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Evolutionary Computation Systems for Biological Data Analysis Protein Family Class Prediction with Particle Swarm Optimization

Dissertation presented to the University of Coimbra in fulfilment of the requirements to obtain a Master's degree in Biomedical Engineering

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FCTUC FACULDADE DE CIÊNCIAS E TECNOLOGIA UNIVERSIDADE DE COIMBRA

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EVOLUTIONARY COMPUTATION SYSTEMS FOR BIOLOGICAL DATA ANALYSIS

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Abstract

Proteins are macromolecules formed by amino acids and they have several biochemical functions. Consequently, they are present in the most important reactions at the cellular level. So, by knowing the principal functions of a new protein sequence is possible to control several metabolic pathways or even certain reactions to a number of stimuli, and so it is possible to gain access to different therapeutic procedures. Therefore, it is essential the creation of computer tools capable of identifying the biochemical functions and applicability of known proteins.

In this thesis, we propose a new method for extraction and selection of features from proteins' primary structure in order to improve efficiency in the prediction of their family classes. The feature extraction consists in a meticulous and differentiated analysis of all possible amino acid subsequences that could be present in the chosen proteins. For each subsequence is assigned a set of different values with statistical significance according its length, high or low frequency, and also the order that amino acids are present consecutively or not in the protein (Scoring). The feature selection is the collection of subsequences that have the highest values in each of the previously highlighted parameters (MaxScoring) in order to initialize the particles in Particle Swarm Optimization.

Two sets of proteins were selected with different characteristics, wherein was possible to prove that our methodology can improve the proteins' class family classification based on SVM when compared with other more common methods. The best results have an average AUC value above 0.80 and it is observed a 10% improvement compared to Amino Acid Composition (AAC),

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and a 20% improvement when compared to the Pseudo Amino Acid Composition Amino Acid (Pse-AAC) and Amphiphilic Pseudo Amino Acid Composition (Am-Pse-AAC).

Resumo

As proteínas são macromoléculas formadas por aminoácidos e possuem diversas funções bioquímicas. Consequentemente, estão presentes nas mais importantes reações metabólicas a nível celular. Assim sendo ao saber quais as principais funções de novas sequencias proteicas é possível controlar diversas vias metabólicas ou ainda reações especificas a certos estímulos, e assim é possível ter acesso a diferentes procedimentos com fins terapêuticos. Logo é essencial a criação de ferramentas computacionais capazes de identificar a funções bioquímicas e a aplicabilidade de proteínas conhecidas.

Nesta tese é proposto um método de extração e seleção de atributos a partir da estrutura primária de proteínas de forma a aumentar a eficiência de previsão das suas classes de famílias. A extração de atributos consiste numa meticulosa e diferenciada análise de todas as possíveis subsequências de aminoácidos que podem estar presentes nas proteínas selecionadas. Para cada subsequência é atribuída um conjunto de diferentes valores com significância estatística conforme o seu comprimento, alta ou baixa frequência, e ainda a ordem em que os aminoácidos estão presentes de forma consecutiva ou não na proteína (Scoring). Já a seleção constitui na escolha das subsequências que apresentem os maiores valores em cada um dos parâmetros salientados anteriormente (MaxScoring) de forma a inicializar as partículas no Particle Swarm Optimization.

Foram escolhidos dois conjuntos de proteínas, com características diferentes, sendo que em ambos foi possível provar que a nova metodologia consegue melhorar a classificação das classes de famílias de proteínas baseada em SVM quando comparada com outros métodos mais comuns. Os melhores resultados obtidos apresentam um valor médio de AUC acima de 0.80 e, nos dois conjuntos é observada uma melhoria acima de 10 % em relação ao de Amino Acid Composition (AAC) e de 20% quando comparado com Pseudo Amino Acid Composition (Pse-AAC) e Amphiphilic Pseudo Amino Acid Composition (Am-Pse-AAC).

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List of Symbols

| a_m | Native amino acids |
|-----------------------------------|--|
| a_m^n | m amino acid in n position |
| $\boldsymbol{b}_{\boldsymbol{m}}$ | Amino Acid m of the set with all coding possibilities of a sequence |
| b_m^n | Amino Acid m of the set with all coding possibilities of a sequence at n |
| | position in a sequence |
| fc | Frequency of consecutives elements |
| fnc | Frequency of non-consecutive elements |
| $f_i(P_j)$ | Frequency of Amino Acid i in Protein j |
| f _u | Normalized frequency of the 20 amino acid in a certain protein |
| | sequence |
| H ¹ | Hydrophobicity correlation function |
| h^1 | Amino acid hydrophobicity value |
| H_1^0 | Amino acid hydrophobicity value |
| $H_1(i)$ | Amino acid i hydrophobicity value after standardization |
| H ² | Hydrophilicity correlation function |
| h^2 | Amino acid hydrophilicity value |
| H_2^0 | Amino acid hydrophilicity value |
| $H_2(i)$ | Amino acid i hydrophilicity value after standardization |
| L | Sequence Length |
| λ | Total Number of order correlation factor |
| M ⁰ | Mass of an amino acid |

| M(i) | Mass of i amino acid after standardization |
|------------------------|---|
| Ν | Total Number of samples in a set of proteins |
| $P_j(n)$ | Protein j |
| s_c^2 | Variance of subsequence FC |
| S_{ki} | Subsequence i with k amino acids in $V(i)$ |
| s_{nc}^2 | Variance of subsequence FNC |
| $oldsymbol{	heta}_j$ | j-tier sequence correlation |
| V(i) | All subsequences possible |
| W | Weight factor for Pse-AAC and Am-Pse-AAC |
| $\overrightarrow{x_J}$ | AAC vector for Protein j |
| x_u | Pse-AAC and Am-Pse-AAC vector for the representation of a certain |
| | Protein |

List of Abbreviations

| AAC | Amino Acid Composition |
|------------|---|
| AFP | Absolute Frequency by Presence |
| AFPC | Absolute Frequency by Presence of Elements Arranged Consecutively |
| AFPN | Absolute Frequency by Presence of Elements Arranged Non |
| | Consecutively |
| Am-Pse-AAC | Amphiphilic Pseudo Amino Acid Composition |
| ASF | Absolute Set Frequency |
| ASFC | Absolute Set Frequency of Elements Arranged Consecutively |
| ASFN | Absolute Set Frequency of Elements Arranged Non Consecutively |
| AUC | Area Under Curve |
| FC | Frequency of Elements Arranged Consecutively |
| FNC | Frequency of elements arranged non consecutively |
| FS | Fisher Score |
| FSC | Fisher Score of elements arranged consequently |
| FSN | Fisher Score of elements arranged non consequently |
| KF | k-Frequency |
| KFC | K-Frequency of elements arranged consequently |
| KFN | K-Frequency of elements arranged non consequently |
| MS | MaxScoring |
| MS - N | MaxScoring Normal |

| MS – WPS | MaxScoring Without Parent Subsequences |
|----------|---|
| Pse-AAC | Pseudo Amino Acid Composition |
| PSO | Particle Swarm Optimization |
| PSO-F | Particle Swarm Optimization Features |
| PSO-W | Particle Swarm Optimization Weights |
| PSO-WF | Particle Swarm Optimization Weights and Features |
| RFP | Relative Frequency by Presence |
| RFPC | Relative Frequency by Presence of Elements Arranged Consecutively |
| RFPN | Relative Frequency by Presence of Elements Arranged Non |
| | Consecutively |
| SVM | Support Vector Machine |
| TF-idf | Term frequency-inverse document frequency |
| TFC-idf | Term frequency-inverse document frequency of Elements Arranged |
| | Consecutively |
| TFN-idf | Term frequency-inverse document frequency of Elements Arranged |
| | Non Consecutively |
| XS | Qui quadrado statistics |

1 Introduction

In this chapter, it is described the motivation, main objectives and the key concepts about proteins and their importance. It will be referred only the most important aspects that can influence the biochemical functions in a protein, like the presence of domains and motifs as well their structure. Also, it is presented a brief overview of the chapters in a comparative scheme.

1.1 | Motivation

The technological advances in the genomic and proteomic sequencing, headed to a large increase of available data whose analysis may leads to discoveries in some areas, such as pharmaceuticals or medicine. This rises the need to create computational tools able to interpret different biological data, in this case proteins. Proteins are present in all cellular metabolic reactions by performing a large number of several functions, so it's important to identify their biological purpose based on the data available in order to not require additional laboratory work in researches with the study of proteins involved.

1.2 | Objectives

The main objective of this thesis is the design of a new method capable of predicting if a protein belongs or not to a class or subclass of proteins, as well as being capable of it effective regardless of the data set supplied. Therefore, all the research is the definition of a possible solution to a problem: the prediction of a protein's biochemical function through its primary structure.

1.3 | Proteins

Proteins are macromolecules formed by amino acids joined by peptides bonds. The sequence of amino acids is determined by the sequences of nucleotide on a process called translation (Campbell, 1996) after a gene as coded to pre-mRNA (Transcription). The sequence of amino acid only is the primary structure of a protein.

The following sections will explain some of the elements that can determine or influence the proteins' biochemical functions as well as the functions themselves.

1.3.1 | Structures

As it is shown in figure 1, a protein can assume four different structures: Primary, secondary, tertiary and quaternary. The first one is the sequence of amino acids and second is the polypeptide chain taking the form of alpha helices or beta strands because of hydrogen bonds processed between specific molecular groups of certain amino acids. In the tertiary structure elements of either alpha helix or beta sheet or both, as well loops and links with no secondary structures are folded into a globular form. At last, the quaternary structure is the association of folded chains with more than one polypeptide.

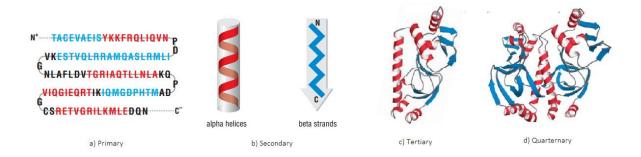


Figure 1 – The different structures which a protein can assume illustrated by the catabolite activator protein (Petsko & Ringe, 2004)

The structure of a protein is important because in order for a polypeptide to function as a protein in most cases must have a stable tertiary structure under physiological conditions. Regarding the amino acids, they have different side chains which allows them to interact differently with each other and with water. These differences affect their contributions to the protein's stability as well to the protein's function. So, the amino acids are categorized as the side chains present in them: hydrophobic, hydrophilic and amphipathic amino acids residues.

Also, certain amino acids are easier found or present in a greater amount in some structures than others. It is the case of leucine and glutamine, which are often found in helices in contrast to valine or isoleucine that are more often in beta sheets. But there also exceptions, like the proline presence does not favor any kind of secondary structure. This is because each side chains of each amino acid are essential for the chemical reactions needed for a protein to assume any kind of form. As for the tertiary structure, most proteins can be unfolded or refolded according to the diluted solution that they are in, indicating that the primary structure contains all the information needed to specify the folded state.

In addition, the water molecules present on the surface of folded protein and the possible bonds created also determines how a protein can be folded, since as referred before, there are different types of amino acids and therefore different kinds of interactions between amino acids and water. So, the side chains of amino acid will be arranged accordingly to the presence of water molecules, the hydrophilic side chains will be arranged to be closer to water molecules and the hydrophobic side chains will settle in the opposite way.

1.3.2 | Protein Domain

A domain is a specific region of a protein structure usually composed by a subsequence of amino acids and is capable of existing on its own in aqueous solution. This segment holds part of the biochemical function of the protein they belong to.

Not all the known domains are continuous segments of amino acids, in many proteins a domain is divided and dispersed across the sequence and also they vary in size, being able to contain an average of 200 amino acids, in which the smallest domain was registered with 57 amino acids and the biggest with 907 residues.

1 Introduction

1.3.3 | Motifs

A motif can be a particular amino acids sequence that is present for a precise biological function, or can be a set of contiguous secondary structure elements that either retain a specific functional importance of a portion of a folded domain. In this sense, it can exist functional motifs (like the ones found in DNA-binding proteins), or simpler structural motifs. This last one does not exist separately from a protein and it can be used as a recognition element for a group of similar proteins. A lot of structural motifs are found in many hormones as well as proteins with NAD cofactors.

The recognition of a motif in a sequence is not easy since most of sequence motifs are discontinuous and the space between their elements can vary significantly and this makes them easier to detected from a structure view rather than from the amino acids sequence. As for the structural motifs, its identification only from the sequence is very difficult given that many different amino acids can lead to same secondary structure making algorithms based in similarity alone not enough.

1.3.4 | Biochemical Functions

The different protein biochemical functions can be categorized into four main functions: binding, catalysis, switching and as structural elements. In binding, proteins bind to a specific or more substrates like Myoglobin binds a molecule of oxygen reversibly to the iron atom in its heme. As for the second category, the proteins are responsible to increase the velocity of every chemical reaction in a living cell. Switching proteins are flexible molecules and their conformation change with ph or a ligand binding, allowing some cellular processes to be controlled. These conformational changes are crucial for the molecular basis of many cancers like the ones that occurs in the GTPase Ras¹ when GTP² is hydrolyzed to GDP³. At last, structural proteins give strength or toughness to living systems, and they depend on specific protein subunits with other proteins or molecules.

