

# Respiratory issues in patients with multiple sclerosis as a risk factor during SARS-CoV-2 infection: a potential role for exercise

Omid Razi<sup>1</sup> · Ana Maria Teixeira<sup>2</sup> · Bakhtyar Tartibian<sup>3</sup> · Nastaran Zamani<sup>4</sup> · Beat Knechtle<sup>5,6</sup>

Received: 27 January 2022 / Accepted: 4 November 2022 / Published online: 21 November 2022 @ The Author(s) 2022

#### Abstract

Coronavirus disease-2019 (COVID-19) is associated with cytokine storm and is characterized by acute respiratory distress syndrome (ARDS) and pneumonia problems. The respiratory system is a place of inappropriate activation of the immune system in people with multiple sclerosis (MS), and this may cause damage to the lung and worsen both MS and infections. The concerns for patients with multiple sclerosis are because of an enhance risk of infection with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). The MS patients pose challenges in this pandemic situation, because of the regulatory defect of autoreactivity of the immune system and neurological and respiratory tract symptoms. In this review, we first indicate respiratory issues associated with both diseases. Then, the main mechanisms inducing lung damages and also impairing the respiratory muscles in individuals with both diseases is discussed. At the end, the leading role of physical exercise on mitigating respiratory issues inducing mechanisms is meticulously evaluated.

Keywords Multiple sclerosis  $\cdot$  COVID-19  $\cdot$  Exercise training  $\cdot$  Renin–angiotensin system  $\cdot$  Respiratory system  $\cdot$  Immune system

Abbreviations	5
---------------	---

ACE2	Angiotensin-converting enzyme 2
Ang I	Angiotensinogen I
Ang II	Angiotensin II
ARDS	Acute respiratory distress syndrome
AT1R	Angiotensin type 1 receptor
Ca <sup>2+</sup>	Calcium
CNS	Central nervous system
COVID-19	Coronavirus disease-2019
MasR	Mas receptor
MS	Multiple sclerosis

NETs	Neutrophil extracellular traps
NF-ĸB	Nuclear factor-kappa B
NK	Natural killer
RAS	Renin-angiotensin system
ROS	Reactive oxygen species
SARS-CoV-2	Severe acute respiratory syndrome corona-
	virus 2
URTI	Upper respiratory tract infection

- <sup>2</sup> Research Center for Sport and Physical Activity, Faculty of Sport Sciences and Physical Education, University of Coimbra, Coimbra, Portugal
- <sup>3</sup> Department of Exercise Physiology, Faculty of Physical Education and Sports Sciences, Allameh Tabataba'i University, Tehran, Iran
- <sup>4</sup> Department of Biology, Faculty of Science, Payame-Noor University, Tehran, Iran
- <sup>5</sup> Institute of Primary Care, University of Zurich, Zurich, Switzerland
- <sup>6</sup> Medbase St. Gallen Am Vadianplatz, Vadianstrasse 26, 9001 St. Gallen, Switzerland

(	Omid Razi
C	omid.razi.physio@gmail.com

beat.knechtle@hispeed.ch

Beat Knechtle

Ana Maria Teixeira ateixeira@fcdef.uc.pt

Bakhtyar Tartibian ba.tartibian@gmail.com

Nastaran Zamani na\_zamani2000@yahoo.com

<sup>1</sup> Department of Exercise Physiology, Faculty of Physical Education and Sport Sciences, Razi University, Kermanshah, Iran

#### Introduction

The respiratory system, anatomically and functionally, is designed to provide and eliminate oxygen and carbon dioxide (CO<sub>2</sub>), respectively, or simply viewed as a gas exchange system [1, 2]. To do so, the respiratory cycle consists of inspiration and expiration which are performed by the help of several muscles. All components of respiratory system, such as pleurae, airway, and vessels, are innervated by afferent and efferent of autonomic nervous system, sympathetic, and parasympathetic nerves especially the vagal nerve. Breath is an autonomic and rhythmic action that is produced by networks of neurons originating from the brainstem, known as pons and medulla oblongata. These neuronal networks enervate thoracic and abdominal muscles. Three main neuronal groups are involved in monitoring the breath rhythm and its duration: (1) inspiratory neurons in dorsomedial medulla, (2) inspiratory and expiratory neurons in ventrolateral medulla, and (3) inspiratory and expiratory discharging neurons in rostral pons. The important characteristic of this system is its ability to modulate breathing patterns in response to changing of external and internal environments [1, 3, 4].

The majority of components of the respiratory system are impaired in some neurological diseases, such as multiple sclerosis (MS) and Alzheimer disease (AD) [5, 6], and this condition may impose further endangering of these individuals during respiratory virus diseases, like the worldwide coronavirus disease-2019 (COVID-19) pandemic. In this context, physicians most often solicit the use of inhaled steroids and also antibiotic medications [7]. Respiratory pathogenesis of both COVID-19 and MS is extensively referred for improper activation of the immune system, renin-angiotensin system (RAS) dysfunction, the existence of some plaques in brain areas monitoring ventilation skeletal muscles [8–13]. Exercise training as a non-pharmacological intervention by several mechanisms, such as improving the immune responses, converting negative RAS axis to positive one, alleviating the plaque progression, can largely mitigate respiratory issues [14]. Thus, the purposes of this narrative review are to meticulously investigate respiratory issues associated with COVID-19 and MS diseases and also better understand the cellular and molecular mechanisms by which neuro-inflammatory autoimmune disease influences lung immunity. Finally, shed light on the positive roles of regular exercise training as a prophylactic or modifying intervention in mitigating such problems is another outstanding aim of this study.

# Respiratory dysfunctions common road between coronavirus and multiple sclerosis

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the third coronavirus disease originated from

animal and it belongs to beta-coronaviruses which induce a disease known as novel COVID-19 [15, 16]. As inferred naturally from the name of the virus, this disease is associated with respiratory infections [17].

In most cases, the disease is without any respiratory signs; however, all sufferers may later manifest different degrees of lung disorders due to damages in lung tissue [18].

Acute respiratory distress syndrome (ARDS) and pneumonia are the common clinical manifests in patients with severe COVID-19 [18–21]. ARDS is clinical disorder associated with systemic inflammation and failure in multiple organs with a high mortality rate related to lung damage [22, 23]. Hence, COVID-19 is associated with some disorders in lung tissue, including airways, lung parenchyma, lung vessels, and neuromuscular disruptions [24].

This virus can infect several systems, including digestive, genitourinary, central nervous system (CNS), and respiratory systems [16, 25, 26]. During COVID-19 disease, the infected individuals may encounter some respiratory problems occurring orderly through this phases: cellular invasion and viral replication in the nasal cavity, replication in lung and immune system activation, pneumonia, ARDS, cytokine storm, and multi-organ failure [27-29]. Many interactive factors contribute to lung tissue damage and impaired respiratory muscles in both COVID-19 and MS diseases included activated immune system and its pro-inflammatory cytokines such as IFN-Y, TNF- $\alpha$ , and IL-1 $\beta$  [8–10, 14, 30–35], central demyelinated lesions/plaques formed in areas monitoring respiratory rhythm, and muscles induced by the function of the activated immune system [36-46], local, and systemic (soluble) imbalance in the RAS axis [12, 13, 47, 48] (Fig. 1).

Respiratory epithelium, especially ciliary airway epithelium, is the critical point of SARS-CoV-2 entering into the host since it expresses the highest levels of SARS-CoV-2 receptors, namely the angiotensin-converting enzyme 2 (ACE2) [49-51]. Epithelium serves as a barrier against pathogens and particles, preventing tissue damage through secreting mucosa and also mucociliar clearance [24]. Upon cell-virus crosstalk and consequent entering into ciliary nasal cells, SARS-CoV-2 travels to lower respiratory tracts (LRTs) and then triggers the extreme production of inflammatory cytokines and chemokines, such as IL-1, IL-6, IL-8, TNF- $\alpha$ , and - $\beta$ , and monocyte chemoattractant protein 1 (MCP-1). These inflammatory mediators recruit leukocytes to the infectious site [52–54]. Increased cytokine levels can devastate airways and alveolar epithelium by triggering the cells apoptotic process and formation of reactive oxygen species (ROS) exacerbating the pneumonia severity. Alveolar damage remarkably impairs gas exchange and leads to respiratory failure [24, 55, 56]. In more detail, for example, TNF- $\alpha$  has an important role in regulating neutrophils influx following lung damage [23, 57]. Neutrophils release toxic oxygen metabolites such as superoxide anion, hydroxyl



**Fig. 1** The main pathways inducing structural and functional pathogenesis of lung tissue through both MS and COVID-19 diseases. The red arrows represent the detrimental events that result from both diseases causing the whole pulmonary issues indicated in the red rectangular box below. *ACE2* angiotensin-converting enzyme 2; *RAS*  renin–angiotensin system; *ROS* reactive oxygen species; *MS* multiple sclerosis; *URTI* upper respiratory tract infection; *LRTI* lower respiratory tract infection; FEV1/FVC ratio, forced expiratory volume in 1 s to forced vital capacity ration

radicals, and hydrogen peroxide which cause cellular oxidative damage in pulmonary endothelium, parenchymal cells, and inflammatory edema [58–60].

Infiltrated neutrophils, therefore, secret neutrophil extracellular traps (NETs) to control lung infection, but their high production is associated with lung damage by turning the alveolar macrophages into the pro-inflammatory M1 phenotype [61]. The main mechanisms for such transformative phenotype ascribed to the NETs induced activation of signaling pathways in pulmonary cells include extracellular signal-regulated kinases 1/2 (ERK1/2), c-Jun N-terminal kinase (JNK), p38, and nuclear factor-kappa B(NF-κB) proteins [62]. Besides, the proteins and dsDNA components located in NETs may act as critical autoantigen sources to trigger local inflammatory cascades [63]. Of note, infectious and damaged epithelium attracting the pro-inflammatory cytokines is associated with reduced secretion of surfactant proteins of A and B [64], resulting in alveolar collapse. It is important to note that COVID-19 often initiates with symptoms akin to influenza [27]. Coronaviruses are the second reason for induced common cold [65]. As mentioned, SARS-CoV-2 can worsen the conditions of some patients with asthma, since it causes infection of the upper and lower respiratory tracts [65–67]. Epithelium of upper respiratory tract needs 3 weeks to return to the previous normal level [68].

Endothelial dysfunction is another lung pathophysiology of COVID-19 disease. In these patients, extended inflammatory cytokine levels will induce some changes or damages in smooth muscle cells of lung vessels including phenotypic switching from the quiescent contractile phenotype to a proliferative, migratory, and synthetic phenotype which is associated with vessel thickening and also reticular small vessels [69, 70]. It has been reportedly illustrated that endothelial cells suffer apoptosis [71]. Increased permeability of lung vessels is another problem that patients with COVID-19 encounter, which is corroborated by alveolar hemorrhage and fibrin deposition [24]. Thus, these disorders in lung microvessels can impair vascular perfusion [71, 72]. Additionally, regarding the expression of ACE2 on microvascular endothelial cells and vascular smooth muscles, SARS-CoV-2 disrupts the relationship between endothelium and smooth muscles which results in disordered vasodilation and vasoconstriction as well as disorders in gas exchange [73]. It has been documented that some central autoimmune diseases are susceptible to other diseases involving immune system [74, 75]. MS is a chronic central disease characterized by inflammatory demyelination. In both patients with MS and its animal model, experimental autoimmune encephalomyelitis (EAE) is initiated with reactivation of T cells crossing the blood-brain barrier (BBB) into the CNS [76]. Patients with MS have a reduction in clearing virus from their lungs that in part stem from lower efficiency of their anti-viral immune responses [74]. MS patients during contracting respiratory viral infection such as influenza and pneumonia experience higher morbidity and severity than individuals without MS disease [77–80]. Thus, MS disease can exacerbate the expansion of respiratory infection that may partly refer to the regulation of inflammatory characteristics of T cells in MS patient's lungs [43]. Interestingly, it has also been revealed that lung is involved in myelin-reactive T cells becoming pathogenic [43]. As a natural procedure, the mobilization of innate (e.g., natural killer; NK) and acquired immune (CD8 <sup>+</sup>T cells) system is cardinal strategy to control the viral replacing and to clear efficiently the respiratory viruses through releasing an anti-viral pro-inflammatory cytokines, like interferon (IFN)- $\gamma$  [81, 82]. A report documented that EAE animals with respiratory infection lowers the production of effector cells, both innate and acquired, and IFN- $\gamma$ , suggesting a reduction in the immune response to infection in patients with MS [82]. MS and animal model of MS are also associated with mobilizing the extensive population of myeloid-derived suppressor cells (MDSCs), especially their CD11b<sup>+</sup> subunit, from bone marrow, blood, spleen, and CNS into the lungs. These myeloid cells inhibit the proliferation of CD8 <sup>+</sup>T cells and consequently their IFN-y production in lungs [74, 78, 83]. MDSCs use various mechanisms to mitigate the immune response including production of IL-10 and synthesis of nitric oxide (NO) through inducible nitric oxide synthase (iNOS) [83-85]. Therefore, MS patients infected with respiratory viruses present increased viral titers, lung pathology, and consequent increases in their mortality. If the patients with MS survived from respiratory infection, their hospitalization lasted 2 times more than individuals without MS and only infected with respiratory viruses, since patients with MS are exposed to extension of relapses after infection [86–89]. It is also reported that the susceptibility of MS patients to respiratory infections may be elevated during relapsing-remitting MS. It has been suggested that patients with MS during the remission phase show a reduction in their innate immune cells. Of these cells, granulocytes (neutrophil, eosinophil, basophil) are the most important to fight against viral infections [90-93]. As a part of immune response, granulocytes migrate to the infectious site, in this case the lungs, and consequently secret effector molecules,

