterns of nuclear autosomal and sex-chromosome divergence among caves,
432 and obtaining such data would be a valuable direction for future cave bear research.

433

434 Although we found a clear association of mitochondrial lineage and cave in Spanish cave bears,
435 the association is not perfect. Specifically, we found a single haplotype that is shared among three
436 caves: Liñares, A Ceza and Arcoia. This shared haplotype is common among Liñares individuals,
437 and separated from a second Liñares haplotype by a single nucleotide mutation. In the second
438 cave, A Ceza, the shared haplotype is considerably diverged from other haplotypes within that

439 cave. In the third cave, Arcoia, both samples investigated have the shared haplotype. These later
440 samples are the remains of juvenile individuals and no other cave bear remains have been found
441 in this cave, raising the possibility that these juveniles (and potentially the A Ceza individual
442 carrying the same haplotype) originate from Liñares. Regardless of the origin of this shared
443 haplotype, while this pattern does imply some degree of movement between caves, the overall
444 evidence for homing behaviour is clear and substantial. An ability to disperse and occupy other
445 caves is further indicated by the sister group relationship found between Eirós cave haplotypes
446 and a haplotype from Chauvet cave in France, two caves that were occupied simultaneously (see
447 Table S3, Supporting Information; Bon *et al.* 2008; 2011). Thus, the Eirós haplotype lineage may
448 be the result of long distance dispersal by female bears from distant caves, rather than movement
449 among localised Spanish caves, which is also consistent with the apparent temporal separation of
450 this lineage from the other Spanish caves.

451

452 ***Wider implications***

453 Homing behaviour has wider implications for species survival and conservation. For example, in
454 extant black bears (*Ursus americanus*), it has been discussed as a potential problem for
455 repopulation programs, as both females and males are able to track back to their home area after
456 being captured by humans and released several kilometres away (Beeman & Pelton, 1976; Rogers
457 & Lynn 1986; Clark *et al.* 2002). The same effect has been observed in Asian black bears (*Ursus*
458 *thibetanus*), where genetic studies showed that 63% of the translocated bears migrate back to
459 their original sites (Mukesh *et al.* 2015). Other well known examples include anadromous fishes,
460 whose ability to return to breeding sites is affected by anthropogenic disruption of freshwater

461 river systems (e.g. Pess *et al.* 2014), and similarly in marine turtles, where anthropogenic coastal
462 development threatens habitats used for egg deposition (e.g. Wallace *et al.* 2011). Although
463 ancient DNA provides the potential to investigate such behavioural patterns in species that have
464 already gone extinct, behavioural inferences based on ancient DNA have been rare (notable
465 examples are Huynen *et al.* 2010; and Allentoft *et al.* 2015). Our study clearly demonstrates the
466 potential utility of ancient DNA in the study of behavioural ecology by revealing evidence of
467 homing behaviour in extinct cave bears, and furthermore, through comparison with a closely
468 related extant species, we have also uncovered clues on the potential causes of cave bear
469 extinction.

470

471 The role of humans in the extinction of the cave bear has been debated (Grayson & Delpech 2003;
472 Munzel & Conrad 2004; Knapp *et al.* 2009; Bon *et al.* 2011; Stiller *et al.* 2014), but explanations
473 that also account for the survival of the sympatric brown bear have remained elusive. It is likely
474 that the high dependence of cave bears on their native caves would have made them more
475 sensitive to human competition for caves for several reasons. First, as noted previously (Grayson
476 *et al.* 2003; Stiller *et al.* 2010), the generally high dependence of cave bears on caves for
477 hibernation would have brought them into severe competition with humans (both Neanderthals
478 and modern humans). Second, their tendency to come back to the same cave site would have
479 made them comparatively predictable prey, which fits to the growing evidence of cave bear
480 hunting, again by both Neanderthals and modern humans (Munzel & Conrad 2004; Wojtal *et al.*
481 2015). And third, this homing behaviour would have prevented a rapid recolonisation of empty
482 caves from neighbouring populations. Overall, these factors could have contributed to the