¹ Protein from a large family of hydrolase enzymes that can bind and hydrolyze guanosine triphosphate, this particular protein is a member of the Ras superfamily of proteins

² Guanosine triphosphate

³ Guanosine diphosphate

1.4 | Thesis Structure

This thesis has six distinctive chapters. In chapter 2 a synopsis of related works is presented. In chapter 3 all the implemented methods, as well the new method, are explained. In chapter 4, it is presented an application and a website created to implement the different methods described before. In chapter 5, it is introduced the different results as they are discussed in chapter 6. At last, the final remarks, conclusions and suggestions for future work are present in chapter 7.

1 | INTRODUCTION

2 | Literature Review

In this chapter, it will be referenced and discussed the several researchs carried out using the primary sequence of a protein to create models that can predict their biochemical functions.

2.1 | Chapter Introduction

There are several factors that can determine or influence the action of a protein in a cell, from its sequence, structure, cellular location, and or even to the presence of other proteins. Thus, several studies address the same problem in different ways: either by predicting subcellular location of a group of proteins or the enzymes subclasses.

So this subject can be divided into three sub problems: protein representation or feature extraction, feature selection and the prediction algorithms used.

2.2 | Features Extraction

The representation of a protein sequence is one of the most important tasks as it may determine the success of any model. It is important that a protein is mathematically well represented in order to have the greatest number of critical information which can characterize it between a set of proteins that share the same function. However, we only have access to a sequence of amino acids, so it is necessary to implement methods to extract features from it. For this purpose, there are also different approaches: methods based on AAC, subsequences, and N-terminal targeting. One of the most basic ways to represent a protein sequence is by counting the 20 amino acids in the sequence: AAC (Nishikawa, et al., 1983). Despite being a fairly simple and old method, it is still used quite successfully in several sets of proteins to predict their subcellular location or subfamily (Zhou & Doctor, 2003), or as a way to establish a performance threshold when proposed a new method with a new set of proteins (Yang, 2011). However, the simplicity of AAC means that many important factors for a biochemical function are not represented.

Pse-AAC technique was developed by Kuo Chen Chou (Chou, 2001) and this method brought something new and unique to the approaches previously used (variations of AAC) to predict cell attributes. Chou showed a new representation of protein sequences, in which was still present in the frequency of the 20 amino acids, as well as their mass, hydrophobicity and hydrophilicity according to the order they appear in the same sequence. Thus, not could only a new interpretation for the first time took account of some chemical properties as well as the order in which the amino acids appeared in the sequence. In addition, using this method a protein sequence is analyzed as set of amino acid pairs (figure 2), because the terms determined to implement this method always needs the current and the immediately following amino acid. In his first work, Chou had used the Pse-AAC to predict cell locations, however this method of feature extraction has been used for other purposes such as prediction of protein structures (Sahu & Panda, 2010), potentially allergenic proteins (Mohabatkar et al., 2013), families of human enzymes (Wu, et al., 2016), among many other studies in various topics.

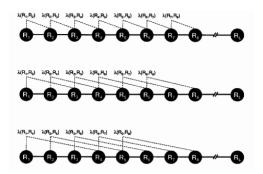


Figure 2 – Schematic Representation of the first three tier sequence order correlation mode along a protein sequence on a Pse-AAC algorithm.

The Pse-AAC was also used in predicting 16 different oxygenase classes according to different target groups (Elrod & Chou, 2003). This work was important in the sense that it determined the homologies between the proteins of each class in order to understand how this

factor could be decisive when using methods based on the AAC. And it proved that despite low homology (30%) between proteins in the same class, with Pse-AAC was able to obtain an accuracy values above 70%, demonstrating that with only the primary sequence is possible to identify and predict a set of proteins with different sequences, whether by class families or their subcellular location.

After the initial proposal of Chou, different variations of his work appeared in order to complete the representation with more relevant biological information, it was the case of isno-PseAAC (Xu et al., 2013), a new approach to Pse-AAC, allowing the incorporation of a factor called the propensity of the position of a specific amino acid to predict Cysteine S-nitrosylation Sites in Proteins. Basically, it is a 21x20 matrix in which each row and column represent the 20 amino acids and the possibility of encoding not one, i.e. the existence of an element x in an amino acid sequence. Thus each element of the array can be translated in the interaction of two consecutive amino acids in the sequence, where is calculated the difference between the terms Pse-AAC for a pair of amino acids between the data set that contains peptide fragments and those that don't.

Besides Pse-AAC, Chou also proposed Am-Pse-AAC to predict enzyme subclasses, in which the molecular weight is not considered and hydrophobicity and hydrophilicity are considered differently in the correlation coefficients, more detail in next chapter (Chou, 2004). The Am-Pse-AAC was also incorporated in different works related to the study of iterations between proteins (Huang, et al., 2014).

It is important to add that all these methods were used for feature extraction, due to the fact that is only based on an amino acid sequence, the same methods were transposed of different biological data, like for RNA sequences (Liu, et al. 2015), and DNA small sequences (Liu et al., 2015).

As for Pse-AAC, it proved to be an effective and simple method to represent a protein leading to the existence of many diverse research, as mentioned earlier. However, it cannot be ignored some aspects that can make this an inadequate protein representation.

Firstly, the protein is observed as a set of pairs, so it is not taken in consideration the existence of protein domains, subsequences crucial to its structure as well as biochemical functionality. Another aspect, it is related to the use of hydrophilicity and hydrophobicity,

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important parameters for intra and inter molecular interactions in the various activities of a protein, as another identification to an amino acid. But instead of having the designation of the single letter amino acid, it has a term that considerers its mass, hydrophilicity and hydrophobicity. Lastly, the protein is not fully analyzed, despite being present the total frequency of each amino acid sequence, the order is considered to a certain amino acid in the sequence, so possible vital information is not considered.

In addition to the above mentioned methods there is the analysis of amino acids sequences present in the protein primary structure through its presence and / or its frequency. Such subsequences can be different combinations of amino acids set (k-tuples), motifs sequences or N-terminal targeting sequences (small peptides present in the protein responsible for the migration of the protein into all specific organelle, such a mobile locator. (Hoglund et al., 2005) (A, et al., 2006).

The biggest problem in using motifs sequences or N-terminal targeting sequences is that it is limited because it is needed a prior knowledge of the type of proteins used and the laboratorial technique performed, and also exist many cellular events that can change or mutate these subsequences.

However, the use of k-tuples is positive in the sense that for the chosen data set is done a thorough search for subsequences that may be potential motifs, targeting sequences or domains. Though, given the existing amino acids and the number of all possible combinations, the number of the all possible subsequences leads to a very high number of potential attributes that can serve as feature to train a classifier. Not to mention that assessment of these same subsequences only by the presence or frequency continues to be insufficient in relation to the biological and chemical knowledge of the subject. It is not only the presence or frequency of a pattern that influence the structure and thereby the biochemical functionality of a protein. Because there are several intra and inter molecular interactions or structures of each amino acid set in the same sequence that can determine different biological purposes for a certain protein.

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2.3 | Feature Selection

After the features extraction is set, how those features are going to be selected is also important, since any efficient prediction model depends on how well described is our study object.

And for proteins, it is even more difficult to choose a correct way of features selection because of the enormous number of biological variables that may contribute to its classification presence in a particular cellular location, or as belonging to a group of proteins that share the same chemical or functional characteristic.

In several researches, there are different methods for features selection, in which some used using the analysis of subsequences of amino acids (k-tuples). There are studies that select k-tuple more frequently, or with higher discriminative power or less independent, by calculating its frequency, Fisher's criterion, the quadratic chi respectively (Yang et al., 2006) (Yiming Yang, 1997). After, the attributes of each metric were evaluated separately in order to realize which is most efficient in the feature selection.

Another feature selection process is by homology analysis of these k-tuples with the proteins belonging to a particular class. In this process are chosen the subsequences that present the highest similarity with the referred proteins (Tian & Skolnick, 2003).

It is also important to note that many studies already used variations of the Particle Swarm Optimization (Kennedy & Eberhart, 1995) to select a set of attributes according to a chosen parameter as a fitness function, like with Chieh-Yuan's work (Tsaia & Chena, 2015), in which the PSO was used to choose a set of k-tuples using as fitness function their similarity with positive proteins (proteins that belong to a class).

In others researches the PSO is used to find the attributes that lead to better an accuracy value when applied different types of predicting algorithms, having as features the terms of the Pse-AAC (Bagyamathi & Inbarani, 2015) (Mandal et al., 2015) (Bin Liu, 2015).

2 | Literature Review

2.4 | Prediction Algorithms

The last factor that contributes to the success of a prediction model is the selection of a prediction algorithm. If is for the prediction of enzymes' subclasses or for identifying subcellular locations different methods were applied.

The nearest-neighbor method is used in a wide variety of researches to predict enzymes or to predict the types of membrane proteins (Shena & Choua, 2005).

Another prediction algorithm used is SVM, in which from protein sequences and defined attributes can determine whether a protein belongs to a class by the characterization of a hyperplane that maximizes the margin that separate two types of data, in this a case is if a sequence belongs or not to a class of proteins (Zhou et al., 2007) (Annette Hoglund, 2006).

Other methods applied were mining association and Bayesian classification rules, in which it was possible to identify different classes of enzymes with rules associated with the protein domain composition (Guda, 2011).

2.5 | Conclusion

For Feature Extraction exist effective methods from protein sequences, however there are always important chemical functionality factors of a protein being ignored or not shown. In Pse-AAC a protein is not fully analysed, while the others methods are based on similarity with known subsequences, therefore not successful for class proteins sets with low homology, mutated proteins or with lost motifs sequences. And some feature selection methods conduct an analysis based only on one factor or parameter.

2.6 | Our Proposal

Based on what it was referred before, it was created a method that from all possible combinations of variable size subsequences are pre-selected (according to more than 24 different metrics) to build a search space used in PSO to select some of them to increase the AUC (area under curve), creating a system capable of evaluating subsequences by different parameters simultaneously.

3 | Methods

In this chapter, it will be discussed in detail the methods based on AAC which have been implemented in detail, and also it will be described all the components and different variables of the proposed method.

3.1 | Chapter Introduction

It is necessary to find variables or parameters capable of identifying a certain set of proteins, in order to distinguish their different classes or subclasses to correctly identifying them, which makes it possible to establish some of the main protein functions.

The biggest obstacle lies in determining correctly these same variables solely with the protein sequence. Therefore, different methods were implemented to define diverse sets of variables.

First, each set of proteins was characterized by the number of each amino acid (AAC). Also, it was applied a method created by Kuo-Chen Chou (Pse-AAC) in which the amino acids' arrangement on the protein sequence as well as their hydrophobicity, hydrophilicity, and mass are taken in to consideration. Furthermore, all possible combinations of subsequences were determined by a predefined number of amino acids present so they could be scored according

to their occurrence and other parameters in each protein sequence (Scoring), and then some of them selected through the highest scores.

Additionally, it was implemented the PSO on all variables previously referenced in order to select the variables best capable of characterizing a certain protein set.

3.1 | Amino Acid Composition – AAC

The Amino Acid Composition is the simplest way to attempt to solve this problem and almost all the previous works made in this field are based on it, plus its results provided were used to show how well a new algorithm can improve them.

Therefore, considering a certain set with N proteins, each protein j (P_j) can be represented as an array of L-amino acids (b_m) :

$$P_{j}(n) = b_{m}^{-1} \dots b_{m}^{-n} b_{m}^{-n+1} b_{m}^{-n+2} \dots b_{m}^{-L-2} b_{m}^{-L-1} b_{m}^{-L}$$
(1)

$$L > 0, 1 < n < L, m > 0, 1 < j \le N$$

where n indicates the amino acid position in the protein sequence, and m identifies an element of b, assuming this contains all the coding possibilities, in a protein sequence, besides the 20 native amino acids because unknown or ambiguous amino acids may also exist.

For AAC, each protein j was represented by a vector $(\vec{x_j})$ with 20 elements, each corresponding to the number of occurrences of an amino acid in the same protein sequence $(f_i(P_i))$, since this amino acid was one of the 20 native amino acids.

$$\overrightarrow{x_j} = \left[f_1(P_j), f_2(P_j), \dots, f_{20}(P_j) \right]$$
(2)

As a result, to train a possible model each sample was a protein sequence represented by 20 features and labeled as 1 or 0, depending on whether they belong or not to a certain class of proteins. Table 1 – List of all elements of b. Each one has a three letter code (3LC), one letter code(1LC) and the name of its chemical structure (Chemical Compound). The first 20 amino acids (from Alanine to Valine) are the 20 native amino acids (kegg, s.d.).

| 3LC | 1LC | CHEMICAL COMPOUND | |
|-----|-----|----------------------|--|
| Ala | А | Alanine | |
| Arg | R | Arginine | |
| Asn | Ν | Asparagine | |
| Asp | D | Aspartic acid | |
| Cys | С | Cysteine | |
| Gln | Q | Glutamine | |
| Glu | E | Glutamic acid | |
| Gly | G | Glycine | |
| His | Н | Histidine | |
| lle | 1 | Isoleucine | |
| Leu | L | Leucine | |
| Lys | К | Lysine | |
| Met | Μ | Methionine | |
| Phe | F | Phenylalanine | |
| Pro | Р | Proline | |
| Ser | S | Serine | |
| Thr | Т | Threonine | |
| Trp | W | Tryptophan | |
| Tyr | Y | Tyrosine | |
| Val | V | Valine | |
| Asx | В | Asn or Asp | |
| Glx | Z | Gln or Glu | |
| Xle | J | Leu or lle | |
| Sec | U | Selenocysteine (UGA) | |
| Pyl | 0 | Pyrrolysine (UAG) | |
| Unk | Х | Unknown | |

Several simulations were made with different preprocessing methods in order to find the best way to deal with this type of data, but better results were obtained without these methods. For example, different data standardization methods were implemented like feature scaling, although it is not required in this case, since all values are in same order of magnitude.