such as histamine, cytokines, chemokines, enzymes, and growth factors [94, 95]. It has been shown that the number of granulocytes, especially neutrophils, is lower during the remitting phase and that may lead to diminished IFN-y production, a stimulating factor of neutrophils or granulocytes, by Th1 cells [90, 96]. The other pathway that may promote the susceptibility of patients with MS to infectious diseases is immunosenescence, which is associated with progressively diminished number of naïve T cells, originated from structurally and functionally thymic involution [97, 98]. The age range of 20 to 40 is a benchmark age range where the majority of individuals may be afflicted with MS disease and live with this disease for a long time even until death [99]. The events that take place in the immunosenescence process result in poorer immune responses in the old patients with MS [100]. Thymus is a lymphoid organ, where the T cells mature, and is a main source for circulating T cells. The thymic size is progressively elevated until puberty and then undergone involution with its parenchymal tissue replaced by fat [98, 101]. Respiratory viruses are leading causes of acute respiratory infections every year affecting mainly older patients with MS and the elderly. Up to date, several reports have described the association between respiratory viral infections with neurological symptoms [102]. Thus, in MS patients, respiratory viruses have placed themselves as relevant agents responsible for CNS pathologies. Aged MS patients who are in advanced phase of the disease do not have enough CD8<sup>+</sup> T cells in their circulation and consequently in their lung tissue to fight against viral antigens and increased infectious risk [98] exacerbating the neurological signs of the patients [103-106]. Lungs are inflamed during respiratory infection, which is associated with increased upregulation of a chemokine, namely CCL20, to attract Th17 cells into the lungs. Through increased gene expression encoding chemokine receptors and integrin receptors on T cells, these immune cells which converted to the pathogenic phenotype are licensed to enter circulation [43, 107]. Circulating pathogenic T cells then increase BBB permeability and lesion load and volume in brain and spinal cord [108], which is equal to worsening the clinical signs of MS patients.

Reduced physical activity during lockdowns, and especially hospitalization, causes respiratory muscle wasting and impaired skeletal muscles that could lead to sarcopenia and cachexia [109–112]. Using mechanical ventilation for several weeks is another factor involved in structural and functional impairment of respiratory muscles [111, 113, 114]. Also, diaphragm, a key inspiratory muscle, during mechanical ventilation is put in an unloaded condition which can be accompanied with atrophy and consequently weakness. Brainstem centers monitoring respiratory rhythm have been documented to switch off sending efferent impulses to respiratory muscles amid long-term usage of mechanical ventilation [115–117]. In support of this claim, reports disclosed that COVID-19 patients during their stay in intensive care unit (ICU) wards experienced diaphragm impairment and a decrease in its thickness [116, 118]. Also, an atrophy in diaphragm fibers and a reduction in its contractile function have also been reported [119, 120]. MS patients also experience such inactivity which is highly similar to those who are bedridden [121]. Inactivity-induced influence on respiratory muscles may also be ascribed to production of ROS by the pro-inflammatory cytokine storm and activated macrophages and monocytes. Reactive oxygen species and resultant oxidative stress increase the apoptosis and proteolytic processes through the expression of caspase-3 and the activation of the ubiquitin-proteasome system [119, 120, 122-127]. The ubiquitin-proteasome system is activated by hyperinflammation conditions as observed in both COVID-19 and MS diseases [128–130]. The ubiquitin system, which is dependent on ATP, is the main mechanism responsible for muscle atrophy [131, 132]. Pro-inflammatory cytokines induce muscle atrophy, particularly in respiratory muscles, through the following additional mechanisms: inhibited protein synthesis due to the changes in anabolic hormones such as insulin-like growth factor 1 (IGF-1), the mitigating function of satellite cells, attenuated expression of myoblast determining protein 1 (MyoD), downregulation of myosin heavy chain (MHC) of slow twitch fibers and increased degeneration and changes in fiber-type phenotype [133–138], increased activation of NF-kB which leads to the activation of the ubiquitin system [137, 139, 140], and hindered expression of the peroxisome proliferator-activated receptor (PPAR), which has a role in preventing inflammatory conditions, all contributing to a catabolic state along with muscle atrophy [141].

In addition to the immune system, intrinsic expression of ACE2 receptors in the skeletal muscle system may play an important role in SARS-CoV-2 entering into muscles and contribute to skeletal muscle morbidities [142]. Indeed, increased virus entrance into respiratory skeletal muscles is also associated with produced pro-inflammatory cytokines and as a consequence ROS formation. Reactive oxygen species induce muscle damage and atrophy that will finally lead to muscle fatigue [110, 143–147]. These species, further, reduce muscle force production by several mechanisms, including attenuating sensitivity of myofibrils to calcium  $(Ca^{2+})$  [148, 149], oxidizing regulatory proteins of sarcoplasmic reticulum (SR) Ca<sup>2+</sup> release channels [150, 151], opening ryanodine-sensitive Ca<sup>2+</sup> release channel resulting in increased  $Ca^{2+}$  concentration [150, 152], inhibiting the function of sarcoplasmic reticulum calcium ATPase (SERCA) which is necessary for ATP hydrolysis [153], impacting on myofibril structure and function [154, 155], altering cross-bridge kinetics [149], oxidizing myosin heavy chain and also increased impairment of myosin function [154, 156], and modifying the function of troponin C [157].

Thus, increment in ROS can incur in  $Ca^{2+}$  dysregulation in cell cytosol (increased intracellular  $Ca^{2+}$  concentration) that in turn activates calpain [145, 158]. Calpain causes the releasing of sarcomere proteins via cleaving cytoskeletal proteins such as titin and nebulin which are anchored to the contractile components [159]. In this context, however, future studies should address whether the direct attack of SARS-CoV-2 on respiratory muscles has a role in their atrophy.

Severe active respiratory syndrome coronavirus 2 also associates with respiratory challenges after entering the body via respiratory or neuronal pathways. Coronavirus is categorized as a virus that after entering to CNS causes lesions in brainstem, a sensitive area for respiratory cycles [3, 160]. It may be concluded that produced lesions cause a neuromuscular impairment of respiratory muscles. Demyelinated lesions which are observed in patients with COVID-19 and MS diseases are actuated by cytokine storm [18, 33, 36, 37, 161, 162]. Upon entering into the body, SARS-CoV-2 identified as a foreign antigen by immune cells triggers serious immune and inflammatory responses which as a consequence cause extensive peripheral and central release of pro-inflammatory cytokines. There is a positive correlation between increased pro-inflammatory cytokines and disease progression [163, 164]. This process suggests the lack of immune regulation in response to respiratory infection.

That the respiratory system in MS patients can be impaired has been neglected by clinicians and scientists due to prominent other signs in these patients. Altered respiratory function and respiratory muscles strength are changes exacerbated with increasing MS disabilities [165-167], and it has even been disclosed that these respiratory issues account for roughly 47% of total deaths in MS patients [168]. There are acute and chronic respiratory failures in MS patients. Respiratory failure happens in the terminal stages of MS and is usually associated with significant bulbar or limb paralysis [169]. Respiratory failure may be acute, typically secondary to demyelinating lesions in the cervical cord or the medulla, or chronic, typically found in the terminal stages of the disease and related to weak respiratory muscles, and ineffective cough, leading to aspiration, atelectasis and pneumonia. Of the two kinds, only acute respiratory failure is potentially reversible with treatment [169–172]. Weakened respiratory muscles, especially expiratory ones, are a prevalent detriment in advanced phase of MS disease [167, 173, 174]. Paraplegic progression from distal to proximal in MS causes impairment in expiratory muscles prior to the diaphragm and intercostal muscles [175]. The regulation of respiratory muscle function is controlled in the regions of the brain stem and spinal cord, dorsal, and ventral respiratory centers. MS patients have centrally demyelinating plaques extended to these respiratory centers which associate with disrupted impulses and neural pathways related to respiratory muscles [166, 167, 169, 172]. Additionally, the majority of MS patients experience autonomic dysfunction, including in the thermal system, which is originated from lesions in brain stem and medulla areas of the brain [176]. Hyperthermia induced by these lesions negatively influences impulse conduction throughout neurons present in respiratory centers that control respiratory muscles [177, 178]. The primary mechanism that can mechanistically explain such reduction in impulse conduction is attributed to the potassium channels expressed in these neurons. Hyperthermia activates two-pore domain  $K^+$  (K2P) channels on respiratory muscles in neuronal hyperpolarization and reduced action potential propagation [179–181].

The above-mentioned pathways can diminish strength and endurance of respiratory muscles [167, 172-174], more predominant in expiratory respiratory ones [172, 174, 182]. The reduction in these muscle fitness components associate with changes in lung volume and capacity [172], including VC, maximal expiratory and inspiratory pressures, forced expiratory volume in the first second (FEV<sub>1</sub>: the volume of air exhaled in the first second during forced exhalation after maximal inspiration), FVC, FEV<sub>1</sub>/FVC ratio, peak expiratory flow (PEF: the highest forced expiratory flow), and total lung capacity [165, 172, 174]. Collectively, these pulmonary issues in patients with MS engender some abnormalities such as disruption in diffusion capacity of gas dispersed across alveolar membrane, ventilation to perfusion ratio, increased physiological dead space, and consequently diminished oxygenation, inefficient cough, reduced respiratory control, dyspnea, and exercise intolerance or reduced exercise capacity [171, 183–185]. All complications related to respiratory muscle impairment can put MS patients in a severe condition or even death upon infection with COVID-19.