483 extinction of the cave bear as modern human populations expanded from Eastern to Western
484 Europe, indeed, advancing in the same direction as the subsequent cave bear extinction. This is in
485 agreement with recent studies that have questioned the relative contribution of Pleistocene
486 climatic changes to cave bear extinction, and suggested instead a major impact of human
487 activities (Knapp *et al.* 2009; Bon *et al.* 2011; Stiller *et al.* 2014). Finally, the lack of evidence of
488 homing behaviour to their maternal caves in Spanish brown bears, a species that lived in
489 widespread sympatry with cave bears but survived the human expansion into Western Europe,
490 further implicates this behaviour as a factor in the extinction of the cave bear.

491

492

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502

503

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671
672 **DATA ACCESSIBILITY:**

673 DNA sequences from the cave and brown bears obtained in this study are deposited in Genbank
674 with accession numbers: KX641289-KX641337. DNA sequence alignment has been deposited in

675 Dryad with the identifier doi:10.5061/dryad.cj965.

676

677 **AUTHORS CONTRIBUTIONS:**

678 G.G.F, A.B, A.G and M.H designed and conceived of the study; G.G.F, A.B and I.N.M.
679 performed molecular work; G.G.F, A.B, B.K and D.F, performed NGS data processing and
680 statistical analysis; A.G, A.G.V, A.C.P, S.C, T.J.T, J.E.O, C.F and G.R collected and identified the
681 ancient remains. G.G.F, A.B, and M.H drafted the manuscript with input from A.G and A.G.V. All
682 authors gave final approval for publication.

683

684

685 **FIGURE LEGENDS**

686

687 **Figure 1.** Map of Northern Spain showing locations of the caves investigated in this study.
688 Circles represent sites with cave bears. Squares are sites with brown bears. Colours are consistent
689 with Fig. 2.

690

691 **Figure 2.** Haplotype networks of A. Iberian cave bears and B. Iberian brown bears, coloured
692 according to the cave in which that haplotype was found (indicated next to each network). Circles
693 are sized relative to haplotype frequency. Dashes along edges indicate single nucleotide mutations.

694

695 **Figure 3.** Time calibrated phylogeny of the Western European *U. spelaeus* cave bear clade. The
696 lower scale shows kyBP. Branch labels indicate posterior clade probabilities ≥ 0.95 , except for
697 terminal tip clades where labels have been removed for simplicity. Nodes are centered on the
698 median estimated divergence time and bars show the 95% HPD. Circles next to taxon names
699 indicate Iberian cave bears and are coloured according to cave (consistent with Fig. 2). The *U.*
700 *ingressus* clade that is sister to the *U. spelaeus* clade and was utilised for molecular dating is
701 shown collapsed for simplicity.

702

703 **Figure 4.** Time calibrated phylogeny of the Western European brown bear clade. The lower scale
704 shows kyBP. Branch labels indicate posterior clade probabilities ≥ 0.95 . Circles next to taxon
705 names indicate Iberian brown bears and are coloured according to cave (consistent with Fig. 2).
706 Two additional representatives of the West European brown bear clade, from Austria (sample Uap)
707 and Bulgaria (GenBank Accession AP012591), were analysed and found to form a well supported
708 sister lineage to the clade shown here that diverged an estimated 68,401 yBP ago (95% HPD
709 50,409–92,631 yBP). This lineage is not shown in order to better visualise divergence times
710 among Iberian brown bear haplotypes.

711

712 **Figure 5.** Time lines of A. Iberian cave bear and B. Iberian brown bear sample ages. Time in yBP
713 is shown on the Y axes. Each point indicates the estimated age of an individual bear. Black points
714 are median phylogenetic age estimates and red points are mean calibrated ^{14}C ages. Error bars
715 show 95% HPD and calibrated ^{14}C uncertainty for phylogenetic age estimates and ^{14}C ages,
716 respectively.