3.2 | Pseudo Amino Acid Composition – Pse-AAC

The main purpose of this methodology is to include more significant information than the AAC by considering the physical and chemical properties of each amino acid present in a certain protein sequence.

According to this method and equation present in 1, the order-correlated factor was defined as:

$$\begin{cases} \theta_{1} = \frac{1}{L-1} \sum_{n=1}^{L-1} \Theta(P_{j}(n), P_{j}(n+1)) \\ \theta_{2} = \frac{1}{L-2} \sum_{n=1}^{L-2} \Theta(P_{j}(n), P_{j}(n+2)) \\ \theta_{3} = \frac{1}{L-3} \sum_{n=1}^{L-3} \Theta(P_{j}(n), P_{j}(n+3)) \\ & \dots \\ \theta_{\lambda} = \frac{1}{L-\lambda} \sum_{n=1}^{L-\lambda} \Theta(P_{j}(n), P_{j}(n+\lambda)) \end{cases}$$
(4)

Subsequently, the correlation factors are determined considering the sequence order correlation between continuous residues along the protein sequence. Therefore, the correlation function was determined by:

$$\Theta\left(P_{j}(n), P_{j}(n+\lambda)\right) = \frac{1}{3}\left\{\left[H_{1}(P_{j}(n+\lambda)) - H_{1}(P_{j}(n))\right]^{2} + \left[H_{2}(P_{j}(n+\lambda)) - H_{2}(P_{j}(n))\right]^{2}\right\}$$
(5)

In the prior mathematical expression, H_1 is the amino acid hydrophobicity value, H_2 is the amino acid hydrophilicity value and M its side-chain mass. These parameters were converted by the following standardization:

$$\begin{cases} H_{1}(i) = \frac{H_{1}^{0}(i) - \sum_{i=1}^{20} \frac{H_{1}^{0}(i)}{20}}{\sqrt{\frac{\sum_{i=1}^{20} \left[H_{1}^{0}(i) - \sum_{i=1}^{20} \frac{H_{1}^{0}(i)}{20}\right]^{2}}{20}}}{\sqrt{\frac{E_{i=1}^{20} \left[H_{2}^{0}(i) - \sum_{i=1}^{20} \frac{H_{2}^{0}(i)}{20}\right]^{2}}{\sqrt{\frac{\sum_{i=1}^{20} \left[H_{2}^{0}(i) - \sum_{i=1}^{20} \frac{H_{2}^{0}(i)}{20}\right]^{2}}{20}}}}{\sqrt{\frac{\sum_{i=1}^{20} \left[H_{2}^{0}(i) - \sum_{i=1}^{20} \frac{M^{0}(i)}{20}\right]^{2}}{20}}}}$$
(6)

where H_1^0 is the amino acid hydrophobicity value taken from Tanford research (C, 1962), H_2^0 is the amino acid hydrophilicity value acquired by Hoop and Woods' work (Hopp TP, 1981) and M^0 the mass of an amino acid (biofor, s.d.).

Consequently, each protein was represented by the resulting vector:

$$x_{u} = \begin{cases} \frac{f_{u}}{\sum_{i=1}^{20} f_{i} + w \sum_{j=1}^{\lambda} \theta_{j}} & (1 \le u \le 20) \\ \frac{w \theta_{u-20}}{\sum_{i=1}^{20} f_{i} + w \sum_{j=1}^{\lambda} \theta_{j}} & (20 + 1 \le u \le 20 + \lambda) \end{cases}$$
(7)

In which f_u is the normalized frequency of the 20 amino acid in a certain protein sequence, θ_j the j-tier sequence correlation and w the weight factor for the amino acid arrangement effect.

3.3 | Amphiphilic Pseudo Amino Acid Composition – Am-Pse-AAC

This method is very similar to Pse-AAC, so the order-correlated factor was defined as:

$$\begin{cases} \theta_{1} = \frac{1}{L-1} \sum_{n=1}^{L-1} H^{1}(P_{j}(n), P_{j}(n+1)) \\ \theta_{2} = \frac{1}{L-1} \sum_{n=1}^{L-1} H^{2}(P_{j}(n), P_{j}(n+1)) \\ \theta_{3} = \frac{1}{L-2} \sum_{n=1}^{L-2} H^{1}(P_{j}(n), P_{j}(n+2)) \\ \theta_{3} = \frac{1}{L-2} \sum_{n=1}^{L-2} H^{2}(P_{j}(n), P_{j}(n+2)) \\ \dots \\ \theta_{2\lambda-1} = \frac{1}{L-\lambda} \sum_{n=1}^{L-\lambda} H^{1}(P_{j}(n), P_{j}(n+\lambda)) \\ \theta_{2\lambda} = \frac{1}{L-\lambda} \sum_{n=1}^{L-\lambda} H^{2}(P_{j}(n), P_{j}(n+\lambda)) \end{cases}$$
(8)

where H^1 and H^2 were the hydrophobicity and hydrophilicity correlation functions given by:

$$H^{1}\left(P_{j}(n), P_{j}(n+\lambda)\right) = h^{1}\left(P(n)\right) \cdot h^{1}\left(P_{j}(n+\lambda)\right)$$
(9)

$$H^{2}\left(P_{j}(n), P_{j}(n+\lambda)\right) = h^{2}\left(P(n)\right) \cdot h^{2}\left(P_{j}(n+\lambda)\right)$$
(10)

where, h^1 is the amino acid hydrophobicity value and h^2 is the amino acid hydrophilicity value. Like for Pse-AAC, these parameters were converted by the following standardization:

$$\begin{cases}
H_{1}(i) = \frac{h_{1}^{0}(i) - \sum_{i=1}^{20} \frac{h_{1}^{0}(i)}{20}}{\sqrt{\frac{\sum_{i=1}^{20} \left[h_{1}^{0}(i) - \sum_{i=1}^{20} \frac{h_{1}^{0}(i)}{20}\right]^{2}}{20}}}{H_{2}(i) = \frac{h_{2}^{0}(i) - \sum_{i=1}^{20} \frac{h_{2}^{0}(i)}{20}}{\sqrt{\frac{\sum_{i=1}^{20} \left[h_{2}^{0}(i) - \sum_{i=1}^{20} \frac{h_{2}^{0}(i)}{20}\right]^{2}}{20}}}
\end{cases}$$
(11)

where h_1^0 is the amino acid hydrophobicity value taken from Tanford research (C, 1962) and h_2^0 is the amino acid hydrophilicity value acquired by Hoop and Woods' work (Hopp TP, 1981).

Consequently, each protein was represented by the resulting vector:

$$x_{u} = \begin{cases} \frac{f_{u}}{\sum_{i=1}^{20} f_{i} + w \sum_{j=1}^{\lambda} \theta_{j}} & (1 \le u \le 20) \\ \frac{w \theta_{u}}{\sum_{i=1}^{20} f_{i} + w \sum_{j=1}^{\lambda} \theta_{j}} & (20 + 1 \le u \le 20 + 2\lambda) \end{cases}$$
(12)

In which f_u is the normalized frequency of the 20 amino acid in a certain protein sequence, θ_j the j-tier sequence correlation and w the weight factor for the amino acid arrangement effect.

3.3 | Scoring

As referred in the last chapter, in this process different subsequences (k-tuples) were designated as possible variables capable of specifying a protein's a class. However, considering all the possible combinations of subsequences, this process can lead to many computational problems. One of the ways found to overcome this difficulty is to assign statistical significance to each of the variables created. In order to do so, different metrics (scores) were determined to specify several degrees of importance in the same variable on a sample or set, each sample being a protein and each variable a k-tuple.

There were several types of scores already applied in previous studies, such as the frequency of a k-tuple on a set of proteins, the linear Fisher discriminant or χ^2 statistics.

Nevertheless, considering the different forms of chemical interactions that may occur in a protein in different sections of its primary structure, other basic metrics can lead to the selection of other k-tuples essential to the correct class prediction. For example, proteins belonging to different classes may differ by the presence or absence of a certain subsequence, the number of copies of it, as well as its subsequence position in the sequence.

As a result, different scores were defined to approach the problem, so the variables may be selected to assess which factor weighs more in the deference between the proteins belonging or not to a class, and to reduce the space search of all possible variables.

3.3.1 | k-tuples

So, one way to extract potential features from the protein sequence is through k-tuples, smaller subsequences with k-amino acids. Basically by defining a k, all possible combinations from 2 to k with the 20 native amino acids were found computationally to proceed to scoring.

Thus, considering a space with all k-tuples variables as V(i), which its dimension is equal to $\sum_{n=1}^{k} 20^{n}$, each element is S_{ki} , an amino acid subsequence with k-amino acids (a_{m}) was exemplified as:

$$S_{ki}(n) = a_m^{-1} a_m^{-2} \dots a_m^{-n} \dots a_m^{-k}$$

$$a_m \in a = \{20 \text{ native amino acids}^*\}$$

$$2 \le k \le 5, i \in \mathbb{N}^+, 1 < n < 5$$
(13)

where n indicates the amino acid position the subsequence, and m identifies an element of a, supposing it has all native 20 amino acids.

3.3.2 | Frequency

In order to define different scores, initially two distinct subsequence frequencies were formulated: frequency of elements arranged consecutively and frequency of elements arranged non consecutively.

This distinction was made because a specific subsequence S_{ki} may be crucial to identify the protein function (first frequency), given that a protein sequence may have unknown or ambiguous amino acids and in the different protein structures can occur several chemical interactions between distant groups of amino acids (second frequency).

3.3.2.1 | Frequency of Elements Arranged Consecutively - FC

Thus, the frequency of a k-tuple variable S_{ki} with n elements arranged consecutively on a protein P_j was defined by:

$$FC(P_j, S_{ki}) = \sum_{n=1}^{L-k+1} [fc(P_j[n:n+k], S_{ki}), a_m^1 = b_m^n]$$
(14)

$$fc(P_{j}[n:n+k], S_{ki}) = \begin{cases} 1, \exists a_{m}^{d} = b_{m}^{c} \\ 0, \exists a_{m}^{d} \neq b_{m}^{c} \end{cases} d = c - n, \forall (P_{j}[n:n+k], S_{ki})(10)$$

The frequency of n elements arranged consecutively is the number of events that all the amino acids of S_{ki} are disposed in the designated order and consecutively in the protein sequence P_j . By analyzing P_j , when an amino acid (b_m^n) in n positon is equal to the first amino acid (a_m^1) of S_{ki} , it is checked whether the immediately remaining subsequent amino acids of P_j match the same order to the last amino acid of the S_{ki} . In this case, it must be beaded 1 to the current frequency and the next the amino acid to analyze in P_j is located at n + k + 1. If not, the next amino acid to be evaluated has n+1 position.

3.3.2.1 | Frequency of elements arranged non consecutively - FNC

Given the previous principles, for a protein P_j it is possible to determine the frequency of elements arranged non consecutively in the following:

$$FNC(P_j, S_{ki}) = \sum_{n=1}^{L-k+1} [fnc(P_j[n; L], S_{ki}), a_m^{-1} = b_m^{-n}]$$
(15)

$$fnc(P_{j}[n:L], S_{ki}) = \begin{cases} 1, \exists a_{m}^{d} = b_{m}^{c} \\ 0, \nexists a_{m}^{d} = b_{m}^{c}, d \leq c - n, \forall S_{ki} \end{cases}$$
(16)

So, in P_j when an n amino acid (b_m^n) is equal to the first amino acid (a_m^1) of S_{ki} , it continues to search for the second amino acid a_m^2 in P_j located after b_m^n regardless the position of the same amino acid may occupy. The current frequency will increase by one when all S_{ki} 's amino acids are found on the sequence protein in the same order they are presented. After, the search for a_m^1 in P_j begins again at the amino acid following the first one found before, and all the amino acids once considered are ignored on the succeeding searches.

The way this frequency is defined, the elements of S_{ki} arranged consecutively in the protein sequence are also being considered, but in less number.

3.3.3 | Data Sets

To correctly evaluate the k-tuples found, and select the best representative features, three sets were created in which each subsequence is evaluated: The set of proteins that will be used to train a model (data_all), the set with the proteins which in the previous set belong to the class (data_pos) and another set with proteins which are not part of it (dara_neg). By doing so, it is possible to review the different k-tuples without the scores being excessively influenced by the unbalance between the number of proteins from the class and the number which do not belong. There is also a possible way to clearly find the features that best characterize each case.