Multiple sclerosis is always associated with some disabilities, including fatigue, strength, coordination, and cognitive signs loss, that progress over time and lead to physical and social inactivity [185]. Besides, several years ago, it has been recommended that MS patients should not participate in physical exercise, just because of increasing their internal temperature during exercise would compromise their clinical signs [186, 187]. Thus, MS patients face a sedentary live [188–190] accompanied with increasing body mass index (BMI) and obesity. Increased BMI and obesity in turn compromise MS severity and even elevate the odds on afflicting MS disease in younger ages [191–195]. Adopted sedentary lifestyle in these patients results in an imbalance between energy intake and expenditure, leading to obesity. Obesity, which is defined as a BMI  $\ge$  30 kg m<sup>-2</sup>, is a metabolic disorder with accumulating fat mass in various body points. Extra burden of body fat through mechanical limitations and reduced thoracic compliance may change lung function/physiology and respiratory rhythm [196, 197]. Hence, increased fat accumulation in areas around ribs, diaphragm, and abdominal cavity implements a mechanical load on chest cavity that abates respiratory compliance (increased stiffness) [198-200]. Elevated intra-abdominal and pleural pressures due to upward and outward movements, respectively, in diaphragm and chest wall preclude airflows toward negative pressure gradient in lungs and pleural space with the lower part of lung system tending to collapse [199, 201, 202]. Generally, increased mechanical load and internal pressures cause a change in respiratory pattern to the quick, shallow type (increased breath rate) [203]. Compromised lung volumes are secondary to the changes of respiratory pattern. The most detrimental alterations in lung volumes and capacities have been observed in expiratory reserve volume (ERV), FVC, forced residual capacity (FRC), total lung capacity (TLC), and tidal volume [204–210]. An impaired lung gas exchange, hypoventilation, and eventually hypoxia have been pinpointed in obese individuals that mostly resulted from regional ventilation-perfusion mismatching; on the other hand, the lower parts of their lungs are often under-ventilated and contrarily overperfused [211, 212]. Adiposity is characterized by deposition of fat in adipocytes, followed by adipocyte hypertrophy and hyperplasia. The hypertrophied adipocyte are infiltrated by macrophages and they in turn release pro-inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ , IL-6) and adipocytokines from the TGF- $\beta$  family, especially TGF- $\beta$ 1 [213–218]. Additionally, an imbalance between some other adipokines, including adiponectin and leptin, also occurs. The concentration of adiponectin, as an anti-inflammatory adipocytokine, and leptin, as a pro-inflammatory cytokine, respectively, decreased and increased in obese individuals [219, 220]. In a more general term, increased secretion of these adipokines into circulation can influence other organs throughout the body and produce some lung disorders, like asthma, COPD, and fibrosis [221, 222]. Increased compensatory lung perfusion in obese individuals can guide circulating TGF-81 to lung tissue. In lungs, TGF- $\beta$ 1 recruits immune cells, such as eosinophils, neutrophils, macrophages, mast cells, and fibroblasts, as well as increases the production and expression of IL-8, cyclooxygenase-2 (COX-2) and prostaglandin E2 (PGE2) in airway smooth muscle cells, leading to airway inflammation and finally asthma [223-227]. Independent of inflammatory responses in airways, TGF- $\beta$ 1 can cause airway remodeling or fibrosis [228]. TGF-β1 modulates the synthetic and secretory functions of epithelial, airway smooth and monocyte cells, and fibroblasts. Increased function of these cells are associated with synthesis and deposition of extracellular matrix (ECM) components. These ECM components include collagen I and IV, elastin, fibronectin, and biglycan [229–231]. Adiposity as a secondary outcome to changing lifestyle in MS patients with detrimental effects on respiratory system can be a risk factor for infectious diseases such as COVID-19 [232] and therefore expose MS patients to higher mortality rate when infected to COVID-19 than non-obese ones. More importantly, accumulated adipose tissue extensively expresses ACE2 enzyme, the receptor for SARS-CoV-2 entering into cells; this tissue thus acting as a reservoir for virus [233, 234].

The infection of SARS-CoV-2 can be fatal and its severity is heterogeneous among individuals that have or do not have underlying diseases [235]. Such heterogeneity in disease severity may be attributed to the underlying diseases that already naturally promote respiratory problems or to differences in ACE2 expression and distribution [236]. Angiotensin-converting enzyme 2 presence in lung tissue can underline the promoted respiratory issues and their severity [237]. Angiotensin-converting enzyme 2 is a dipeptidyl carboxypeptidase expressed remarkably on numerous tissues and organs, including lungs, vascular endothelia, cardiovascular tissue, stomach, small intestine, colon, skin, Ranvier nodes, thymus, bone marrow, spleen, liver, kidneys, and brain [73, 162, 238, 239]. However, ACE belongs to RAS. RAS comprises two arms or axes in which one of them is detrimental/ pathological and another is protective with opposing effects [240, 241]. The main pathological axes are angiotensin II (Ang II also known as Ang1-8)/ACE/angiotensin type 1 receptor (AT1R). Angiotensin-converting enzyme or ACE cleaves angiotensinogen I (Ang I) to form Ang II exerting its actions by binding to AT1R. The protective axis consists of angiotensin 1-7 (Ang1-7)/ACE2/Mas receptor (MasR), and sometimes angiotensin type 2 receptor (AT2R) is also taken into account in this axis. Angiotensin-converting enzyme 2 produces Ang1-7 via catalyzing Ang II [240, 242, 243]. The second axis has anti-inflammatory, anti-proliferative, anti-fibrotic, anti-apoptotic, and vasodilatory functions [244]. There are two RAS types, namely, systemic and local RAS [245]. Indeed, ACE2 is found in two forms, membrane associated and soluble which is catabolically activated [246, 247]. The upregulation of the detrimental axis of RAS has been observed in disease circumstances [248–250], and it has been found that the activity of soluble ACE2 decreases in disease conditions [250]. Soluble and membrane-associated ACE2 as protective axis of RAS were downregulated following infection with SARS-CoV-2 which may contribute to increased viral entering and lysing of ACE2-positive cells [12, 48, 251, 252]. On the other hand, attenuated ACE2 is associated with loss of protective effects of ACE2 and increased Ang II in both mRNA and protein levels [247]. The circulating and tissue levels of ACE2 in some diseases, such as cardiovascular and chronic kidney diseases, and also smokers with chronic obstructive pulmonary disease (COPD) are increased as a compensatory response and it may be an explanation for why some persons with underling diseases are at higher risk of COVID-19-induced mortality

[253, 254]. Increased plasma level of Ang II has been also reported in patients infected with SARS-CoV-2 which had positive correlation to viral load and lung damage. Therefore, it may be possible to inhibit the detrimental effects of COVID-19 through suppressing of Ang II [12, 252, 255, 256]. Angiotensin II or Ang II exerts its pro-inflammatory and pro-fibrotic action through binding AT1R on lung cells [247]. Contrarily, the protective role of the ACE2/Ang1-7/ MasR axis has been identified in several models of lung damage including initial type of SARS [252]. Angiotensinconverting enzyme 2 suppresses the production of Ang II, the activity of ACE and AT1R activation in order to prevent severe lung failure by mediating the production of bioactive peptide of Ang1-7 which activates MasR and AT2R signaling [257-260]. Angiotensin 1-7 promotes its beneficial functions by inhibiting ERK1/2 and natural NF-KB pathways and also prevents bronchial responsiveness, which is a hallmark characteristic of chronic asthma [258, 259]. In MS patients associated with SARS-CoV-2, notable differences were observed in the numbers of lung NK cells, CD8<sup>+</sup> T cells, inflammatory monocytes, and myeloid-derived suppressor cells (MDSCs). This leads to increased lung cell infiltration, suppressive monocytes in the bone marrow, blood, spleen, and CNS, and a decrease in anti-viral CD8<sup>+</sup> T-cell function. It is worth noting that increased concentration of ACE, and dysregulation in ACE/ACE2 balance, has been observed in diseases associated with ARDS, like in patients with severe COVID-19. Produced imbalance favors the detrimental axis of RAS, which can impair lung function due to inflammation, fibrosis, and lung edema, the latter resulting from promoted permeability of lung blood vessels [252, 258, 261, 262].

Majority of the human studies have measured soluble ACE2 in blood, while membrane-associated ACE2 assessment needs more investigation in future. It has been acknowledged that using ACE2 blockers or antibodies disrupting viral entering into the cell during COVID-19 infection, may endanger patients, since these strategies abate the protective effects of ACE2 and its anti-inflammatory activity and as a consequence promote lung susceptibility to damage [263, 264]. Furthermore, utilizing analogue receptors or recombinant soluble ACE2 is another strategy to reduce viral binding in a competitive manner to membrane-associated ACE2 and finally through this procedure mitigate infection and viral load. In this context, soluble ACE2 acts as a decoy receptor and reduces the binding of SARS-CoV-2 to local/membrane-associated ACE2 and as a result reduces lung damages induced by COVID-19 disease [264, 265]. Of note, based on evidence, increased levels of soluble ACE2 point to the attenuation of membrane-associated ACE2 levels [266].

As mentioned above, COVID-19 patients have a lower protective axis compared with controls or rather, patients with COVID-19 disease illustrated higher circulatory Ang II levels which were correlated to viral load [255, 267].

Angiotensin II receptor blockers (ARBs) improve ACE2/Ang1-7/MasR axis of RAS which is associated with assuaged ROS production, inhibiting lung fibrosis via mitigation of collagen deposition, reducing the disruption of alveolar walls through anti-inflammatory influences mediated by suppression of NF- $\kappa$ B pathway, and also by reducing the production of pro-inflammatory cytokines (IL-6, TNF- $\alpha$ ) [268–271]. Collectively, RAS manipulation may abate SARS-induced tissue damages [12].

In any case, details about the expression and distribution of ACE2 receptors are scarce in MS patients and they should be identified in future research. SARS-CoV-2's receptor availability can increase the virus entering into the lung cells and worsen the disease complications [236]. Both ACE2 and transmembrane serine protease 2 (TMPRSS2) are expressed on the apical membrane of alveolar cell type 2 (AT2). The virus binds to the ACE2 receptor through its spike glycoprotein (S) and then TMPRSS2 helps SARS-CoV-2 to fuse with the host cell membrane for the release of its genome [272]. Thus, other factors such as TMPRSS2 may be critical in regulating COVID-19 disease, although it remains to be clarified in future research. As above mentioned, RAS has been found peripherally in circulation and centrally in the CNS [273]. The main sources of RAS components in CNS are glial cells (especially astrocytes) and neurons [274]. Increased expression and activation of detrimental components of RAS have been reported in circulation, cerebrospinal fluid (CSF) and brain tissue (especially on lesions) of MS patients [13, 47, 275], while there was a reduction in the protective ACE2 component [13]. This observed status in MS patients is associated with exacerbating neurological signs [259, 276, 277]. Importantly, the detrimental axis is activated in the early steps of experimental autoimmune encephalomyelitis (EAE), as an animal model of MS, but the protective axis is activated during the end time point of this model [278]. The majority of studies have concentrated on the inflammatory role of the detrimental axis of RAS. Growing scientific literature, using the EAE model, has reported that ACE inhibitors and ARBs and improvement of the protective axis can attenuate the clinical scores and inflammation [243, 279-281]. Thus, RAS axes should be taken into account for therapeutic purposes in the treatment COVID-19.

A correlation has been detected between pulmonary damage and changes in its function [282]. Recovered patients from COVID-19 still experience impairments in pulmonary functional capacity for several months [283]. Such functional disorders or functional reduction have been proved in forced expiratory flow, forced expiratory volume in 1 s to forced vital capacity (FEV1/FVC) [284]. Otherwise, ground-glass opacities are observed in the early and progressive phases of the disease [285]. Patients' age, comorbidities, history of cigarette smoking, the duration of hospital admission, and also the type of medication administration are the critical determinants in the severity of pulmonary disorders [283, 286].

It has been shown that there is a mutual relationship between having chronic respiratory disease and increasing cerebral infraction; on the other hand, it is shown that there is a significant relationship between impaired respiratory function and both brain atrophy and volume of white matter lesions [5, 167, 171, 287].

Remarkable brain and brainstem demyelination influence motor pathways, especially those that innervate limbs, which lead to mobility weakness or impairment. Multiple sclerosis is a neuro-inflammatory and demyelinated disease associated with lesions throughout the CNS, which depending on the involved brain area incurs in some disabilities [288, 289]. Pulmonary dysfunction manifested in MS primarily include impaired respiratory muscles that result in pulmonary weakness and cough. Expiratory muscles are probably more at risk to suffer impairment. It must be mentioned that there is a close correlation between disease severity and higher reductions in respiratory muscles force. In this context, after pulmonary function tests (PFTs) it has been indicated that MS patients have a low vital capacity (14%) in the supine position. Thus, respiratory dysfunction in MS patients may partly reflect the demyelinated lesions in the monitoring area of respiratory centers in the brainstem and cervical spinal cord and as a result, they can weaken the expiratory respiratory muscles. As a whole, impaired expiratory respiratory muscles may be accompanied by a higher risk of respiratory infections like pneumonia. Respiratory infection-induced mortality of MS patients is twice times higher than the general population, [172, 173, 287, 290-293]. It is worth noting that the side effects of some MS-modifying drugs such as Fingolimod, Tranquilizes, muscle relaxants, and opioids may be the main factor in the reduction of some lung function values and slowdown the ventilation action.

It has also been shown that pneumonia is the common consequence in all coronavirus and MS patients. Respiratory problems in MS patients initiate by disease progression. The systemic pro-inflammatory milieu in MS patients alone contributes to skeletal muscle weakness and these complications may increase with COVID-19 infection. In this context, it is necessary to identify the direct attack of SARS-CoV-2 on skeletal muscles in future research.

Additionally, both diseases share the same initial mechanisms and symptoms; thus, individuals with MS may experience and be placed in the intolerable condition after coronavirus infection, just like resurging the chronic relapses increasing the clinical symptoms that may lead to their death [294, 295].