3.3.4 | Scores

Diverse categories of scores were created in order to analyze each subsequence (S_{ki}) considering different characteristics, such as its number of elements (a_m) , its frequency in a defined protein set, how discriminant it is for different types of proteins, among others.

3.3.4.1 | Absolute Set Frequency – ASF

This score is the simplest way to evaluate the significance of a subsequence on a certain protein set. Basically, it determines how important a subsequence is by measuring its frequency on each protein. Some repeated subsequences can be important in a protein structure and consequently in its metabolic function.

For the subsequence S_{ki} and certain set with N proteins, this score was the sum of the subsequence frequency in each protein as it is defined next.

3.3.4.1.1 | Absolute Set Frequency of Elements Arranged Consecutively – ASFC

$$ASFC(S_{ki}) = \sum_{j=1}^{N} FC(P_j, S_{ki})$$
(17)

3.3.4.1.2 | Absolute Set Frequency of Elements Arranged Non Consecutively – ASFN

$$ASFN(S_{ki}) = \sum_{j=1}^{N} FNC(P_j, S_{ki})$$
(18)

3|Methods

3.3.4.2 | Absolute Frequency by Presence – AFP

This score evaluates whether a certain subsequence is present or not in a protein sequence, rather than its frequency. Consequently, the score will be higher the more proteins has the subsequence in question. This score was defined this way because a certain k-tuple can be very common in some proteins, however, it is not expressed significantly throughout the defined set or by its frequency in the sequence.

So, for a set with N proteins, a subsequence (S_{ki}) will have as a score the number of proteins in which it had a frequency higher than 0. This score was used in order to take into consideration the two types of frequency.

3.3.4.2.1 | Absolute Frequency by Presence of Elements Arranged Consecutively – AFPC

For a certain subsequence S_{ki} the Absolute Frequency by Presence of Elements Arranged Consecutively is determined by:

$$AFPC_i(S_{ki}) = \sum_{j=1}^{N} Ec\left(P_j, S_{ki}\right)$$
(19)

$$Ec(P_{j}, S_{ki}) = \begin{cases} 1, & if \ FC(P_{j}, S_{ki}) > 0\\ 0, & if \ FC(P_{j}, S_{ki}) = 0 \end{cases}$$
(20)

In which N represents the total number of proteins in a defined set, P_i is one of those proteins.

3.3.4.2.2 | Absolute Frequency by Presence of Elements Arranged Non Consecutively – AFPN

The AFPN is defined by:

$$AFPN(S_{ki}) = \sum_{j=1}^{N} Enc(P_j, S_{ki})$$
(21)

$$Enc(P_j, S_{ki}) = \begin{cases} 1, if \ FNC(P_j, S_{ki}) > 0\\ 0, if \ FNC(P_j, S_{ki}) = 0 \end{cases}$$
(22)

3.3.4.3 | Relative Frequency by Presence – RFP

This score is similar to the last one, the difference is that RFP takes into account the number of proteins in the defined set. Of course, in MaxScoring the previous score and RFP will lead to the selection of the same sequences in data_all. However, this score was not created

to be used in data_all, but rather to create a different selection for the remaining sets, which shall be addressed in the next section with a more detailed description of MaxScoring.

Also the two types of frequency were considered, therefore the two types of scores shall be described next.

3.3.4.3.1 | Relative Frequency by Presence of Elements Arranged Consecutively – RFPC

$$RFPC(S_{ki}) = \frac{1}{N} \sum_{j=1}^{N} Ec\left(P_j, S_{ki}\right)$$
(23)

$$Ec(P_{j}, S_{ki}) = \begin{cases} 1, & if \ FC(P_{j}, S_{ki}) > 0\\ 0, & if \ FC(P_{j}, S_{ki}) = 0 \end{cases}$$
(24)

3.3.4.3.2 | Relative Frequency by Presence of Elements Arranged Non Consecutively – RFPN

$$RFPN(S_{ki}) = \frac{1}{N} \sum_{j=1}^{N} Enc\left(P_j, S_{ki}\right)$$
(25)

$$Enc_{j}(S_{ki}) = \begin{cases} 1, if \ FNC_{j}(S_{ki}) > 0\\ 0, if \ FNC_{j}(S_{ki}) = 0 \end{cases}$$
(26)

3.3.4.4 | k-Frequency – KF

It would be incorrect to evaluate the frequency of the variables disregarding the length of each one. The more amino acids the variable has, the lower its frequency is comparing to the features of AAC. In this sense, to try a solution, this score was created, which attempts to benefit variables with more elements, or at least to enable their evaluation more evenly.

Therefore, KF is the ratio between the sum of a subsequence frequency in each protein and k, variable defined when all k-tuples were defined. [see 3.1 k-tuples]

In this score, both frequencies were used like it is shown below.

3.3.4.4.1 | K-Frequency of elements arranged consequently – KFC

$$KFC(S_{ki}) = \sum_{j=1}^{N} \frac{FC(P_j, S_{ki})}{k}$$
(27)

3.3.4.4.2 | K-Frequency of elements arranged non consequently – KFN

$$KFN(S_{ki}) = \sum_{j=1}^{N} \frac{FNC(P_j, S_{ki})}{k}$$
(28)

3.3.4.5 | Fisher Score – FS

This score is based on the Fisher linear discriminant analysis, a method used in pattern recognition, machine learning, and other fields of expertise, to find a linear combination of features that characterizes and/or separates two or more classes of objects or events. Considering a class of proteins, it was defined that the two events assessed would be if the protein belongs (labeled as 1) or not to the same class (labeled as 0).

In previous works, the same score was used in order to find the discriminating variables between different protein classes with extended formulas, but in this case the way that all sets and classes are defined is not possible to do the same, and also before knowing what features separate different classes, it is more important to know what subsequences can characterize a protein in a way to show if it belongs or not to a certain class, since, proteins share similar characteristics when it comes to their functionality: some have multiple functions, and therefore, multiple classes.

Accordingly, for a certain set of N protein, FS of a certain subsequence S_{ki} is a ratio in which the numerator is the square of the difference between the mean frequency of S_{ki} in proteins labeled as 1 and 0, and the denominator is the sum of the variances of those frequencies.

3.3.4.5.1 | Fisher Score of elements arranged consequently – FSC

$$FSC(S_{ki}) = \frac{\left|\overline{FC(P_{J},S_{ki})}\right|_{l=1} - \overline{FC(P_{J},S_{ki})}\right|_{l=0}|^{2}}{s_{c}^{2}|_{l=1} + s_{c}^{2}|_{l=0}}$$
(29)

3.3.4.5.2 | Fisher Score of elements arranged non consequently – FSN

$$FSN(S_{ki}) = \frac{\left|\frac{FCN(P_{j}, S_{ki})}{s_n^2}\right|_{l=1} - \frac{FC(P_{j}, S_{ki})}{s_n^2}\right|_{l=0}^{l}}{s_n^2|_{l=1} + s_n^2|_{l=0}}$$
(30)

3.3.4.5 | Term frequency-inverse document frequency - TF-idf

The concept around this score is often used in information retrieval and text mining. Basically, Tf-idf is a statistical measure that shows how important a word is in one or more documents by evaluating how infrequently the word is across them. In a protein sequence, rare subsequences are also important to predicts its classes. The presence of one or few rare subsequences can be crucial to define a particular functionality, and all the scores before aren't able to evaluate that, therefore the less frequent subsequence is the higher the score is.

The tf-idf of a term t in a document d is defined as the product of the term frequency (tf) with the term inverse document Frequency (idf). The tf is the quotient between the number of times t appears in d and the total number of terms or words present in the same document. As for idf, it is determined by the logarithm of the total number of documents divided by the number of documents with the same term in it. Consequently, the tf-idf of t in several documents is the sum of the operation explained above for each document.

In this case, the documents are the protein sequences and the words or terms are the subsequences. Therefore, for a certain subsequence S_{ki} and for the protein sequence j the term frequency can be defined by:

$$tf = \frac{f(P_j, S_{ki})}{L_j} k_i \tag{31}$$

where f is one of the type of frequencies previous defined, k_i is the number of amino acids present on S_{ki} and L_j is the subsequence length.

This tf was considered in this way because a protein sequence doesn't have a total number of terms or words, it's a sequence, thus considering a potential term as an amino acid set with equal size as the subsequence involved, the total number of terms could be the number of times that the protein can contain the subsequence, taking into account how its frequency has been set. As a result, the total number of terms in a protein is the ratio between its total number of amino acids, and the total number of amino acids in a subsequence.

More detailed formulas are showed next, considering a set with N proteins and a certain subsequence, using both types of frequencies.

3.3.4.5.1 | Term frequency-inverse document frequency of Elements Arranged Consecutively – tfc-idf

$$tfc - idf(S_{ki}) = \sum_{j=1}^{N} \frac{FC(P_j, S_{ki})}{L_j} k_i \times \log_{10}\left(\frac{N}{FAPC(S_{ki})}\right)$$
(32)

3.3.4.5.2 | Term frequency-inverse document frequency of Elements Arranged Non Consecutively – tfn-idf

$$tfn - idf(S_{ki}) = \sum_{j=1}^{N} \frac{FNC(P_j, S_{ki})}{L_j} k_i \times \log_{10}\left(\frac{N}{FAPN(S_{ki})}\right)$$
(33)

3.3.4.6 | X^2 statistics – XS

This score is based on \mathcal{X}^2 statistics, which is basically used to evaluate if the distribution of two categorical variables differ from each other in a single population. Essentially, this can be used to determine if there is a significant correlation between the two variables. In this case the two categorical variables are whether a protein belongs or not to a class and if it contains a certain subsequence or not.

So, by using the two-way contingency table it was defined as a score:

$$XS(S_{ki}) = \frac{N \times (AD - CB)^2}{(A + C) \times (B + D) \times (A + B) \times (C + D)}$$
(34)

where A is the number of positive proteins with S_{ki} , B is the number of negative proteins that have the it, C stands for the number of positives proteins that not contain it, D is the number of negative proteins without the subsequence in theirs sequences and N is the number of proteins involved (positive and negative).

It is important to add that this score has a natural value of zero if both variables are independent, so it will be used only if appear subsequences scored above 0.

This score also was used for both defined frequencies.

3.3.5 | MaxScoring

Once the scoring is complete, the next step is the selection of a number of subsequences for each score. So, depending on the chosen set of protein, it is defined the number of variables for each score (M). The M variables with the highest score in each category previous defined will be selected, and after those subsequences it will became features of the same set to construct a potential model through SVM classifiers. This is only for data_all, for

the data_neg and the data_pos sets of M/2 variables will be selected for each score, and then the combined features of both will became the features to a model.

For the score RFP, this process works a little different. For the data_neg and the data_pos sets, the variables are compared in both sets, and they are selected for each set if the score was higher in it. For example, during the search for the most significant variables, if "aa" has a high score, it would only be chosen for the set where it showed a higher score.

In addition, MaxScoring was executed in two different ways. The first one the variables were selected only by the score they presented (MaxScoring Normal – MS-N). As for the second one, besides the score it is important the variables shared similarities. Basically when searching for variables with the high scores if it is found two variables that distinguish in length but the larger contains the smaller, the last one is eliminated from the search (MaxScoring without parent subsequences – MS-WPS).

3.4 | Particle Swarm Optimization – PSO

The Particle Swarm optimization algorithm (PSO) is a computational method that optimizes a given problem by iteratively measuring the quality of the various solutions.

In this context, PSO is used to select the best features to characterize each class by using as fitness function the AUC (area under the curve) obtained.

3|Methods

```
1: //initialize all particles
    2: Initialize
   3: repeat
                         for each particle i in S do
   4:
                                              update the particle's best position
   5:
                               if f(x_i) < f(pb_i) then
    6:
    7:
                                      pb_i = x_i
    8:
                                   end if
   9:
                                           /update the global best position
                                  if f(pb_i) < f(gb) then
10:
                                            gb = pb_i
11:
                                   end if
12.
                         end for
13:
14:
15:
                            //update particle's velocity and position
                   for each particle i in S do
16:
17:
                                 for each dimension d in D do
                                              v_{i,d} = v_{i,d} + C_1 * Rnd(0,1) * [pb_{i,d} - x_{i,d}] + C_2 * Rnd(0,1) * [gb_d - x_{i,d}] + C_2 
18:
                                              x_{i,d} = x_{i,d} + v_{i,d}
19:
20:
                                  end for
21:
                         end for
22:
                            //advance iteration
23:
                     it = it + 1
24:
25: until it > MAX_ITERATIONS
```

As shown in Figure 3, the PSO consists in the initialization of a group de particles and

Figure 3 - Pseudo Code of the Particle Swarm Optimization Algorithm used for feature Selection (ist, s.d.)

search for best solution/particle by updating theirs positions $(x_{i,d})$ and velocities. In each iteration, each particle is updated by the following two best values. The best solution achieved during a iteration is pb_i, and gb_d is the best value obtained by any particle in the population S in any iteration. The particle updates its velocity and positions with the equations expressed in the figure where v_{i,d} is the particle velocity, Rnd () is a random number, C1 and C2 are learning factors.

3.4.1 |Hybrid Features – Initialization of the particles

Comparing to previous studies, in this context, the PSO was used differently for feature selection. From the prior sections, it is clear that are different types of variables as potential features for a class classifier, each different type of variable concerning different aspects that needed to be evidenced at a given dataset.

Accordingly, it was decided to use the PSO to group all these variables so it can be considered all the chemical and biological indications discussed before, and also all can be fairly part of a possible model at the same time. This means a distinct particles initialization and the presence of features with different backgrounds - Hybrid features.

So we have two different sets of variables: AAC and k-tuples variables selected by MaxScoring. The variables involving AAC were always present in each particle, as well as the k-tuples.

So, for a given dataset was created a number of particles same as the number of the existing scores for the three types of sets. [See section 3.3]. Thus, with all scores it existed 12 scores for data_all and 12 scores from other sets, making a total of 24 particles. Although it may be less if the score XS is not relevant, that is if in MaxScoring the maximum values always found were 0. This means that each particle represents a score for a determined set. Each particle dimension will be 20 (AAC) plus the number of all k-tuples found in all considered scores.

The Initialization of each particle was done accordingly to each score, where each element of x_i (a specified particle) was 1 in the positions relative to variables from AAC to the k-tuples variables that belong to the score, the remaining elements are 0. For example, considering the particle representative of a FSC for data_all, all elements corresponding to the k-tuples selected by Max Scoring for that score and that set, as well as to the AAC have the value of 1, while the others will be equal to 0.

The particles initialization was defined in a way to reduce the search-space without ignoring all possible combinations of different variables as best features.

3.4.2 | Different Uses of Particle position to perform Feature Selection

The Feature Selection is performed through the values defining the position particle (x_i) by three different ways.

The first one, every feature chosen as input for the PSO algorithm is used to construct a possible model but the vector with the frequencies of each one was multiplied by the corresponding position vector of a certain particle – PSO-W. This way, the position components of a particle were viewed as weights, because it was correct to assume some features could be more important than others and this could be a solution to find how important each one can be.

Another method is removing features if its corresponding position component from the particle was less than 0 - PSO-F.

And lastly, the two previous approaches are considered, so the features in which the position components of particle vector position are negative are ignored and for the remaining features their frequencies will be multiplied by the corresponding weights – PSO-WF

3.4.3 | Fitness Function – Data Structure

For each interaction, each particle consists of a defined data structure to construct a model through SVM, so then could be used in a test set to obtain the AUC.

After defining which PSO will be used to deal with the chosen features in each particle, for each protein it will be determined the variable frequency, depending on the nature of the hybrid feature. If is a k-tuple three situation could occur.

The first two was being able to choose how to determine a k-tuple frequency in a protein sequence, if FC or FNC, because they are related and performing different types of frequency in different k-tuples even if they were chosen by a score with other type of frequency is not completely wrong. And also because k is set for 5 so the variables are small enough to be significant for both types of frequency. It was decided to do this also to see how both frequencies can influence the classifiers' performance as well as to begin the implementation with simpler tests to see the improvement as the work progresses.

The other scenario was the definition of each subsequence frequency according to its provided score. If a k-tuple is present in different scores influenced by different frequencies, then a duplicate is created for that k-tuple, resulting into two representative vectors. One with the FC of that variable and the another with the absolute difference between FC and FNC. This way, the type of score will be considered not only by selecting which k-tuple will be in the algorithm but also how the data will be constructed for the respective class classifier.

4|Practical Applications

In this chapter, it was described two essential tools that were created to enable the use of the referred methods easily by any user. Thus, it was created a graphical interface in C # and a website, in order to complement the research with a practical component.

4.1 |C# Application

A C# Application that handles all necessary scripts was created in order to the methods could be implemented properly by any user not familiar with the covered procedures or with no experience in manipulating the different scripts created, or even who is interested in performing experiments with other datasets.

At first the user can see the project name, and two buttons one of which directs the user to a task menu (figure 4), and the other opens a window that allows the user to verify all python necessary libraries in order to run any command.

The task menu (figure 5) consists in 5 different buttons and each of them shows a different type of task: date set settings, algorithms based on AAC, tasks related to the Scoring, the application of PSO, and finally the view of excel files containing results.



Figure 4 – Initial C# Application window

To perform any task, first the user must choose which data set he wants to use if any was previously saved, or he has the option to add data set by choosing the files present on the computer. When choosing a dataset is allowed to the user to execute AAC, Pse-AAC or Am-Pse-AAC (figure 6) and the results can be viewed by clicking on files button.

If the user selects a dataset and a number of range of classes, the user can start the Scoring, MaxScoring, and he can also choose a score or more and try to predict proteins classes without using PSO.

After having performed the scoring and MaxScoring to a certain data set at least once, the user can use to the PSO according to the settings.

In Files Manager (last button), the user as access to all files with results of the chosen dataset. The user can visualize and perform basic operations like determining the mean, the minimum and maximum of a group of values.

| Evolutionary Computation Systems for Biological Data Analysis

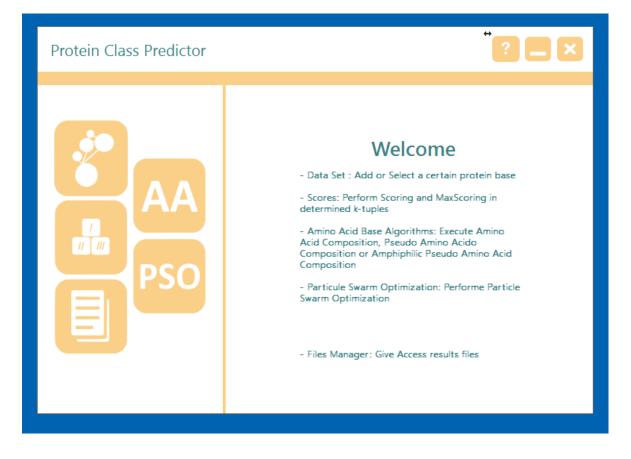


Figure 5 – Task Menu Panel

The control of python scripts as well as of files that are being created is done by executing intermediate python scripts (created on purpose for this interface) through the computer command line. This leads to the command prompt window appear when starting a task, or doing the implementation of a method, to inform the different stages of the running method.

It is important to point out that the application warns the user about the minimal requisites needed to perform all this methods, as well as the high execution time. Also to run this application is needed a great number of files and scripts.

| Protein Class Predictor | | | | | |
|--|---|--|--|--|--|
| AA M M M M M M M M M M M M M M M M M M | Scoring k: No MaxScoring New Scores No dataset k: M: Type: Y | Select Scores No dataset k: Y M: Type: | | | |
| | | Scores: | | | |

Figure 6 – Scoring and MaxScoring Panel

4.2 |Web Page

In addition to the application it was created a Web Page in which the user has information about the project (figures 7 and 8). It is also where is available the application as well as recommendations on how to use is. Similarly has a sector in case a possible user does not want to use the application, but wants test the proposed methods, he can submit files and the application of the methods is performed with the resources available at the moment, user only receives the results or the associated notifications.

| Welc | ome | | |
|------|--------------|-------------|---------------------|
| | About | Downloads | Co a factor control |
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| | Methods | Contacts | |
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Figure 7 – Initial Menu Shown at the created Web Page



Figure 8 – The available information about the used methods at the created Web Page

4 | Practical Applications

5 | Results

In this chapter, it was discussed the main characteristics of the chosen data sets, in what way the proposed method was executed, all the different testes performed, as well as the results obtained in order to be properly discussed in the next chapter.

5.1 |Materials

In order to verify the explained methods efficiency two well-known sets of proteins were chosen. These sets of proteins were used to implement various algorithms that are based only on primary sequence or other attributes.

One of the chosen proteins sets is present at Protein Classification Benchmark Collection (Paolo Sonego, 2006) and It is referred as SCOP40mini. This set has 1357 proteins belonging to a protein superfamily named SCOP95, in which each protein, considering its biological chemical characteristics may or may not belong to one of the 55 different groups (55 classification tasks).

The protein sequences and the definition of the train and test sets are provided for the same database. As shown in table 9 in Appendix, the train and test sets have a low percentage of positives (proteins belonging to the class), when compared to the negatives number present (proteins which are not part of the class), which makes the classification task more difficult but also realistic in a possible scenario where the methods referred above can be used as future research tools.

The second set was created by Kuo-Chein Chou and David W.Elrod in order to predict the enzymes family classes (Chou & Elrod, 2003). From ENZYME database, it was selected a set of oxidoreductases whose sequences were obtained through SWISS-PROT. This protein set, which will be referred to as Chou and Elrod 2003, it contains 2640 proteins distributed by 16 classes of different enzymes families (table2).

In this set, it was defined two different subsets of proteins: Train set to create a possible classifier and a test set to obtain a AUC in order to estimate the classifier performance. The proteins of each subset were chosen randomly and the negatives are in the same number as the positives, in order to have a more balanced data set. It was only used 85% samples of each class, and then it is divided into 15% and 75% for test and train sets.

| Class | Train Set | Test Set |
|-------|-----------|----------|
| 1 | 177 | 89 |
| 2 | 122 | 61 |
| 3 | 109 | 55 |
| 4 | 73 | 37 |
| 5 | 63 | 32 |
| 6 | 172 | 87 |
| 7 | 36 | 18 |
| 8 | 33 | 17 |
| 9 | 143 | 72 |
| 10 | 52 | 27 |
| 11 | 86 | 44 |
| 12 | 52 | 27 |
| 13 | 145 | 73 |
| 14 | 87 | 44 |
| 15 | 47 | 24 |
| 16 | 86 | 44 |

Table 2 - Number of Proteins (positives) in each Subset for each class of Chou and Elrod 2003.

For this set of proteins, one of the problem encountered is the number of proteins present in some classes, in which the size of the train size is very small, like classes 7, 8 or 15.

The selection of each of these data sets was made with different objectives. First in a research like this, it is necessary to prove that the method is effective and better than any other applied in the same data set, for this was chosen Chou and Elrod 2003, because it was used before by other methods and it is a balanced data set, created to use methods based on AAC.

Second, it is also important to show that the method works with other data set with different characteristics, and for this was chosen SCOP40 mini, a data set with classes with a very low percentage of negatives. And finally, it is necessary analyze the proposed method in order to understand what are the key variables or attributes that can influence its performance, as well as what its main limitations, for that SCOP40mini was also used.

5.2 |Experiments

In order to understand all work performed is important to explain how the different methods were implemented, the applications used or created, all the defined variables, as well as the different experiments conducted to demonstrate the efficiency of the proposed method.

5.2.1 |Execution Scripts and Files

All methods were implemented in Python along with tools present in scikit-learn (developers, s.d.), Openpyxl (OpenPyxl, s.d.), numpy and scipy.

The methods implementation from the C# application has already been addressed in Chapter 4, but it is necessary to refer the files and folders created and involved in the execution of the main python scripts, in order to demonstrate the structure used for an easy understanding if in need to check or change a specific file.

So, three principal python scripts were created: one is responsible for implementing all the methods related to feature extraction and selection (FeatureGeneration.py), another for the creation and application of SVM classifiers (Classifiers.py) and the last one creates the necessary structures to serve as input to train classifiers (processData.py).

For each data set is created a folder that contains subfolders for each class, other for materials and other for results. For each class, the subfolder contains all the files created from scoring (text files with a simple table in which each row is a subsequence and each column one score), and MaxScoring (text files with the chosen subsequences). In materials folder are all files containing the protein sequences as well as files with the information necessary to identify the train and test sets. As for the results folder, subfolders are created according to different tests with different chosen variables, if PSO is used for each class the created classifiers are stored, as well as the particle's components and subsequences for best results.

5 | Results

5.2.2 |Variables

The results as well as the research were carried out according to two distinct stages: the study of the different variables involved and the search for the best combination of them. The first step involves performing tests in which only one variable is changed in order to realize how it influences the average AUC obtained, and the second stage is finding the best combination of variables that provides the best average AUC for a chosen dataset.

The variables in question are k, type of MaxScoring, M, type of PSO, type of frequency and the number of iterations, in order to obtain the best average AUC value with the lowest number of iterations (lowest running time required).

For SCOP40mini, the proposed algorithm was tested with M values from 6 to 20, but only the results obtained with M equal to 10 and 20 are been illustrated. Regarding the k used, the PSO was only implemented with k equal to 3 and 5, since when trying to predict the families' classes only using MaxScoring, it was concluded that these values were the most significant (higher average AUC). As for the other dataset, M values used were never higher than 10 to avoid overfitting.

For PSO, the different parameters were defined based on previous researches, therefore, the inertial constant is 0.5 plus a random value between 0 and 1 divided by 2, the sum between the cognitive and social parameters (C1 e C2) must be less or equal to 4, so both are considered 2. The maximum velocity of particles was set as 2.5 and the stopping criterion was the number of iterations that the global best value remains the same, and it can be decided by the user. In the experiments, it was defined 100, 200, 500 iterations.

The performance of each method was evaluated according to the average AUC obtained, as well as the accuracy in order to compare with results from other papers. The experiments were performed to compare the AAC, Pse-AAC, Am-Pse-AAC, each individual score, and then all scores in combination and finally with all the features from AAC and the scores selected by the PSO.