#### Pleiotropic roles of physical exercise

Physical exercise is a challenge on approximately the wholebody system. The movement demands, the control of skeletal muscles, the cardiovascular, and particularly the pulmonary system helps to maintain the intensity of a given exercise for longer times. Persistent contributions of regular exercise, specialty endurance mode causes adaptations in all of these physiological systems [296, 297]. The majority of respiratory muscles including expiratory and inspiratory muscles are skeletal muscles [298]. In admitted COVID-19 patients with a severe condition of mechanical ventilation, it is necessary to strengthen pulmonary muscles during the recovery period [24]. Furthermore, individuals with changes in the motor system and generally with disabilities, most probably experience functionally respiratory disorders [299]. Endurance training might be one recovery strategy to improve the function of pulmonary muscles (Fig. 2). It has been revealed that endurance training primarily increases the number and size of mitochondria and capillaries in skeletal muscles and as a consequence converts the fibers phenotype to the more oxidative type [296, 297, 300-302]. Increased myoglobin and glycogen content and the increase use of fat as a fuel source are other adaptations that occur in skeletal muscles [303, 304]. The main functional alterations in the respiratory system induced by endurance training are as following: (1) increased tidal volume and breath rate which collectively promote maximal pulmonary ventilation and (2) improved pulmonary perfusion as a result of increased pulmonary blood flow in the higher area of the lungs [305]. These adaptations in pulmonary muscles and in respiratory function are partly amenable to increased VO<sub>2max</sub> and lactate thresholds. A study conducted on severe acute respiratory syndrome (SARS) survivals showed that 6 weeks of combined training (endurance and resistance) improved cardiopulmonary and muscle (upper and lower limbs) fitness and performance,



**Fig. 2** The schematic diagram of protective or modifying role of physical activity. The positive effects of regular physical exercise on four body systems is indicated by dashed rectangles and also the positive marks. The effects of positive changes in MS patients are, then, identified by the solid rectangle and also dashed arrows for every item. *FEV* forced expiratory volume; *URTI* upper respiratory tract

infection; *COVID-19* coronavirus disease-19; *QOL* quality of life; Ach, acetylcholine; TLR, toll-like receptor; HPA, hypothalamus–pituitary–adrenal axis; *RAS* renin–angiotensin system; *Treg* T regulatory cells; *OPCs* oligodendrocyte precursor cells; *MBP* myelin basic protein; *PLP* myelin proteolipid protein; *ROS* reactive oxygen species increased predicted VO<sub>2max</sub>, and elevated health-related quality of life (QOL) [306]. Increased VO<sub>2max</sub>/VO<sub>2peak</sub> induced by exercise training mainly comes from improving and reducing blood circulation and pressure, respectively, as well as refining cardiovascular function [307]. Additionally, a reduction in breathlessness and an improvement in muscle endurance and strength can increase contribution to physical exercise and independency in doing personal duties, all promoting QOL [308–310]. Thus, MS patients with the contribution of progressive endurance training following SARS-CoV-2 can expedite their recovery and also improve their quality of life through independence from others in daily tasks [305].

Nowadays, the training of respiratory muscles is the newest training trend to rehabilitate individuals who have a problem with their respiratory muscles or even to enhance performance in persons whose professions benefit from improving the strength of respiratory muscles [311, 312]. This training model is implemented in guise of trained expiratory and inspiratory respiratory muscles, and a combination of them [313]. It has been proved that this type of rehabilitating training in MS patients is involving positive adaptations and improvements in respiratory muscle strength, spirometer parameters, cough efficiency, fatigue, and dyspnea [290, 314–317]. Besides, respiratory training improves the strength and endurance components of respiratory muscles and as a result promote lung functional capacity and performance [318]. Due to enhancement of components related to respiration, such as slowdown breathing rate and assuaged carbon dioxide production, dyspnea is diminished secondary to the respiratory training [317, 319]. Inspiratory muscle training has affirmative effects on cardiac function by involving in autonomic nervous system; for example, increasing parasympathetic activity [320, 321]. Elevated exercisemediated intrathoracic pressure triggers baroreflex activity leading to promoted venous return which in turn mitigates heart sympathetic activation during resting condition [175]. Despite potential influences on respiratory muscles, exercise training defies the cardiac problems incurred in MS and COVID-19 diseases and therefore prevents exacerbating ventilation process. There are several training models escalating respiratory muscle strength and endurance [322–324]. One of these models is swim training [299].

Swim training increases respiratory work; hence, this training type promotes pulmonary volumes by strengthening the respiratory muscles, especially the diaphragm [325].

Other functional changes in the form of adaptations that occurred in the pulmonary system induced by exercise training include (1) reduced fatigability, (2) increased expiratory lung volume, (3) elevated vital capacity, (4) increased diaphragm thickness, (5) enhanced function of inspiratory muscles [326, 327], (6) increased TLC, (7) promoted FRC [299], (8) increased FEV1 [299], (9) promoted FVC, (10) increased PEF [299, 328, 329], and (11) increased strength and endurance of respiratory muscles [330].

Maintenance of diaphragm activity under mechanical ventilation may prevent its atrophy [331]. Otherwise, it has been identified that increased concentration of metabolites in respiratory muscles may partly explain the fatigue of exercising organs; in such a way, metabolites trigger the firing rate of afferent nerves to the autonomous nervous system.

Increased strength of outflow of sympathetic nerve, by corollary, causes vasoconstriction and as result fatigue in exercising organs [332, 333]. Inspiratory and expiratory muscle training inflict a load on the diaphragm and as a result, increases cross-sectional area and strength and endurance of the diaphragm and also improves fatigue tolerance [334–336].

Single exercise sessions, or acute exercise, impact on the immune system by recruiting leukocytes from other organs to circulation, acquiring active phenotype of both innate and adaptive cells including NK cells, active T and B lymphocytes [337, 338], and increased release of immune modulatory peptides, such as anti-inflammatory cytokines [339]. Thus, acute exercise causes the immune activation and this may influence defense mechanisms against pathogens. Although the increased immune function may be efficacious in healthy persons, this condition can aggravate the circumstance of MS individuals particularly those who suffer from COVID-19. It is documented that regular physical exercise can attenuate respiratory issues through effectuating positive responses of the immune system or reducing pro-inflammatory cytokines as causative agents of respiratory issues in COVID-19 and MS patients (Fig. 2) [29, 340, 341]. IL-6 may be one of the outstanding mechanisms by which exercise induces a mitigated inflammatory environment. Exercise training increases the production of IL-6 from adipocytes, macrophages, monocytes, brain, liver, and skeletal muscles [340, 342, 343]. The increased circulatory concentration of IL-6 is associated with attenuated production of pro-inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ ) from inflammatory cells [340, 342] as well as promoted antiinflammatory cytokines, such as IL-1 receptor antagonist (IL-1ra), IL-4, and IL-10 [342, 344]. Furthermore, blockage of IL-1β receptors, which inhibits its signal transduction, maybe another anti-inflammatory function induced by IL-6 [345]. Produced anti-inflammatory cytokines reduce antigen presentation by antigen-presenting cells (APCs) which are necessary to maintain inflammatory responses [346]. The upregulation of IL-6 in lung tissue after exercise training has also been shown in lung injury in animal models [347]. IL-6 dampens pulmonary inflammation through increasing superoxide dismutase (SOD) and also restricts the disruption of alveolar barrier induced by neutrophils [348, 349]. A negative correlation between IL-6 and IL-10 has been shown with neutrophils density in lung tissue. Increased

concentration of IL-6 and also activation of the hypothalamus-pituitary-axis (HPA) induced by physical exercise increase the release of cortisol, a circulatory anti-inflammatory factor. Initially increased exercise-induced cortisol reduces pro-inflammatory production by acting on its own receptors on immune cells [340, 344]. It has also been shown that IL-6 can activate HPA per se [348, 350, 351]. The previous evidence corroborates this claim since an increase in IL-6 receptors and an enlargement have been observed in adrenal glands [350]. Exercise directly mitigates pulmonary inflammation by increasing glucocorticoid receptors on inflammatory lung cells. It also dampens the levels of proinflammatory cytokines in inflammatory lung tissue induced by endotoxin in animal models [347, 348]. Reduced proinflammation plays a critical role in abating the permeability of microvascular endothelium [352] and accordingly reduces ROS and lung edema [62]. Importantly, it has been revealed that enhanced pulmonary antioxidants, particularly SOD, induced by regular exercise can attenuate ARDS produced through viral infection. Enzymatic antioxidants degrade free radicals culminating in the reduction of lung damages [62, 353-355].

Exercise training has an extensive effect on the vague tone of the parasympathetic nerve. The increased efferent reflex of a sympathetic nerve is associated with releasing acetylcholine (Ach) from its terminals. Ach binds to nicotinic receptors on immune cells attenuating the production of pro-inflammatory cytokines as well as acts on macrophages by converting their phenotype from M1 (pro-inflammatory phenotype) to M2 (anti-inflammatory phenotype) [356, 357]. Reduced toll-like receptors (TLRs), especially TLR4, on circulatory monocytes may be another way through which exercise impacts the changes of immune status. Activated intracellular signals of these receptors trigger the production of pro-inflammatory cytokines [358, 359]. Therefore, regular exercise revolves negative immune response to a positive one. Another change in immune function resulting from exercise training is increased circulatory number of T regulatory (Treg) cells [360]. These cells secret anti-inflammatory cytokines like IL-10 and transforming growth factorbeta (TGF- $\beta$ ) and also increase the proportion of Th2 to Th1 which is related to promoting anti-inflammatory cytokines [361]. There is cross-reactivity between pro-inflammatory cytokines and microglial cells; in such a way, reduced proinflammatory cytokines induced by exercise training, mitigates reactivated microglia (microgliosis) and consequently decreased microgliosis associated with assuaging the produced pro-inflammatory cytokines released by reactive microglia [362, 363]. The changes of detrimental immune responses to reparative/positive responses may be efficacious to mitigate pulmonary damages resulting from COVID-19 disease and to improve the strength and endurance of the respiratory muscles in MS patients infected with COVID-19.

Importantly, attenuated pro-inflammatory cytokines provided by physical exercise can also be beneficial for reducing neuronal loss and for reducing the demyelination induced by MS/COVID-19 in brain areas monitoring respiratory muscles and ventilation cycle [32, 364, 365]. Thus, exercise training establishes an appropriate balance in lung infection, tissue homeostasis, and immune response.

It has been found that exercise inhibits alveolar macrophages polarization to pro-inflammatory M1 phenotype by reducing NETs production and suppressing ERK1/2 and NF- $\kappa$ B pathways in lung cells and macrophages. This action can culminate in the mitigation of lung damages [366]. Besides, exercise enhances sputum clearance throughout the pulmonary system, which can be attributed to increased activity of nasal epithelial sodium channels (ENaC), promoted ventilation, shear force, and body movements [367]. Damped neutrophilic inflammation has been reported in individuals with pulmonary problems after participation in regular exercise programs and is associated with the diminishment of complement receptors [368]. Thus, exercise could reduce lung inflammation induced by infection, particularly in patients with MS.

Increased remyelination, or rather, ceased demyelination mediated by regular physical exercise could be attributed to the following: (1) increased central expression of neurotrophic factors and their receptors expressed in brain areas, particularly on oligodendrocyte precursor cells (OPCs), which can elevate the proliferation and differentiation of OPCs to adult (myelinating) oligodendrocytes enveloping neural axon [369, 370], (2) increased number of mitochondria, which is associated with mitigating the production of pro-inflammatory cytokines, reduces myelin damage induced by oxidative stress [371], (3) increased antioxidant enzymes [372], (4) upregulation of some myelin protein expression, such as myelin main protein (MBP) and proteolipid protein (PLP) [373], which are expressed on myelin sheath and also essential for myelin formation and thickness [374, 375], and (5) phenotypic conversion of microglia from M1 (pro-inflammatory) to M2 (anti-inflammatory) type and maintain them in inactivation or resting state as well as increasing their phagocytic function for expediting clearance of debris [376, 377]. Collectively, contribution in regular physical exercise can preclude plaque/lesion extension to the areas of the brain more related to respiratory centers and even restore nerve impulses through remyelinating processes. The relationship between immune and pulmonary systems in MS individuals with COVID-19 disease is per se complex and there is no information related to exercise training and respiratory system in MS patients who have been infected with COVID-19.