5.2.3 |SVM training

For each class was created a classifier in which will only identify whether a given protein belongs or not to the same class, so for each test study were created 55 independent classifiers to SCOP40mini, and 16 independent classifiers to Chou and Elrod 2003.

In the first phase of the results, the classifiers were created according to the default settings in scikit-learn for a linear kernel, because it was necessary analyze quickly each of the variables that can influence the proposed method.

As for the second phase, the SVM classifiers were defined according to the best parameters obtained by performing a search grid for each class, which was performed for the gaussian kernel with the gamma values between 0.001 and 0.0001, and C values between 1 and 100, and for the linear kernel with the same C values referred above.

5.3 |New Method Analysis – Variables Study

In this section, it is demonstrated the different results obtained from the different possible combinations that led to the best results.

Due to the high number of subsequences as variables, and given that the proposed method has a high runtime, that is why it was first performed an analysis without search grid in order to understand what would be the definitions that could lead to better results. At this stage the tests were performed only with Scop40mini data set. It is worth mentioning that in some results are illustrated only values obtained for some classes, because the statistical tendencies observed in the showed classes are the same for the remaining classes.

5|Results

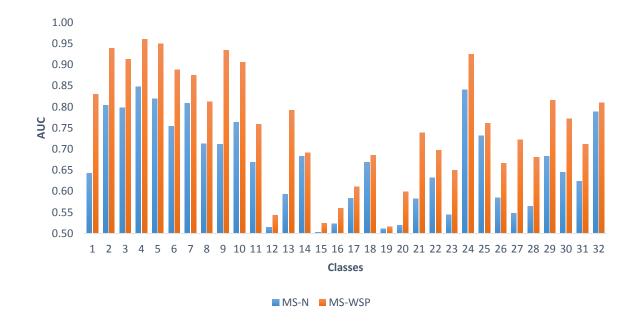
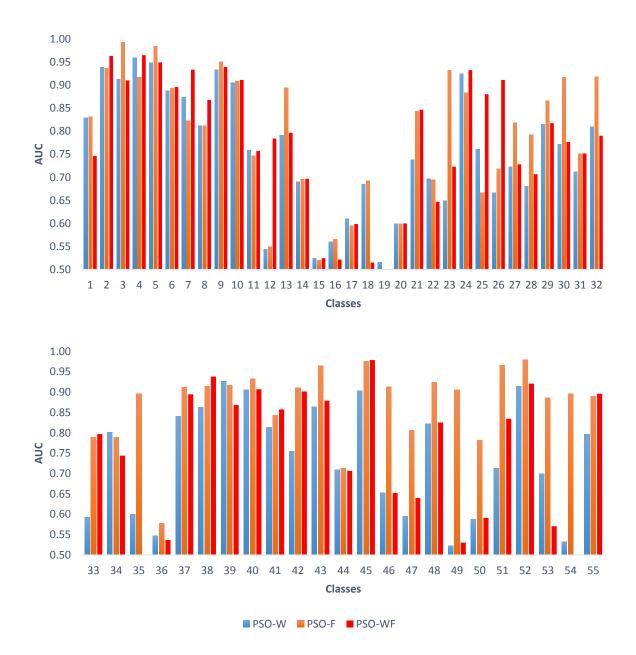
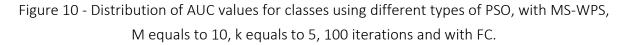


Figure 9 – Distribution of AUC values for some classes using different types of MaxScoring with PSO-W, M equals to 10, k equals to 5, 100 iterations and with FC.

One of the variables to be considered is the type of MaxScoring used, as shown in Figure 9, all classes using MS-N leads to lower AUC values than when using MS-WPS. The average difference between the values obtained by different types of MaxScoring is approximately 0.08, in which it was registered a maximum difference of 0.22 for Class 9.

As for different types of PSO (Figure 10), it was observed higher AUC values for classes like 11, 19, 22, 34 and 39 with PSO-W, while for classes 2, 4, 6, 7, 8, 10, 12, 14, 20, 21, 24, 25, 26, 33, 38, 41, 45 and 55, the best results were obtained when using PSO-WF. About the remaining classes the best values were documented using PSO-F. Thus, PSO-F was the type of PSO with the best results in a larger number of classes. The average AUC obtained for PSO-W was 0.749, for PSO-F 0.82 and 0.77 for PSO-WF. This tendency repeats itself for the other data set as well.





As for the M values, all the experiments were performed with M equal to 10, however it was decided to test with other M values in order to realize if AUC is affected.

According to Figure 11, approximately 26 classes obtained better AUC values with higher M, however the impact of the M increase on average AUC values was insignificant, having registered an average AUC value of 0.823 for M equal to 10 and 0.811 for M as 20. Other tests were performed with M values less than 10, but it was always obtained an average AUC value slightly lower than with M equal to 10 (about 0.04 less).

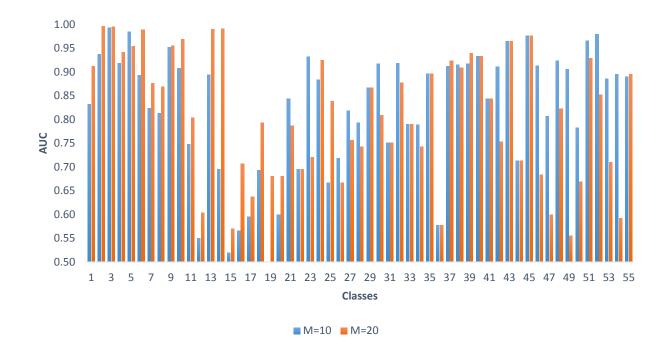


Figure 11 - Distribution of AUC values for each class using different M values, with MS-WPS, PSO-F, k equals to 5 and with FC.

About the type of PSO used with each type of frequency, observing Table 3, there were better results when used FC. However, when used both frequencies the average AUC value improves slightly in some tests, providing the best results shown in the next section.

Regarding the number of iterations required to obtain the best results, it was possible to observe during the tests that with higher number of samples in the train sets more iterations are necessary. The number of iterations was also influenced by the value of k, the higher it was more iterations were needed. For k equal to 5 only with more than 100 iterations it was possible to obtain a mean AUC over 0.8 but with k equal to 3 was possible to obtain acceptable AUC values with more than 50 iterations. The use of search grid also led to being necessary to increase the number of iterations, since each iteration a new selection of features meant another set of parameters to train a SVM classifier.

Table 3 – The increase in the average AUC value when used PSO-F, M equals to 10, k equals to 5, 100 iterations and with two different frequencies (and FC FNC) for 20 classes.

| Class | FC | FNC |
|---------|----------|----------|
| 1 | 0,233483 | 0,158408 |
| 2 | 0,320504 | 0,257898 |
| 3 | 0,246956 | 0,245434 |
| 4 | 0,143836 | 0,118287 |
| 5 | 0,074159 | 0,120413 |
| 6 | 0,096942 | 0,204587 |
| 7 | 0,367482 | 0,219164 |
| 8 | 0,19356 | 0,160297 |
| 9 | 0,024344 | 0,06000 |
| 10 | 0,101499 | 0,129862 |
| 11 | 0,199975 | 0,148064 |
| 12 | 0,358346 | 0,195423 |
| 13 | 0,477099 | 0,38855 |
| 14 | 0,467939 | 0,467176 |
| 15 | 0,370102 | 0,146819 |
| 16 | 0,366739 | 0,11385 |
| 17 | 0,30528 | 0,123602 |
| 18 | 0,420807 | 0,289907 |
| 19 | 0,363742 | 0,301501 |
| 20 | 0,230901 | 0,279814 |
| Average | 0,268185 | 0,206453 |

5.4 |Results with best average AUC value

In this section, it is present the main results required a short analysis of the best results obtained, as well as a comparison between the proposed method and methods based on AAC.

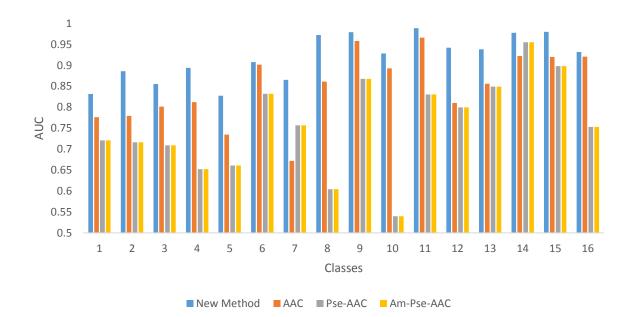


Figure 12 - AUC Values obtained in each class with different approaches using Chou and Elrod 2003. For Pse-AAC λ was 10, Am-Pse-AAC λ was 9, and for the proposed method was used PSO-F with M equal to 10, k equal to 5, MS-WSP and 200 iterations.

As For Figure 12, it was observed that the proposed method achieved better results than Pse-AAC, Am-Pse-AAC and AAC in all classes for Chou and Elrod 2003, also that the AAC and Pse-AAC with the training sets created, had similar AUC values. It should be emphasized that the classes in the AAC based methods have a worse performance (class 5, 7 and 8), and the proposed method achieved values above 0.80.

Referring to figure 13, the implementation of Pse-AAC and Am-Pse-AAC led to very low AUC values, most classes obtained 0.5. Despite the failure of the methods based one AAC, witj the proposed method was obtained AUC values above 0.70.

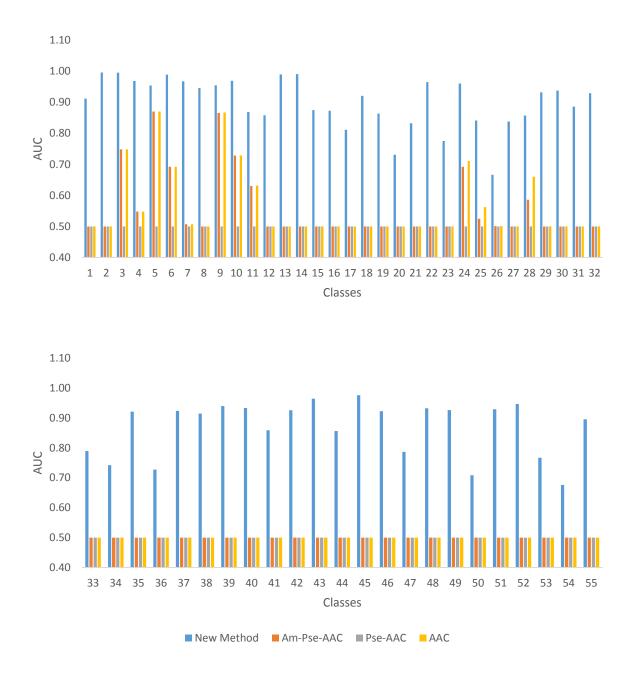


Figure 13 - AUC Values obtain in each class with different approaches using SCOP40mini. For Pse-AAC λ was 10, and Am-Pse-AAC λ was 9, for the proposed method was used PSO-F with M equal to 10, k equals to 5, MS-WSP, and 200 iterations without search grid.

Table 4 – Average AUC Values and Average Accuracy values obtained with different approaches using Chou and Elrod 2003. For Pse-AAC λ was 10, Am-Pse-AAC λ was 9, and for the proposed method was used PSO-F with M equal to 10, k equal to 5, MS-WSP, and 100 iterations. Also it is present the improvement determined comparing to AAC and Am-Pse-AAC.

Chou and Elrod 2003

| | A | | | 1 (0() |
|---|-------------|------------------|---------|----------|
| Methods | Average AUC | Average Accuracy | Improve | ment (%) |
| Implemented | | | | |
| AAC | 0,849 | 0,848 | - | - |
| Pse-AAC | 0,759 | 0.756 | -10,603 | - |
| Am-Pse-AAC | 0,759 | 0,756 | -10,603 | - |
| MaxScoring - All Scores | 0,573 | 0,515 | -51,352 | -24,506 |
| Proposed Method | 0,919 | 0,920 | 8, 245 | 21,080 |
| Previous Studies | | | | |
| Am-Pse-AAC and SVM (Chou, 2004) | - | 0,809 | | |
| Am-Pse-AAC with Covariant-Discriminant Algorithm (Chou, 2004) | - | 0,766 | | |
| Covariant-Discriminant Algorithm (Chou, 2001) | - | 0,755 | | |

Based on the Table 4, it can be affirmed that the proposed method can improve any of the others known methods, because it was obtained an average AUC value of 0.919 and average Accuracy value of 0.92, higher than any of the other methods implemented, and higher than any other values registered in other researches with the same data set.

Table 5 - Average AUC Values and Average Accuracy values obtained with different approaches using SCOP40mini. For Pse-AAC λ was 10, Am-Pse-AAC λ was 9, and for the proposed method was used PSO-F with M equal to 10, k equal to 5, MS-WSP, and 500 iterations. Also it is present the improvement determined comparing to AAC and Am-Pse-AAC.

| | SCOP | 40mini |
|-------------------------|-------------|-----------------|
| | | |
| Methods | Average AUC | Improvement (%) |
| Implemented | | |
| AAC | 0,537 | - |
| Pse-AAC | 0,500 | -6,890 |
| Am-Pse-AAC and SVM | 0,534 | -0,554 |
| MaxScoring - All Scores | 0,646 | 20,298 |
| Proposed Method | 0,888 | 65,363 |

Regarding Table 5, with the proposed method achieved an improvement of 65% compared to AAC, and without search grid reached better results than with SVM classifiers using as features the distance matrixes Lempel-Ziv-Welch and by Partial Match (Gaspari, sd), whose average AUC values are respectively 0.8288 and 0.8551 (pongor, s.d.)

Table 6 – Principal SVM Classifiers parameters found in Search grid for Chou and Elrod 2003 set.

| Kernel | с | gama |
|--------|-----|--------|
| | 10 | 0,0001 |
| rah | 100 | 0,0001 |
| rgb | 10 | 0,001 |
| | 1 | 0,001 |

Considering classifiers' parameters for the dataset Chou and Elrod 2003, it was possible to determine that the best parameters were set to a Gaussian kernel, and C ranges from 1.10 to 100 and gamma the range was between 0.0001 and 0.001.

About the subsequences chosen by PSO, it can be established that are present subsequences with different k and some amino acids weren't selected in both datasets. Observing Table 7, the score with a larger number of selected features is the XSN_ALL, the score XSN with data_all, and the less influential was RFPN_Pos (because in some classes this score didn't have any selected features). It is important to add that in each class the number of features selected by each score is different, each class had its unique combination of number of features per score.

Table 7 – The average number of features selected in each score in each class and set for Chou and Elrod 2003

| Scores | Average of number features |
|-------------|----------------------------|
| AFPC_All | 5,67 |
| AFPC_Neg | 2,67 |
| AFPC_Pos | 2,60 |
| AFPN_All | 6,20 |
| AFPN_Neg | 3,47 |
| AFPN_Pos | 3,53 |
| ASFC_All | 5,33 |
| ASFC_Neg | 2,67 |
| ASFC_Pos | 2,27 |
| ASFN_All | 5,67 |
| ASFN_Neg | 2,93 |
| ASFN_Pos | 2,93 |
| FSC_All | 5,53 |
| FSN_All | 5,07 |
| KFC_All | 5,33 |
| KFC_Neg | 2,67 |
| KFC_Pos | 2,27 |
| KFN_All | 5,67 |
| KFN_Neg | 2,93 |
| KFN_Pos | 2,93 |
| RFPC_Neg | 2,13 |
| RFPC_Pos | 2,60 |
| RFPN_Neg | 3,40 |
| RFPN_Pos | 2,00 |
| TFC-idf_All | 5,60 |
| TFC-idf_Neg | 2,27 |
| TFC-idf_Pos | 3,00 |
| TFN-idf_All | 5,00 |
| TFN-idf_Neg | 3,00 |
| TFN-idf_Pos | 2,80 |
| XSC_All | 6,00 |
| XSN_All | 6,47 |

| Data set | Scoring | Max Scoring | PSO | PSO with search grid |
|------------------------|---------|-------------|-----|----------------------|
| Scop40mini | 50 | 5 | 360 | 600 |
| Chou and Elrod 2003 | 37 | 3 | 200 | 475 |

Table 8 – Estimated running time (hour) for each step method per class.

Finally, it should be distinguished in Table 8, the running time required for SCOP40mini was higher for the Chou and Elrod 2003, and the process with higher execution time was the PSO with search grid. The estimate was based on files created during the process.

5|Results

6 Discussion

In this chapter, it will be discussed the results presented in the previous chapter, including how the proposed method can be influenced, its effectiveness in predicting proteins' class families in both datasets, as well as its major limitations.

6.1 | Proposed Method and its Variables

As for the variables that can influence the proposed method, the obtained results are quite interesting.

Regarding the type of PSO used, PSO-F had better results, meaning in a space of possible features, they were selected with equal weight to best characterize a class. The PSO-W and PSO-WF were created on the idea that each score can assign features that can hold different types of interactions in a protein, and therefore some interactions can be more important than others. But in the results was expressed that every iteration present in each feature must have the same weight as the others.

Regarding the type of MaxScoring, MS-WPS led to better results, in which allows to conclude that the shorter subsequences that are contained in subsequences with higher length will be ignored, so in most cases the longer subsequence has a score value higher than the other subsequences but lower than its parent subsequence (shorter sequence).

So for example a subsequence "aaa" and the subsequence "aaaa", the one with more amino acids is more important despite the score being smaller, which makes sense because the more specific a subsequence is, more information as feature has.

As for different values of M, the best value to be used is 10, since with higher M values can exceed the number of selected features recommended to not occur overfitting. M values lower than 10 can lead to use PSO on the scores in an insignificant way, because by giving reduced search spaces within each score, the PSO will be choosing between sequences and not scores, and what it is desired is selection of the best subsequence in each score obtaining a multi parameter representation of a protein.

About the types frequently used, when comparing both, FC led to better results, making possible to conclude that the interactions between consecutive amino acids are more relevant than those more distant. However, the best results are found when using the FC and the difference between the two frequencies. For some classes was necessary both frequencies in particulars features, and this is similar to what happens in some proteins.

Regarding the number of iterations, this increases when there is more information available, if k or the number of samples in the training set is higher or the search grid is used. And with more necessary iterations more run time is needed, making the method too time consuming. However, the need to increase the number of iterations is normal with the increase of information or more variables present in order to converge to a solution.

These statements are true for most classes but not for all, so it would be interesting to analyze why, and what dataset characteristics that leads to other types of PSO or frequency.

6.2 |Performance

As can be seen in the results, with the proposed method the average AUC value was always higher than the value obtained by other methods based on AAC for the two different data sets. With the proposed method, It was achieved good results regardless of the data set characteristics, such as the percentage of positives in SCOP40mini which is extremely reduced. The other methods on an unbalanced data set failed to attain reasonable results (about 0.5 AUC).

6.3 |Limitations

Despite the favorable results obtained with the proposed method, there are certain characteristics that lead to its disadvantage, and one of them is the scoring, it is the first step and requires a value k. In this work as mentioned above, it was used k equal to 5, which already leads to the calculation of scores for over 3 million combinations for only one class. This leads that chosen number k is conditional to the computing power it is available, and the domains and motifs have more than 5 amino acids.

The biggest limitation is the implementation of the method with the PSO, which needs high computing power because it is a very time consuming iterative method and uses as fitness functions the AUC obtained by training new SVM classifiers each iteration. Because of it, the high M number, or a high number of samples used could lead to a higher running time needed. 6|Discussion

7 | Conclusion

In this chapter, it will be discussed the main achievements of this thesis as well as reflect on the possible applications of this method.

7.1 |Proposed Method

Given the results and their discussion the main objective of this thesis was achieved. It has created a new method for extraction and selection of features from the primary structure of a protein that led to better results than many of the methods mentioned, and also it was successful when applied to two different datasets. Unlike methods based on AAC, this method continued to show good results with date set with a low percentage of positives.

With this method was accomplished a representation of proteins that takes into account various aspects and events that other methods so far had not done, also using the AUC as fitness function and the combination of multiple scores, it was something never done before.

7.2 |Practical Applications

Another positive aspect of this work is that it was not only created a new method, as it was also created an C # application and a website that make all this research practical and able to performed by any user, so the method can be applied easily and appropriately. And also, by having created these tools and making then available it may contribute to validate even better the proposed method because it allows the user to handle all variables adjacent to the method and test it in any dataset.

7.3 |Future Work

Despite the many positive aspects of this thesis, there is still much work to be done so that the using this method can have a significant impact on several areas of interest (medicine, pharmaceutical industry, etc.). This is because this method still has some limitations that can be eliminated.

One of these limitations is the running time of the method. It is crucial to form and plan different options to significantly reduce this execution time without affecting the method performance, in order to properly be used as a research pool.

Another limiting aspect is the definition of the number of iterations that the AUC remains unchanged to stop the iterative PSO, because this is not able to lead to the true convergence of different particles for the best possible AUC value, since this can be obtained before or after the number of iterations set.

Furthermore, it is also necessary to validate this proposed method in different biological data whose structure is also a sequence of coding elements (letters), such as RNA and DNA. This highlights the advantages in using the proposed method, because it does not require anything other than a set with samples consisting of a sequence of letters, and it can be used for different purposes, like the prediction of classes, cell locations, molecular targets, etc., depending of the set objective.

Also in the future, after the resolution of the method's limitations and the search and storage of different optimized SVM classifiers, this research has the objective of building a Multiclass classifier to have a structure that predicts what class a protein belongs and not only if it is part or not of a single set.

In conclusion, all existing sub-sequences in the protein sequence are analyzed by different statistical importance: by their rarity, size, discriminatory power, independence and others. Then they are selected according to these different scores, leading to a significant reduction in the search space as well as a different particle initialization for Particle Swarm Optimization.

This method primes a different analysis of a protein sequence, along with a minor loss of information since this analysis is not only supported with one or two factors. By having such

promising results this method brings new ideas to improve even a better solution to the problem in hands.

7|Conclusion

Appendix

Table 9 - The number and percentage (Rate) of positives in train and test sets for each class considering the number of negatives in SCOP40mini dataset.