It has been postulated that low-to-moderate-intensity exercise in contrast to high-intensity exercise, causes a decrement in upper respiratory tract infections (URTI) and symptoms [378]. Besides, the individuals with moderate exercise levels also experience lower URTI incidence compared to their sedentary counterparts [379]. The main mechanisms related to reducing URTI induced by moderate regular exercise training have been attributed to the following (Fig. 2): first, increased salivary immunoglobulin A (s-IgA) which is the first line of the body defense against foreign pathogens, like respiratory viruses. This factor binds to respiratory viruses and eliminates them through opsonization [380, 381]. Second, immune phenotype changes from T helper 1 (Th1) to Th2 (improving Th1/Th2 balance). Th1 cells produce pro-inflammatory chemokines when exposed to pathogens, but their excessive responses can incur in tissue damages in the lungs [382]. In this context, moderate exercise training attenuates immune cells infiltration to lungs and lymph nodes drainage and release of pro-inflammatory cytokines by Th1 are reduced [383]. Third, increased IL-2 levels in lung tissue enhance differentiation and maturation of Treg cells. Increased number of Treg is congruent with establishing an anti-inflammatory milieu in lungs [382]. Anti-inflammatory cytokines such as IL-4 exert another role in reducing detrimental pro-inflammatory conditions. Interleukin-4 facilitates the differentiation of naïve Th to Th2 phenotype which has an anti-inflammatory function as well as co-stimulates B cells to secrete virus-neutralizing antibodies. Viral antibodies reduce the virus load through inhibiting the infection of cells and opsonizing the infected cells [383]. Fourth, increased soluble TNF- $\alpha$  receptor which is capable to bind to circulatory TNF- $\alpha$  through which mitigates membrane-binding propensity and consequently reduces activation of NF-kB signal pathways. Fifth, an increment in eosinophil chemoattractants causes extravasation of eosinophils into the infected lung tissue where their ribonucleases can degrade virus's single-stranded RNA and suppresses virus replication [383]. Initial increases in cortisol induced by chronic exercise may act as an assuaging factor of pro-inflammatory condition produced by infection and as a consequence reduces lung susceptibility to infection [383, 384]. In this matter, professionals should be aware that prescribing a proper exercise protocol in MS patients with SARS-CoV-2 infection is essential, since higher core body temperature (hyperthermia) in individuals with MS may act as an endogenous stress factor that causes a higher CNS recruitment and higher exertion. In this case, higher exertion will lead to increased concentration of stress hormones and result in impairment of the host immune system which it may endanger MS patients with compromised immune system [385, 386]. Sixth, exercise increases the circulation of IL-6 derived from exercising skeletal muscles and, it, in turn, upregulates anti-inflammatory cytokines, including IL-1ra and IL-10. These anti-inflammatory cytokines mitigate the extended inflammation originated from respiratory virus infection [383, 387]. Besides, increased recruitment of NK and cytotoxic T cells also occurs following regular exercise training, improving immune defense against foreign pathogens [388, 389]. Exercise-mediated increases in immunosurveillance and attenuated inflammation have been observed in some parts of the body, including the upper respiratory tract (URT), lung, blood, and skeletal muscles, among others [389, 390]. Thus, regarding a reverse relationship between mediated exercise training and URTI incidence and duration [391, 392] and also fatality and pneumonia rates [393–395], either individuals with a clinical condition or healthy are encouraged to regularly practice physical exercise. It is worth noting that highly fitted persons have lower basic levels of inflammatory biomarkers compared with unfitted ones [396].

As mentioned, host susceptibility to SARS-CoV-2 is dependent on binding between host ACE2 and spike (S) glycoprotein of the virus which is known as the S1 subunit [397]. Although there are not enough reports regarding exercise training on ACE, especially ACE2 as local or lung tissue receptor, the changes of other subunits in other organ systems like kidneys, heart, brain, skeletal muscles, and circulation mediated by exercise are available [398, 399]. The most beneficial and prophylactic effects of exercise maybe induced through changes in RAS (Fig. 2) [400, 401]. Based on a literature review and recently original reports, regular exercise downregulates systemic and local ACE/ AngII/AT1R axis and also upregulates all components of ACE2/Ang1-7/MasR axis, as well as transfers the axis balance to the protective axis [400, 402, 403]. Upregulated protective axis of RAS increases the bioavailability of prostaglandins (PGs) and bradykinin as well as enhances anti-inflammatory environment, augments anti-fibrotic and antioxidant defenses, and normalizes oxidative stress and anti-apoptotic environment [404-406]. These responses in RAS can improve lung blood flow and consequently lead to reduced oxygen deficiency in MS patients infected with COVID-19 [407, 408]. Besides, it has been claimed that exercise training reduces lung lesions and fibrosis through the normalization of RAS axes and reducing collagen deposition [271, 399, 409]. By affecting this system, exercise training can attenuate the susceptibility of individuals to detrimental functions of COVID-19 infection or mitigate the severity of disease by the following additional strategies: (1) mitigated severity of comorbidities [247] and as a result reduced COVID-19-induced mortality rates [410, 411] and (2) warded off the diminishing effects of COVID-19 on ACE2 via increasing ACE2 activity and its concentration [412], although the positive or negative effects of increasing ACE2 should be investigated. Thus, RAS manipulation and its normalization may be a potential treatment for health optimization against the COVID-19 pandemic.

Since adipose tissue can play a role as the viral reservoir [233] and in the sense that obesity causes many structural

and functional issues in respiratory system, weight loss via lifestyle changes may reverse such respiratory problems [413]. Physical exercise has profound effects on body composition by increasing fat oxidation and improving muscle mass, which has a leading role on fat oxidation and consequent weight loss. By the same token, exercise should have enough intensity to influence lipid oxidation and metabolic factors [414]. There are several pathways by which exercise causes weight loss, including increased aerobic capacity measured by maximal oxygen consumption (VO2 max) and altered body composition resulting in part from elevating muscle mass [415–417]. Promoted muscle mass is associated with more consumption of glucose and lipid as fuels and as a result dampens insulin resistance [418, 419]. In addition, increasing activation of AMP-activated protein kinase (AMPK) and peroxisome proliferator-activated receptor gamma coactivator 1 (PGC)- $\alpha$  is another mechanism through which exercise facilitates lipid and glucose oxidation [420, 421]. PGC- $\alpha$  increases aerobic capacity of muscle tissue by impacting on mitochondrial biogenesis [422, 423]. Besides, changes in some genes involving in lipogenesis and lipolysis are another adaptation that occurs during and after exercise. In this context, it has been disclosed that lipolysis [peroxisome proliferator-activated receptor (PPAR)-α, cytochrome c oxidase (COX) IV] and lipogenesis [fatty acid synthase (FAS), and acetyl-CoA carboxylase (ACC)] genes are upregulated and downregulated, respectively [424]. The initiation of exercise elevates catecholamine hormones, including adrenaline and noradrenaline. Upon release, these hormones bind to their  $\beta$ -adrenergic receptors expressed on adipose tissue yielding intracellular signal and consequent phosphorylation of hormone-sensitive lipase to promote lipolysis in this tissue [425]. It is worth noting that exercise reverses increased adiponectin induced by obesity. Elevated anti-inflammatory adipokine increases the expression of farnesoid X receptor (FXR) as a regulator of multiple metabolic pathways. FXR then activates adaptor phosphotyrosine protein interacting with the PH domain and leucine zipper 1 (APPL1) to increase lipolysis [426, 427]. Additionally, physical exercise establishes a balance among some adipomyokines, such as myostatin (MST), TGF- $\beta$ 1, and activin A, as members of the transforming growth factor- $\beta$  superfamily  $(TGF-\beta)$  and follistatin (FST). These adipo-myokines, particularly TGF-β members, are upregulated in adiposity and inflammatory condition, while FST inhibits their function through binding to them. Generally, FST increases muscle mass and consequently reduces body fat [428-432]. Therefore, physical exercise is a dynamic lifestyle that mitigates weight gain or obesity, as a risk factor for severe COVID-19, in MS patients and as a result, reverses the changes in lung mechanics and function. It has been suggested that weight loss associates with improving in peak expiratory flow and some spirometer indices [433–435] markedly increases in lung volumes (TLC, FRC, ERV) [436–438], diminishing airway hyper-responsiveness in asthmatic and non-asthmatic obese individuals [434, 439, 440].

As mentioned in the previous section, increased core body temperature in MS patients can influence the respiratory center and nerves in the brain monitoring respiratory muscles and ventilation rhythm. Thus, improving heat strain engendered in MS patients during coronavirus infection, as a febrile virus, would help MS patients to reduce the detrimental effects of hyperthermia on respiratory muscles, especially their fatigability [441]. Although physical exercise is notorious as a heat stressor, long-term exposure to physical exercise is associated with some adaptations in thermal regulation to diminish its compromised effects [442]. Exercise training causes adaptive changes in the cardiovascular system and hemodynamic and hematological factors, including increased contractile strength of cardiac muscle, increased plasma volume, and reduced vasoconstriction at the subcutaneous level [443-445]. These adaptations are associated with supplying deep or core organs with higher cardiac output and followed by transferring the core temperature to the body surface [446, 447]. Exercise increases antioxidant enzymes and therefore reduces and elevates reactive oxygen species (ROS) production and nitric oxide (NO) bioavailability, respectively [448-451]. Besides, increased plasma ATP concentration in response to exercise-induced hypoxia and shear stress, interacts with P2Y receptors to elevate the vasodilation factors, such as NO and prostaglandin E2 (PGE2). These exercise-induced alterations attenuate vascular damages and promote microvessel dilation [452–460]. Some other adaptive mechanisms yielded by exercise amenable to dampening core body temperature are increased sweat rate through elevating cholinergic sensitivity, higher efficiency of eccrine sweat gland in sweat production per each gland, increased number and sensitivity of muscarinic receptors responsible for sweating [442, 461]. Therefore, exercise abates the threshold for commencing subcutaneous blood flow and sweat production in response to promoting core body temperature. Generally speaking, maintaining core body temperature in a narrative range mediated by exercise can preserve the normal impulses along neurons enervating respiratory muscles, followed by the attenuation of clinical signs and premature whole and respiratory fatigue in MS patients.

#### Conclusion

Our review investigated molecular mechanisms of respiratory impairments and lung damage in MS patients with COVID-19. We found that regular exercise training changes the responses of the immune system and also increases some aspects of innate and adaptive immunity against SARS-CoV-2 virus to cope with lung damages. Generally speaking, physical exercise training can mitigate the negative effects of COVID-19 disease on lung tissue and respiratory muscles in MS patients and expedites their recovery following COVID-19 infection.

Acknowledgements Not applicable.

Author contributions OR, BT, and NZ conceptualized and wrote the first draft. AMT, BK, and OR developed the study concept. BT, AMT, and BK reviewed and edited the final version of manuscript. All authors contributed to the article and approved the submitted version.

**Funding** Open access funding provided by University of Zurich. The authors declare that no funds, grants, or other support were received during the preparation of this manuscript.

**Data availability** Data sharing not applicable to this article as no datasets were generated or analyzed during the current study.

### Declarations

**Conflict of interest** The authors have no competing interests to declare that are relevant to the content of this article.

Ethical approval No applicable.

Consent to participate No applicable.

**Consent to publication** Authors consent for the publication of the manuscript.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

## References

- Person A, Mintz ML (2006) Anatomy and physiology of the respiratory tract. Springer, Disorders of the respiratory tract, pp 11–15
- 2. Ward JP, Ward J, Leach RM (2010) The respiratory system at a glance. John Wiley & Sons
- 3. Waugh A, Grant A (2014) Ross & Wilson anatomy and physiology in health and illness E-book. Elsevier health sciences, Amsterdam
- Credland N (2016) Respiratory anatomy and physiology. Routledge, Respiratory care, pp 15–28
- Buyse B, Demedts M, Meekers J, Vandegaer L, Rochette F, Kerkhofs L (1997) Respiratory dysfunction in multiple sclerosis: a prospective analysis of 60 patients. Eur Respir J 10:139–145

- Smeltzer SC, Utell MJ, Rudick RA, Herndon RM (1988) Pulmonary function and dysfunction in multiple sclerosis. Arch Neurol 45:1245–1249
- 7. Ronsen O (2005) Prevention and management of respiratory tract infections in athletes. New Stud Athl 20:49
- Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ (2020) COVID-19: consider cytokine storm syndromes and immunosuppression. Lancet 395:1033–1034
- Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, Wang T, Zhang X, Chen H, Yu H (2020) Clinical and immunological features of severe and moderate coronavirus disease 2019. J Clin Investig 130:2620–2629
- Link H (1998) The cytokine storm in multiple sclerosis. Mult Scler J 4:12–15
- 11. Killestein J, Rep MH, Barkhof F, Roos MT, Adèr HJ, van Lier RA, Polman CH (2001) Active MRI lesion appearance in MS patients is preceded by fluctuations in circulating T-helper 1 and 2 cells. J Neuroimmunol 118:286–294
- Kuba K, Imai Y, Rao S, Gao H, Guo F, Guan B, Huan Y, Yang P, Zhang Y, Deng W (2005) A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. Nat Med 11:875–879
- Kawajiri M, Mogi M, Higaki N, Matsuoka T, Ohyagi Y, Tsukuda K, Kohara K, Horiuchi M, Miki T, Kira J (2009) Angiotensin-converting enzyme (ACE) and ACE2 levels in the cerebrospinal fluid of patients with multiple sclerosis. Mult Scler J 15:262–265
- 14. Pedersen BK, Saltin B (2006) Evidence for prescribing exercise as therapy in chronic disease. Scand J Med Sci Sports 16:3–63
- Chen Y, Liu Q, Guo D (2020) Emerging coronaviruses: genome structure, replication, and pathogenesis. J Med Virol 92:418–423
- Maleki BH, Tartibian B (2021) COVID-19 and male reproductive function: a prospective, longitudinal cohort study. Reproduction 161:319–331
- Rahimi B, Vesal A, Edalatifard M (2020) Coronavirus and Its effect on the respiratory system: is there any association between pneumonia and immune cells. J Fam Med Prim Care 9:4729
- Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X (2020) Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. The Lancet 395:497–506
- Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, Qiu Y, Wang J, Liu Y, Wei Y (2020) Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet 395:507–513
- 20. Wu C, Chen X, Cai Y et al (2020) Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 pneumonia in Wuhan, China: JAMA intern med. J Emerg Med 58:713
- Wu F, Zhao S, Yu B, Chen YM, Wang W, Song ZG, Hu Y, Tao ZW, Tian JH, Pei YY (2020) A new coronavirus associated with human respiratory disease in China. Nature 579:265–269
- 22. Zilberberg MD, Epstein SK (1998) Acute lung injury in the medical ICU: comorbid conditions, age, etiology, and hospital outcome. Am J Respir Crit Care Med 157:1159–1164
- Krishnadasan B, Naidu BV, Byrne K, Fraga C, Verrier ED, Mulligan MS (2003) The role of proinflammatory cytokines in lung ischemia-reperfusion injury. J Thorac Cardiovasc Surg 125:261–272
- Brosnahan SB, Jonkman AH, Kugler MC, Munger JS, Kaufman DA (2020) COVID-19 and respiratory system disorders: current knowledge, future clinical and translational research questions. Arterioscler Thromb Vasc Biol 40:2586–2597
- 25. Zou X, Chen K, Zou J, Han P, Hao J, Han Z (2020) Singlecell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable

- Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W (2020) Detection of SARS-CoV-2 in different types of clinical specimens. JAMA 323:1843–1844
- Biscardi A (2020) Coronavirus impacts on respiratory system and its phases. In: Biscardi A (ed) Medical reports and case studies. Bologna, Italy
- Yufang S, Ying W, Changsun S (2020) COVID-19 infection: the perspective on immune response. Cell Death Differ. https://doi. org/10.1038/s41418-020-0530-3
- Chowdhury MA, Hossain N, Kashem MA, Shahid MA, Alam A (2020) Immune response in COVID-19: a review. J Infect Public Health. https://doi.org/10.1016/j.jiph.2020.07.001
- Chen C, Zhang X, Ju Z, He W (2020) Research progress on the mechanism of cytokine storm induced by new coronavirus pneumonia and related immunotherapy [J/OL]. Chinese J Burns 36:E005
- QinC Z (2020) Dysregulation of immune response in patients with COVID-19 in Wuhan China. Clin Infect Dis 71(15):762– 768. https://doi.org/10.1093/cid/ciaa248
- 32. Li Y, Li H, Fan R, Wen B, Zhang J, Cao X, Wang C, Song Z, Li S, Li X (2016) Coronavirus infections in the central nervous system and respiratory tract show distinct features in hospitalized children. Intervirology 59:163–169
- 33. Sorenson M, Furst J, Mathews H, Jason LA (2017) Dysregulation of cytokine pathways in chronic fatigue syndrome and multiple sclerosis. In: Sorenson M (ed) Fatigue: biomedicine health & behavior
- 34. Killestein J, Den Drijver B, Van der Graaff W, Uitdehaag BM, Polman C, Van Lier RA (2001) Intracellular cytokine profile in T-cell subsets of multiple sclerosis patients: different features in primary progressive disease. Mult Scler J 7:145–150
- 35. Wygrecka M, Jablonska E, Guenther A, Preissner KT, Markart P (2008) Current view on alveolar coagulation and fibrinolysis in acute inflammatory and chronic interstitial lung diseases. Thromb Haemost 99:494–501
- 36. Smith KJ, McDonald W (1999) The pathophysiology of multiple sclerosis: the mechanisms underlying the production of symptoms and the natural history of the disease. Philosophical transactions of the royal society of London. Series B 354:1649–1673
- Flachenecker P, Bihler I, Weber F, Gottschalk M, Toyka KV, Rieckmann P (2004) Cytokine mRNA expression in patients with multiple sclerosis and fatigue. Mult Scler J 10:165–169
- Netland J, Meyerholz DK, Moore S, Cassell M, Perlman S (2008) Severe acute respiratory syndrome coronavirus infection causes neuronal death in the absence of encephalitis in mice transgenic for human ACE2. J Virol 82:7264–7275
- Gu J, Korteweg C (2007) Pathology and pathogenesis of severe acute respiratory syndrome. Am J Pathol 170:1136–1147
- Mao X-Y, Jin W-L (2020) The COVID-19 pandemic: consideration for brain infection. Neuroscience 437:130
- Li YC, Bai WZ, Hashikawa T (2020) Response to commentary on "the neuroinvasive potential of SARS-CoV-2 may play a role in the respiratory failure of COVID19 patients." J Med Virol 92(7):707–709
- 42. Desforges M, Le Coupanec A, Dubeau P, Bourgouin A, Lajoie L, Dubé M, Talbot PJ (2020) Human coronaviruses and other respiratory viruses: underestimated opportunistic pathogens of the central nervous system? Viruses 12:14
- 43. Odoardi F, Sie C, Streyl K, Ulaganathan VK, Schläger C, Lodygin D, Heckelsmiller K, Nietfeld W, Ellwart J, Klinkert WE (2012) T cells become licensed in the lung to enter the central nervous system. Nature 488:675–679

- Steelman AJ (2015) Infection as an environmental trigger of multiple sclerosis disease exacerbation. Front Immunol 6:520
- 45. Baig AM, Khaleeq A, Ali U, Syeda H (2020) Evidence of the COVID-19 virus targeting the CNS: tissue distribution, hostvirus interaction, and proposed neurotropic mechanisms. ACS Chem Neurosci 11:995–998
- 46. Li Y, Bai W, Hashikawa T (2020) The neuroinvasive potential of SARS-CoV2 may be at least partially responsible for the respiratory failure of patients with COVID-19. J Med Virol 92(6):552–555
- Mogi M, Horiuchi M (2013) Effect of angiotensin II type 2 receptor on stroke, cognitive impairment and neurodegenerative diseases. Geriatr Gerontol Int 13:13–18
- Imai Y, Kuba K, Penninger JM (2008) The discovery of angiotensin-converting enzyme 2 and its role in acute lung injury in mice. Exp Physiol 93:543–548
- Astuti I (2020) Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2): an overview of viral structure and host response. Diabetes Metab Syndr 14:407–412
- Shereen MA, Khan S, Kazmi A, Bashir N, Siddique R (2020) COVID-19 infection: origin, transmission, and characteristics of human coronaviruses. J Adv Res 24:91–98
- 51. Sungnak W, Huang N, Bécavin C, Berg M, Queen R, Litvinukova M, Talavera-López C, Maatz H, Reichart D, Sampaziotis F (2020) SARS-CoV-2 entry factors are highly expressed in nasal epithelial cells together with innate immune genes. Nat Med 26:681–687
- Li G, Fan Y, Lai Y, Han T, Li Z, Zhou P, Pan P, Wang W, Hu D, Liu X (2020) Coronavirus infections and immune responses. J Med Virol 92:424–432
- Mason RJ (2020) Pathogenesis of COVID-19 from a cell biology perspective. Eur Respir J. https://doi.org/10.1183/13993003. 00607-2020
- 54. Tang NLS, Chan PKS, Wong CK, To KF, Wu AKL, Sung YM, Hui DSC, Sung JJY, Lam CWK (2005) Early enhanced expression of interferon-inducible protein-10 (CXCL-10) and other chemokines predicts adverse outcome in severe acute respiratory syndrome. Clin Chem 51:2333–2340
- 55. Chan JFW, Zhang AJ, Yuan S, Poon VKM, Chan CCS, Lee ACY, Chan WM, Fan Z, Tsoi HW, Wen L (2020) Simulation of the clinical and pathological manifestations of coronavirus disease 2019 (COVID-19) in a golden Syrian hamster model: implications for disease pathogenesis and transmissibility. Clin Infect Dis 71:2428–2446
- 56. Lim YX, Ng YL, Tam JP, Liu DX (2016) Human coronaviruses: a review of virus-host interactions. Diseases 4:26
- 57. Naidu BV, Woolley SM, Farivar AS, Thomas R, Fraga CH, Goss CH, Mulligan MS (2004) Early tumor necrosis factor-α release from the pulmonary macrophage in lung ischemia-reperfusion injury. J Thorac Cardiovasc Surg 127:1502–1508
- Splettstoesser WD, Schuff-Werner P (2002) Oxidative stress in phagocytes—"the enemy within." Microsc Res Tech 57:441–455
- Chandel NS, Schumacker PT, Arch RH (2001) Reactive oxygen species are downstream products of TRAF-mediated signal transduction. J Biol Chem 276:42728–42736
- Li J-M, Fan LM, Christie MR, Shah AM (2005) Acute tumor necrosis factor alpha signaling via NADPH oxidase in microvascular endothelial cells: role of p47phox phosphorylation and binding to TRAF4. Mol Cell Biol 25:2320–2330
- Tomar B, Anders H-J, Desai J, Mulay SR (2020) Neutrophils and neutrophil extracellular traps drive necroinflammation in COVID-19. Cells 9:1383
- 62. Mussi R, Camargo E, Ferreira T, De Moraes C, Delbin M, Toro I, Brancher S, Landucci E, Zanesco A, Antunes E (2008) Exercise training reduces pulmonary ischaemia–reperfusion-induced inflammatory responses. Eur Respir J 31:645–649

- De Perrot M, Liu M, Waddell TK, Keshavjee S (2003) Ischemia– reperfusion–induced lung injury. Am J Respir Crit Care Med 167:490–511
- 64. Wang J, Nikrad MP, Phang T, Gao B, Alford T, Ito Y, Edeen K, Travanty EA, Kosmider B, Hartshorn K (2011) Innate immune response to influenza A virus in differentiated human alveolar type II cells. Am J Respir Cell Mol Biol 45:582–591
- Chilvers M, McKean M, Rutman A, Myint B, Silverman M, O'Callaghan C (2001) The effects of coronavirus on human nasal ciliated respiratory epithelium. Eur Respir J 18:965–970
- 66. Johnston SL, Pattemore PK, Sanderson G, Smith S, Lampe F, Josephs L, Symington P, O'Toole S, Myint SH, Tyrrell DA (1995) Community study of role of viral infections in exacerbations of asthma in 9–11 year old children. BMJ 310:1225–1229
- 67. Pedersen M, Sakakura Y, Winther B, Brofeldt S, Mygind N (1983) Nasal mucociliary transport, number of ciliated cells, and beating pattern in naturally acquired common colds. Eur J Respir Dis Suppl 128:355–365
- Rautiainen M, Kiukaanniemi H, Nuutinen J, Collan Y (1992) Ultrastructural changes in human nasal cilia caused by the common cold and recovery of ciliated epithelium. Ann Otolo Rhinol Laryngol 101:982–987
- 69. Bai HX, Wang R, Xiong Z, Hsieh B, Chang K, Halsey K, Tran TML, Choi JW, Wang D-C, Shi L-B (2020) Artificial intelligence augmentation of radiologist performance in distinguishing COVID-19 from pneumonia of other origin at chest CT. Radiology 296:E156–E165
- Magro C, Mulvey JJ, Berlin D, Nuovo G, Salvatore S, Harp J, Baxter-Stoltzfus A, Laurence J (2020) Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: a report of five cases. Transl Res 220:1–13
- Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, Mehra MR, Schuepbach RA, Ruschitzka F, Moch H (2020) Endothelial cell infection and endotheliitis in COVID-19. Lancet 395:1417–1418
- 72. Flammer AJ, Anderson T, Celermajer DS, Creager MA, Deanfield J, Ganz P, Hamburg NM, Lüscher TF, Shechter M, Taddei S (2012) The assessment of endothelial function: from research into clinical practice. Circulation 126:753–767
- 73. Hamming I, Timens W, Bulthuis M, Lely A, Gv N, van Goor H (2004) Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. J Pathol 203:631–637
- 74. Glenn JD, Smith MD, Xue P, Chan-Li Y, Collins S, Calabresi PA, Horton MR, Whartenby KA (2017) CNS-targeted autoimmunity leads to increased influenza mortality in mice. J Exp Med 214:297–307
- Razi O, Tartibian B, Laher I, Govindasamy K, Zamani N, Rocha-Rodrigues S, Suzuki K, Zouhal H (2022) Multimodal benefits of exercise in patients with multiple sclerosis and COVID-19. Front Physiol. https://doi.org/10.3389/fphys.2022. 783251
- 76. Razi O, Parnow A, Rashidi I, Pakravan N, Nedaei SE, Motl RW (2022) Aerobic training improves blood-brain barrier and neuronal apoptosis in experimental autoimmune encephalomyelitis. Iran J Basic Med Sci 25:245
- 77. Jick S, Li L, Falcone G, Vassilev Z, Wallander M-A (2014) Mortality of patients with multiple sclerosis: a cohort study in UK primary care. J Neurol 261:1508–1517
- 78. Lalmohamed A, Bazelier M, Van Staa T, Uitdehaag B, Leufkens H, De Boer A, De Vries F (2012) Causes of death in patients with multiple sclerosis and matched referent subjects: a population-based cohort study. Eur J Neurol 19:1007–1014