| 1216643,0766660,892186612,65176612,513386563,39216573,105396525,64126541,806466526,5956540,767426526,0596541,3689262812,7896281,41910061513,99256173,891011761515,9886171,28117361510,61526173,7712346546,1756550,7614436546,1756550,7615426546,0366550,9116416545,9076551,0617516429,3256443,7118666429,3256443,7624536562,67256403,7625656399,23136401,8926106701,4766710,89276033,93106573,812812460414,57466043,992914060414,5746643,9929140604 </th <th>Class</th> <th></th> <th>Train Set</th> <th></th> <th></th> <th>Test Set</th> <th></th> | Class | | Train Set | | | Test Set | |
|---|-------|-----------|-----------|----------|-----------|-----------|----------|
| 18 661 2,65 17 661 2,11 3 38 656 5,48 6 657 0,90 4 23 656 3,39 21 657 3,10 5 39 652 5,54 12 654 1,80 6 46 652 6,05 9 654 1,36 9 100 615 1,39 25 617 3,81 11 73 615 10,61 52 617 7,77 12 34 654 6,17 5 655 0,76 14 43 654 6,17 5 655 0,76 14 43 654 6,03 6 655 0,91 15 42 654 6,03 6 655 0,91 16 41 654 5,90 7 655 1,66 17 642 9,32 5 | | Positives | Negatives | Rate (%) | Positives | Negatives | Rate (%) |
| 3386565,4866570,904236563,39216573,105396525,64126541,806466526,5956540,767426526,0596541,3689262812,7896281,41910061513,99256173,891011761515,9886171,28117361510,61526177,7712346546,1756550,9114436546,1756550,9115426546,0366550,9116416545,9076551,0617516429,3256440,7719476426,82246443,5920666429,3256440,7719476562,67256401,6921236397,66256417,6923656399,23136401,9926106701,4766710,892710364414,57466443,972812460418,8296041,6729140 </td <td>1</td> <td>21</td> <td>664</td> <td>3,07</td> <td>6</td> <td>666</td> <td>0,89</td> | 1 | 21 | 664 | 3,07 | 6 | 666 | 0,89 |
| 4236563,39216573,105396525,64126541,806466526,5956540,767426526,0596541,3689262812,7896281,41910061513,99256173,891011761515,9886171,28117361510,61526750,7614436546,1756550,7614436546,1756550,7615426546,0366550,9116416545,9076551,6617516427,36206443,5118666429,3256440,7719476426,82246443,5920666429,3256400,7821236397,66256403,7824536399,23136401,98256556399,23136401,9826106701,4766710,892710360414,57466047,982812460418,8296041,472914 | 2 | 18 | 661 | 2,65 | 17 | 661 | 2,51 |
| 5 39 652 5,64 12 654 1,80 6 46 652 6,59 5 654 0,76 7 42 652 6,05 9 654 1,36 8 92 628 12,78 9 628 1,41 9 100 615 13,99 25 617 3,89 10 117 615 15,98 8 617 7,77 12 34 652 4,96 18 653 2,68 13 43 654 6,17 5 655 0,76 14 43 654 6,03 6 655 0,91 16 41 654 5,90 7 655 0,76 17 651 642 9,32 5 644 0,77 18 66 642 9,32 5 644 0,77 19 47 642 6,82 24 644 3,59 20 66 642 9,32 5 644 0,77 21 27 660 3,93 10 660 1,49 22 3 655 | 3 | 38 | 656 | 5,48 | 6 | 657 | 0,90 |
| 64466526,5956540,767426526,0596541,3689262812,7896281,41910061513,99256173,891011761515,9886171,28117361510,61526177,7712346524,96186532,6813436546,1756550,7614436546,0366550,9115426546,0366550,9116416545,9076544,0117516427,36206440,7719476426,82246443,5920666429,3256440,7721226603,93106601,4922346562,67266573,8124536399,23136401,69256436,911,4766710,8926106701,4766710,892710360418,8296043,762812460418,8296043,752914060418,8296043,7531 <t< td=""><td>4</td><td>23</td><td>656</td><td>3,39</td><td>21</td><td>657</td><td>3,10</td></t<> | 4 | 23 | 656 | 3,39 | 21 | 657 | 3,10 |
| 7426526,0596541,3689262812,7896281,41910061513,99256173,891011761515,9886171,28117361510,61526177,7712346524,96186532,6813436546,1756550,7614436546,0366550,9116416545,9076551,0617516427,36206443,01186666429,3256440,7719476426,82246443,59206666429,3256440,7721276603,93106601,4922346562,67266573,8123186562,67266673,8124536399,23136043,97256456399,23136043,9726106701,4766710,89271336041,457466047,08281246041,703256043,97291406041,816156042,4231< | 5 | 39 | 652 | 5,64 | 12 | 654 | 1,80 |
| 89262812,7896281,41910061513,99256173,8910111761515,9886171,28117361510,61526177,7712346524,96186532,6813436546,1756550,7614436546,0366550,9115426546,0366550,9116416545,9076551,0617516427,36206443,0118666429,3256440,7719476426,82246443,5920666429,3256440,7721276603,93106601,4922346562,67266573,8123186562,67266673,8124536399,23136401,7025646429,32136403,9726106701,4766710,8927136441,5156443,97281246454,93106573,81291406041,85296041,4730137 <td>6</td> <td>46</td> <td>652</td> <td>6,59</td> <td>5</td> <td>654</td> <td>0,76</td> | 6 | 46 | 652 | 6,59 | 5 | 654 | 0,76 |
| 910061513,99256173,891011761515,9886171,28117361510,61526177,7712346524,96186532,6813436546,1756550,7614436546,0366550,9115426546,0366550,9116416545,9076551,0617516427,36206443,0118666429,3256440,7719476426,82246443,5920666429,3256440,7721276603,93106601,4922346562,67266573,8123186562,67266573,8124536399,23136401,99256136399,23136403,97291406041,8296041,47301376041,849126043,93311346041,816156042,4233166672,3456690,7434116681,6286701,1835 <td>7</td> <td>42</td> <td>652</td> <td>6,05</td> <td>9</td> <td>654</td> <td>1,36</td> | 7 | 42 | 652 | 6,05 | 9 | 654 | 1,36 |
| 1011761515,9886171,28117361510,61526177,7712346524,96186532,6813436546,1756550,7614436546,0366550,9115426546,0366550,9116416545,9076551,0617516427,36206443,01186666429,3256440,7719476426,82246443,59206666429,3256440,7721276603,93106601,4922346562,67256403,7623186562,67256403,7624536399,23136401,9925656399,23136401,9926106041,88296043,97271336041,816156042,42281246041,816156042,42311346041,816156042,4233166672,3456690,7434116681,6286701,1835 </td <td>8</td> <td>92</td> <td>628</td> <td>12,78</td> <td>9</td> <td>628</td> <td>1,41</td> | 8 | 92 | 628 | 12,78 | 9 | 628 | 1,41 |
| 11 73 615 10,61 52 617 7,77 12 34 652 4,96 18 653 2,68 13 43 654 6,17 5 655 0,76 14 43 654 6,03 6 655 0,91 15 42 654 6,03 6 655 0,91 16 41 654 5,90 7 655 1,06 17 51 642 7,36 20 644 3,01 18 66 642 9,32 5 644 0,77 19 47 642 6,82 24 644 3,59 20 66 642 9,32 5 644 0,77 21 27 660 3,93 10 660 1,49 22 34 656 2,67 26 657 3,81 23 18 656 2,67 26 640 3,96 24 53 639 9, | 9 | 100 | 615 | 13,99 | 25 | 617 | 3,89 |
| 12346524,96186532,6813436546,1756550,7614436546,1756550,7615426546,0366550,9116416545,9076551,0617516427,36206443,01186666429,3256440,7719476426,82246443,59206666429,3256440,7721276603,93106601,4922346562,67266573,8124536397,26256403,76256556399,23136401,9926106701,4766710,892710360418,8296041,473013760418,8296041,473113460418,16156042,4233166672,3456690,7434116681,6286701,1835516557,2856510,7034556557,2856510,70 | 10 | 117 | 615 | 15,98 | 8 | 617 | 1,28 |
| 13436546,1756550,7614436546,1756550,7115426546,0366550,9116416545,9076551,0617516427,36206443,01186666429,3256440,77194776426,82246443,59206666429,3256440,7721276603,93106601,4922346564,93106571,5023186562,67266573,8124536399,23136401,99256156399,23136401,9926106701,4766710,892710360414,57466041,972812460417,03256043,972914060418,8296041,473113460418,16156042,4233166672,3456690,7434116881,6286701,1835516507,2856510,70 | 11 | 73 | 615 | 10,61 | 52 | 617 | 7,77 |
| 14 43 654 6,17 5 655 0,76 15 42 654 6,03 6 655 0,91 16 41 654 5,90 7 655 1,06 17 51 642 7,36 20 644 3,01 18 666 642 9,32 5 644 0,77 19 477 642 6,82 24 644 3,59 20 666 642 9,32 5 644 0,77 21 27 660 3,93 10 660 1,49 22 34 656 2,67 26 657 3,81 24 53 639 9,23 13 640 1,99 25 655 639 9,23 13 604 1,97 28 124 604 17,03 25 604 3,97 29 140 604 18,82 9 604 1,97 31 134 604 | 12 | 34 | 652 | 4,96 | 18 | 653 | 2,68 |
| 15 42 654 6,03 6 655 0,91 16 41 654 5,90 7 655 1,06 17 51 642 7,36 20 644 3,01 18 66 642 9,32 5 644 0,77 19 47 642 6,82 24 644 3,59 20 66 642 9,32 5 644 0,77 21 27 660 3,93 10 660 1,49 22 34 656 4,93 10 657 3,81 23 18 656 2,67 26 657 3,81 24 53 639 9,23 13 640 1,99 25 655 639 9,23 13 640 1,99 26 10 670 1,47 6 671 0,89 27 103 604 14,57 46 604 1,91 32 140 604 18,89 <td>13</td> <td>43</td> <td>654</td> <td>6,17</td> <td>5</td> <td>655</td> <td>0,76</td> | 13 | 43 | 654 | 6,17 | 5 | 655 | 0,76 |
| 16 41 654 5,90 7 655 1,06 17 51 642 7,36 20 644 3,01 18 666 642 9,32 5 644 0,77 19 47 642 6,82 24 644 3,59 20 666 642 9,32 5 644 0,77 21 27 660 3,93 10 660 1,49 22 34 656 2,67 26 657 3,81 24 53 639 7,66 25 640 3,76 25 65 639 9,23 13 640 1,99 26 10 670 1,47 6 671 0,89 27 103 604 14,57 46 604 3,97 28 124 604 17,03 25 604 3,97 29 140 604 18,82 9 604 1,47 30 137 604 | 14 | 43 | 654 | 6,17 | 5 | 655 | 0,76 |
| 17 51 642 7,36 20 644 3,01 18 66 642 9,32 5 644 0,77 19 477 642 6,82 24 644 3,59 20 666 642 9,32 5 644 0,77 21 27 660 3,93 10 660 1,49 22 34 656 4,93 10 657 1,50 23 18 656 2,67 26 657 3,81 24 53 639 9,23 13 640 1,99 26 10 670 1,47 6 671 0,89 27 103 604 14,57 46 604 7,98 28 124 604 17,03 25 604 3,97 29 140 604 18,82 9 604 1,47 30 137 604 18,16 15 604 2,42 31 134 604 | 15 | 42 | 654 | 6,03 | 6 | 655 | 0,91 |
| 18 66 642 9,32 5 644 0,77 19 47 642 6,82 24 644 3,59 20 666 642 9,32 5 644 0,77 21 27 660 3,93 10 660 1,49 22 34 656 4,93 10 657 1,50 23 18 656 2,67 26 657 3,81 24 53 639 9,23 13 640 1,99 26 10 670 1,47 6 671 0,89 27 103 604 14,57 46 604 7,08 28 124 604 17,03 25 604 3,97 39 140 604 18,82 9 604 1,47 30 137 604 18,16 15 604 2,42 31 134 604 <td>16</td> <td>41</td> <td>654</td> <td>5,90</td> <td>7</td> <td>655</td> <td>1,06</td> | 16 | 41 | 654 | 5,90 | 7 | 655 | 1,06 |
| 19476426,82246443,5920666429,3256440,7721276603,93106601,4922346564,93106571,5023186562,67266573,8124536397,66256403,76256556399,23136401,9926106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,49126041,953113460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 17 | 51 | 642 | 7,36 | 20 | 644 | 3,01 |
| 20666429,3256440,7721276603,93106601,4922346564,93106571,5023186562,67266573,8124536397,66256403,76256556399,23136401,9926106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,49126042,423113460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 18 | 66 | 642 | 9,32 | 5 | 644 | 0,77 |
| 21276603,93106601,4922346564,93106571,5023186562,67266573,8124536397,66256403,76256556399,23136401,9926106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,16156042,423113460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 19 | 47 | 642 | 6,82 | 24 | 644 | 3,59 |
| 22346564,93106571,5023186562,67266573,8124536397,66256403,76256556399,23136401,9926106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,16156042,423213460418,16156042,4233166672,3456690,7434116581,6286701,1835516507,2856510,76 | 20 | 66 | 642 | 9,32 | 5 | 644 | 0,77 |
| 23186562,67266573,8124536397,66256403,76256556399,23136401,9926106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 21 | 27 | 660 | 3,93 | 10 | 660 | 1,49 |
| 24536397,66256403,76256556399,23136401,9926106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,49126041,953113460418,16156042,4232166672,3456690,7434116681,6286701,1835516507,2856510,76 | 22 | 34 | 656 | 4,93 | 10 | 657 | 1,50 |
| 25656399,23136401,9926106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,49126041,953113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 23 | 18 | 656 | 2,67 | 26 | 657 | 3,81 |
| 26106701,4766710,892710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,49126041,953113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 24 | 53 | 639 | 7,66 | 25 | 640 | 3,76 |
| 2710360414,57466047,082812460417,03256043,972914060418,8296041,473013760418,49126041,953113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 25 | 65 | 639 | 9,23 | 13 | 640 | 1,99 |
| 2812460417,03256043,972914060418,8296041,473013760418,49126041,953113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 26 | 10 | 670 | 1,47 | 6 | 671 | 0,89 |
| 2914060418,8296041,473013760418,49126041,953113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 27 | 103 | 604 | 14,57 | 46 | 604 | 7,08 |
| 3013760418,49126041,953113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 28 | 124 | 604 | 17,03 | 25 | 604 | 3,97 |
| 3113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 29 | 140 | 604 | 18,82 | 9 | 604 | 1,47 |
| 3113460418,16156042,423213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 30 | 137 | 604 | 18,49 | 12 | 604 | 1,95 |
| 3213460418,16156042,4233166672,3456690,7434116681,6286701,1835516507,2856510,76 | 31 | 134 | 604 | | | 604 | |
| 33166672,3456690,7434116681,6286701,1835516507,2856510,76 | 32 | 134 | 604 | 18,16 | 15 | 604 | 2,42 |
| 34116681,6286701,1835516507,2856510,76 | 33 | 16 | 667 | | | 669 | |
| <i>35</i> 51 650 7,28 5 651 0,76 | | 11 | | | | | |
| | | | | | | | |
| 30 25 050 3,70 31 051 4,55 | 36 | 25 | 650 | 3,70 | 31 | 651 | 4,55 |

| 37 | 161 | 592 | 21,38 | 12 | 592 | 1,99 |
|---------|-------|--------|-------|-------|--------|------|
| 38 | 159 | 592 | 21,17 | 14 | 592 | 2,31 |
| 39 | 159 | 592 | 21,17 | 14 | 592 | 2,31 |
| 40 | 155 | 592 | 20,75 | 18 | 592 | 2,95 |
| 41 | 151 | 592 | 20,32 | 22 | 592 | 3,58 |
| 42 | 166 | 592 | 21,90 | 7 | 592 | 1,17 |
| 43 | 168 | 592 | 22,11 | 5 | 592 | 0,84 |
| 44 | 142 | 592 | 19,35 | 31 | 592 | 4,98 |
| 45 | 166 | 592 | 21,90 | 7 | 592 | 1,17 |
| 46 | 53 | 644 | 7,60 | 16 | 644 | 2,42 |
| 47 | 55 | 644 | 7,87 | 14 | 644 | 2,13 |
| 48 | 63 | 644 | 8,91 | 6 | 644 | 0,92 |
| 49 | 53 | 644 | 7,60 | 16 | 644 | 2,42 |
| 50 | 19 | 663 | 2,79 | 11 | 664 | 1,63 |
| 51 | 32 | 654 | 4,66 | 16 | 655 | 2,38 |
| 52 | 34 | 654 | 4,94 | 14 | 655 | 2,09 |
| 53 | 13 | 668 | 1,91 | 7 | 669 | 1,04 |
| 54 | 13 | 667 | 1,91 | 10 | 667 | 1,48 |
| 55 | 15 | 668 | 2,20 | 5 | 669 | 0,74 |
| Average | 70,93 | 635,62 | 9,80 | 14,02 | 636,44 | 2,15 |
| | | | | | | |

Appendix

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