- Sumelahti ML, Hakama M, Elovaara I, Pukkala E (2010) Causes of death among patients with multiple sclerosis. Mult Scler J 16:1437–1442
- Marrie RA, Elliott L, Marriott J, Cossoy M, Blanchard J, Leung S, Yu N (2015) Effect of comorbidity on mortality in multiple sclerosis. Neurology 85:240–247
- Bender BS, Croghan T, Zhang L, Small P Jr (1992) Transgenic mice lacking class I major histocompatibility complex-restricted T cells have delayed viral clearance and increased mortality after influenza virus challenge. J Exp Med 175:1143–1145
- Thomas PG, Keating R, Hulse-Post DJ, Doherty PC (2006) Cell-mediated protection in influenza infection. Emerg Infect Dis 12:48
- Zhu B, Bando Y, Xiao S, Yang K, Anderson AC, Kuchroo VK, Khoury SJ (2007) CD11b+ Ly-6Chi suppressive monocytes in experimental autoimmune encephalomyelitis. J Immunol 179:5228–5237
- Gabrilovich DI, Nagaraj S (2009) Myeloid-derived suppressor cells as regulators of the immune system. Nat Rev Immunol 9:162–174
- Sinha P, Clements VK, Bunt SK, Albelda SM, Ostrand-Rosenberg S (2007) Cross-talk between myeloid-derived suppressor cells and macrophages subverts tumor immunity toward a type 2 response. J Immunol 179:977–983
- Marrie RA, Elliott L, Marriott J, Cossoy M, Blanchard J, Tennakoon A, Yu N (2014) Dramatically changing rates and reasons for hospitalization in multiple sclerosis. Neurology 83:929–937
- De Keyser J, Zwanikken C, Boon M (1998) Effects of influenza vaccination and influenza illness on exacerbations in multiple sclerosis. J Neurol Sci 159:51–53
- Montgomery S, Hillert J, Bahmanyar S (2013) Hospital admission due to infections in multiple sclerosis patients. Eur J Neurol 20:1153–1160
- Oikonen M, Laaksonen M, Aalto V, Ilonen J, Salonen R, Erälinna JP, Panelius M, Salmi A (2011) Temporal relationship between environmental influenza A and epstein-barr viral infections and high multiple sclerosis relapse occurrence. Mult Scler J 17:672–680
- 90. Pavelek Z, Angelucci F, Souček O, Krejsek J, Sobíšek L, Klímová B, Šarláková J, Halúsková S, Kuča K, Vališ M (2020) Innate immune system and multiple sclerosis. Granulocyte numbers are reduced in patients affected by relapsing-remitting multiple sclerosis during the remission phase. J Clin Med. https:// doi.org/10.3390/jcm9051468
- 91. Saferding V, Blüml S (2020) Innate immunity as the trigger of systemic autoimmune diseases. J Autoimmun 110:102382
- Yadav SK, Mindur JE, Ito K, Dhib-Jalbut S (2015) Advances in the immunopathogenesis of multiple sclerosis. Curr Opin Neurol 28:206–219
- Cadman E, Lawrence R (2010) Granulocytes: effector cells or immunomodulators in the immune response to helminth infection? Parasite Immunol 32:1–19
- Eyerich S, Metz M, Bossios A, Eyerich K (2020) New biological treatments for asthma and skin allergies. Allergy 75:546–560
- Ueda Y, Kondo M, Kelsoe G (2005) Inflammation and the reciprocal production of granulocytes and lymphocytes in bone marrow. J Exp Med 201:1771–1780
- 96. Javan MR, Aslani S, Zamani MR, Rostamnejad J, Asadi M, Farhoodi M, Nicknam MH (2016) Downregulation of immunosuppressive molecules, PD-1 and PD-L1 but not PD-L2, in the patients with multiple sclerosis. Iranian J Allergy, Asthma Immunol 15(4):296–302
- 97. Grebenciucova E, Berger JR (2017) Immunosenescence: the role of aging in the predisposition to neuro-infectious complications

arising from the treatment of multiple sclerosis. Curr Neurol Neurosci Rep 17:1–10

- Ibáñez AO, Laviñeta JC, Blanco TA (2022) Immunosenescence: the role of age in multiple sclerosis. In: Ostolaza Ibáñez A (ed) Neurología (English edition). Navarra, Spain
- Tullman MJ (2013) Overview of the epidemiology, diagnosis, and disease progression associated with multiple sclerosis. Am J Manag Care 19:S15-20
- Buscarinu MC, Reniè R, Morena E, Romano C, Bellucci G, Marrone A, Bigi R, Salvetti M, Ristori G (2022) Late-onset MS: disease course and safety-efficacy of DMTS. Front Neurol. https://doi.org/10.3389/fneur.2022.829331
- Appay V, Sauce D, Prelog M (2010) The role of the thymus in immunosenescence: lessons from the study of thymectomized individuals. Aging (Albany NY) 2:78
- 102. Algahtani H, Subahi A, Shirah B (2016) Neurological complications of middle east respiratory syndrome coronavirus: a report of two cases and review of the literature. Case rep neurol med 2016. https://doi.org/10.1155/2016/3502683
- Edwards S, Zvartau M, Clarke H, Irving W, Blumhardt L (1998) Clinical relapses and disease activity on magnetic resonance imaging associated with viral upper respiratory tract infections in multiple sclerosis. J Neurol Neurosurg Psychiatry 64:736–741
- 104. Narod S, Johnson-Lussenburg C, Zheng Q, Nelson R, Alperovitch A, Berr C, Sibley W (1985) Clinical viral infections and multiple sclerosis. Lancet (London, England) 326:165
- 105. Andersen O, Lygner P-E, Bergström T, Andersson M, Vablne A (1993) Viral infections trigger multiple sclerosis relapses: a prospective seroepidemiological study. J Neurol 240:417–422
- Burks JS, DeVald B, Jankovsky LD, Gerdes JC (1980) Two coronaviruses isolated from central nervous system tissue of two multiple sclerosis patients. Science 209:933–934
- 107. Hirota K, Yoshitomi H, Hashimoto M, Maeda S, Teradaira S, Sugimoto N, Yamaguchi T, Nomura T, Ito H, Nakamura T (2007) Preferential recruitment of CCR6-expressing Th17 cells to inflamed joints via CCL20 in rheumatoid arthritis and its animal model. J Exp Med 204:2803–2812
- Souberbielle BE, Szawlowski PW, Russell WC (1995) Is there a case for a virus aetiology in multiple sclerosis? Scott Med J 40:55–62
- Berg-Weger M, Morley JE (2020) Loneliness and social isolation in older adults during the Covid-19 pandemic: implications for gerontological social work. Springer, Cham
- Morley JE, Kalantar-Zadeh K, Anker SD (2020) COVID-19: a major cause of cachexia and sarcopenia? J Cachexia Sarcopenia Muscle 11:863–865
- 111. Hermans G, Van den Berghe G (2015) Clinical review: intensive care unit acquired weakness. Crit Care 19:1–9
- 112. Ferrante LE, Pisani MA, Murphy TE, Gahbauer EA, Leo-Summers LS, Gill TM (2018) The association of frailty with post-ICU disability, nursing home admission, and mortality: a longitudinal study. Chest 153:1378–1386
- 113. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, Xiang J, Wang Y, Song B, Gu X (2020) Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. Lancet 395:1054–1062
- 114. Dres M, Demoule A (2018) Diaphragm dysfunction during weaning from mechanical ventilation: an underestimated phenomenon with clinical implications. Crit Care 22:1–8
- 115. Dres M, Jung B, Molinari N, Manna F, Dubé B-P, Chanques G, Similowski T, Jaber S, Demoule A (2019) Respective contribution of intensive care unit-acquired limb muscle and severe diaphragm weakness on weaning outcome and mortality: a post hoc analysis of two cohorts. Crit Care 23:1–9

- 116. Goligher EC, Dres M, Fan E, Rubenfeld GD, Scales DC, Herridge MS, Vorona S, Sklar MC, Rittayamai N, Lanys A (2018) Mechanical ventilation–induced diaphragm atrophy strongly impacts clinical outcomes. Am J Respir Crit Care Med 197:204–213
- 117. Jonkman A, Jansen D, Heunks LM (2017) Novel insights in ICUacquired respiratory muscle dysfunction: implications for clinical care. Ann Update Intensive Care Emerg Med 2017:291–301
- 118. Demoule A, Jung B, Prodanovic H, Molinari N, Chanques G, Coirault C, Matecki S, Duguet A, Similowski T, Jaber S (2013) Diaphragm dysfunction on admission to the intensive care unit. Prevalence, risk factors, and prognostic impact—a prospective study. Am J Respir Crit Care Med 188:213–219
- 119. Levine S, Nguyen T, Taylor N, Friscia ME, Budak MT, Rothenberg P, Zhu J, Sachdeva R, Sonnad S, Kaiser LR (2008) Rapid disuse atrophy of diaphragm fibers in mechanically ventilated humans. N Engl J Med 358:1327–1335
- 120. Hussain SN, Mofarrahi M, Sigala I, Kim HC, Vassilakopoulos T, Maltais F, Bellenis I, Chaturvedi R, Gottfried SB, Metrakos P (2010) Mechanical ventilation–induced diaphragm disuse in humans triggers autophagy. Am J Respir Crit Care Med 182:1377–1386
- 121. White LJ, Castellano V (2008) Exercise and brain health—implications for multiple sclerosis. Sports Med 38:91–100
- 122. Jaber S, Petrof BJ, Jung B, Chanques G, Berthet J-P, Rabuel C, Bouyabrine H, Courouble P, Koechlin-Ramonatxo C, Sebbane M (2011) Rapidly progressive diaphragmatic weakness and injury during mechanical ventilation in humans. Am J Respir Crit Care Med 183:364–371
- 123. Hooijman PE, Beishuizen A, Witt CC, de Waard MC, Girbes AR, Spoelstra-de Man AM, Niessen HW, Manders E, van Hees HW, van den Brom CE (2015) Diaphragm muscle fiber weakness and ubiquitin–proteasome activation in critically ill patients. Am J Respir Crit Care Med 191:1126–1138
- 124. Conti P, Ronconi G, Caraffa A, Gallenga C, Ross R, Frydas I, Kritas S (2020) Induction of pro-inflammatory cytokines (IL-1 and IL-6) and lung inflammation by coronavirus-19 (COVI-19 or SARS-CoV-2): anti-inflammatory strategies. J Biol Regul Homeost Agents 34:1
- 125. Powers SK, Kavazis AN, DeRuisseau KC (2005) Mechanisms of disuse muscle atrophy: role of oxidative stress. Am J Physiol-Regulatory, Integr Comparative Physiol 288:R337–R344
- Powers SK, Kavazis AN, McClung JM (2007) Oxidative stress and disuse muscle atrophy. J Appl Physiol 102:2389–2397
- 127. McClung JM, Kavazis AN, DeRuisseau KC, Falk DJ, Deering MA, Lee Y, Sugiura T, Powers SK (2007) Caspase-3 regulation of diaphragm myonuclear domain during mechanical ventilation–induced atrophy. Am J Respir Crit Care Med 175:150–159
- Costamagna D, Costelli P, Sampaolesi M, Penna F (2015) Role of inflammation in muscle homeostasis and myogenesis. Mediators inflamm. https://doi.org/10.1155/2015/805172
- Londhe P, Guttridge DC (2015) Inflammation induced loss of skeletal muscle. Bone 80:131–142
- Degens H (2010) The role of systemic inflammation in agerelated muscle weakness and wasting. Scand J Med Sci Sports 20:28–38
- Bonaldo P, Sandri M (2013) Cellular and molecular mechanisms of muscle atrophy. Dis Model Mech 6:25–39
- 132. Bodine SC, Latres E, Baumhueter S, Lai VK-M, Nunez L, Clarke BA, Poueymirou WT, Panaro FJ, Na E, Dharmarajan K (2001) Identification of ubiquitin ligases required for skeletal muscle atrophy. Science 294:1704–1708
- Reid MB, Li Y-P (2001) Tumor necrosis factor-α and muscle wasting: a cellular perspective. Respir Res 2:1–4
- Frost RA, Lang CH, Gelato MC (1997) Transient exposure of human myoblasts to tumor necrosis factor-α inhibits serum and

insulin-like growth factor-I stimulated protein synthesis. Endocrinology 138:4153-4159

- Goodman MN (1991) Tumor necrosis factor induces skeletal muscle protein breakdown in rats. American J Physiol-Endocrinol Metab 260:E727–E730
- 136. Broussard SR, MCCusker RH, Novakofski JE, Strle K, Hong Shen W, Johnson RW, Freund GG, Dantzer R, Kelley KW (2003) Cytokine-hormone interactions: tumor necrosis factor α impairs biologic activity and downstream activation signals of the insulin-like growth factor I receptor in myoblasts. Endocrinology 144:2988–2996
- Li YP, Schwartz RJ, Waddell ID, Holloway BR, Reid MB (1998) Skeletal muscle myocytes undergo protein loss and reactive oxygen-mediated NF-κB activation in response to tumor necrosis factorα. FASEB J 12:871–880
- 138. Phillips T, Leeuwenburgh C (2005) Muscle fiber-specific apoptosis and TNF- $\alpha$  signaling in sarcopenia are attenuated by life-long calorie restriction. FASEB J 19:1–33
- 139. Cai D, Frantz JD, Tawa NE Jr, Melendez PA, Oh B-C, Lidov HG, Hasselgren P-O, Frontera WR, Lee J, Glass DJ (2004) IKKβ/ NF-κB activation causes severe muscle wasting in mice. Cell 119:285–298
- 140. Jackman RW, Cornwell EW, Wu CL, Kandarian SC (2013) Nuclear factor-κB signalling and transcriptional regulation in skeletal muscle atrophy. Exp Physiol 98:19–24
- 141. Batista M Jr, Peres S, McDonald M, Alcântara PSMd, Olivan M, Otoch JP, Farmer S, Seelaender M (2012) Adipose tissue inflammation and cancer cachexia: possible role of nuclear transcription factors. Cytokine 57:9–16
- 142. Ferrandi PJ, Alway SE, Mohamed JS (2020) Last Word on Viewpoint: the interaction between SARS-CoV-2 and ACE2 may have consequences for skeletal muscle viral susceptibility and myopathies. J Appl Physiol 129:872–872
- 143. Bahat G (2020) Covid-19 and the renin angiotensin system: implications for the older adults. J Nutr Health Aging 24:699–704
- 144. Betters JL, Criswell DS, Shanely RA, Van Gammeren D, Falk D, DeRuisseau KC, Deering M, Yimlamai T, Powers SK (2004) Trolox attenuates mechanical ventilation–induced diaphragmatic dysfunction and proteolysis. Am J Respir Crit Care Med 170:1179–1184
- Kondo H, Miura M, Itokawa Y (1991) Oxidative stress in skeletal muscle atrophied by immobilization. Acta Physiol Scand 142:527–528
- 146. Kondo H, Miura M, Kodama J, Ahmed SM, Itokawa Y (1992) Role of iron in oxidative stress in skeletal muscle atrophied by immobilization. Pflugers Arch 421:295–297
- 147. Reid MB, Khawli F, Moody MR (1993) Reactive oxygen in skeletal muscle. III. Contractility of unfatigued muscle. J Appl Physiol 75:1081–1087
- Smith MA, Reid MB (2006) Redox modulation of contractile function in respiratory and limb skeletal muscle. Respir Physiol Neurobiol 151:229–241
- 149. Andrade FH, Reid MB, Westerblad H (2001) Contractile response to low peroxide concentrations: myofibrillar calcium sensitivity as a likely target for redox-modulation of skeletal muscle function. FASEB J 15:309–311
- Anzai K, Ogawa K, Ozawa T, Yamamoto H (2000) Oxidative modification of ion channel activity of ryanodine receptor. Antioxid Redox Signal 2:35–40
- Abramson JJ, Salama G (1989) Critical sulfhydryls regulate calcium release from sarcoplasmic reticulum. J Bioenerg Biomembr 21:283–294
- 152. Fabisiak JP, Ritov VB, Kagan VE (2000) Reversible thioldependent activation of ryanodine-sensitive Ca2+ release

channel by etoposide (VP-16) phenoxyl radical. Antioxid Redox Signal 2:73-82

- 153. Daiho T, Kanazawa T (1994) Reduction of disulfide bonds in sarcoplasmic reticulum Ca (2+)-ATPase by dithiothreitol causes inhibition of phosphoenzyme isomerization in catalytic cycle. This reduction requires binding of both purine nucleotide and Ca2+ to enzyme. J Biol Chem 269:11060–11064
- 154. Yamada T, Mishima T, Sakamoto M, Sugiyama M, Matsunaga S, Wada M (2006) Oxidation of myosin heavy chain and reduction in force production in hyperthyroid rat soleus. J Appl Physiol 100:1520–1526
- 155. Haycock JW, Jones P, Harris JB, Mantle D (1996) Differential susceptibility of human skeletal muscle proteins to free radical induced oxidative damage: a histochemical, immunocytochemical and electron microscopical study in vitro. Acta Neuropathol 92:331–340
- 156. Coirault C, Guellich A, Barbry T, Samuel JL, Riou B, Lecarpentier Y (2007) Oxidative stress of myosin contributes to skeletal muscle dysfunction in rats with chronic heart failure. Am J Physiol-Heart Circulatory Physiol 292:H1009–H1017
- 157. Plant DR, Lynch GS, Williams DA (2000) Hydrogen peroxide modulates Ca2+-activation of single permeabilized fibres from fast-and slow-twitch skeletal muscles of rats. J Muscle Res Cell Motil 21:747–752
- 158. Shanely RA, Zergeroglu MA, Lennon SL, Sugiura T, Yimlamai T, Enns D, Belcastro A, Powers SK (2002) Mechanical ventilation–induced diaphragmatic atrophy is associated with oxidative injury and increased proteolytic activity. Am J Respir Crit Care Med 166:1369–1374
- Koh TJ, Tidball JG (2000) Nitric oxide inhibits calpain-mediated proteolysis of talin in skeletal muscle cells. Am J Physiol Cell Physiol 279:C806–C812
- 160. Román GC, Spencer PS, Reis J, Buguet A, Faris MEA, Katrak SM, Láinez M, Medina MT, Meshram C, Mizusawa H (2020) The neurology of COVID-19 revisited: a proposal from the environmental neurology specialty group of the world federation of neurology to implement international neurological registries. J Neurol Sci 414:116884
- 161. Wan S, Yi Q, Fan S, Lv J, Zhang X, Guo L, Lang C, Xiao Q, Xiao K, Yi Z (2020) Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP). MedRxiv. https://doi. org/10.1101/2020.02.10.20021832
- 162. Zhang Y, Geng X, Tan Y, Li Q, Xu C, Xu J, Hao L, Zeng Z, Luo X, Liu F (2020) New understanding of the damage of SARS-CoV-2 infection outside the respiratory system. Biomed Pharmacother 127:110195
- 163. Li C, Yang P, Zhang Y, Sun Y, Wang W, Zou Z, Xing L, Chen Z, Tang C, Guo F (2012) Corticosteroid treatment ameliorates acute lung injury induced by 2009 swine origin influenza A (H1N1) virus in mice. PLoS ONE. https://doi.org/10.1371/journal.pone. 0044110
- 164. Younan P, Iampietro M, Nishida A, Ramanathan P, Santos RI, Dutta M, Lubaki NM, Koup RA, Katze MG, Bukreyev A (2017) Ebola virus binding to Tim-1 on T lymphocytes induces a cytokine storm. MBio 8:e00845-e917
- 165. Boşnak Güçlü M, Güçlü Gündüz A, Nazliel B, Irkec C (2012) Comparison of functional exercise capacity, pulmonary function and respiratory muscle strength in patients with multiple sclerosis with different disability levels and healthy controls. J rehabilit med 44(1):80–86
- Aboussouan LS (2005) Respiratory disorders in neurologic diseases. Clevel Clin J Med 72:511
- Mutluay F, Gürses H, Saip S (2005) Effects of multiple sclerosis on respiratory functions. Clin Rehabil 19:426–432

- Hirst C, Swingler R, Compston D, Ben-Shlomo Y, Robertson NP (2008) Survival and cause of death in multiple sclerosis: a prospective population-based study. J Neurol Neurosurg Psychiatry 79:1016–1021
- McCool FD, Tzelepis GE (2012) Dysfunction of the diaphragm. N Engl J Med 366:932–942
- Howard R, Wiles C, Hirsch N, Loh L, Spencer G, Newsom-Davis J (1992) Respiratory involvement in multiple sclerosis. Brain 115:479–494
- Tzelepis GE, McCool FD (2015) Respiratory dysfunction in multiple sclerosis. Respir Med 109:671–679
- 172. Smeltzer SC, Skurnick JH, Troiano R, Cook SD, Duran W, Lavietes MH (1992) Respiratory function in multiple sclerosis: utility of clinical assessment of respiratory muscle function. Chest 101:479–484
- 173. Farhat MR, Loring SH, Riskind P, Weinhouse G (2013) Disturbance of respiratory muscle control in a patient with early-stage multiple sclerosis. Eur Respir J 41:1454–1456
- Westerdahl E, Gunnarsson M, Wittrin A, Nilsagård Y (2021) Pulmonary function and respiratory muscle strength in patients with multiple sclerosis. Multiple Sclerosis Intern. https://doi.org/ 10.1155/2021/5532776
- 175. Dereli M, Kahraman BO, Kahraman T (2022) A narrative review of respiratory impairment, assessment, and rehabilitation in multiple sclerosis. Dubai Medical J. https://doi.org/10.1159/00052 1444
- 176. Razi O, Tartibian B, Teixeira AM, Zamani N, Govindasamy K, Suzuki K, Laher I, Zouhal H (2022) Thermal dysregulation in patients with multiple sclerosis during SARS-CoV-2 infection. The potential therapeutic role of exercise. Multiple Scler Relat Disord 59:103557
- 177. Linker R, Mohr A, Cepek L, Gold R, Prange H (2006) Core hypothermia in multiple sclerosis: case report with magnetic resonance imaging localization of a thalamic lesion. Mult Scler J 12:112–115
- 178. White K, Scoones D, Newman P (1996) Hypothermia in multiple sclerosis. J Neurol Neurosurg Psychiatry 61:369–375
- Braun AP (2012) Two-pore domain potassium channels: variation on a structural theme. Channels 6:139–140
- Griffin JD, Boulant JA (1995) Temperature effects on membrane potential and input resistance in rat hypothalamic neurones. J Physiol 488:407–418
- Wechselberger M, Wright CL, Bishop GA, Boulant JA (2006) Ionic channels and conductance-based models for hypothalamic neuronal thermosensitivity. Am J Physiol-Regul, Integr Comparative Physiol 291:R518–R529
- Ray AD, Mahoney MC, Fisher NM (2015) Measures of respiratory function correlate with fatigue in ambulatory persons with multiple sclerosis. Disabil Rehabil 37:2407–2412
- 183. Koseoglu B, Gokkaya N, Ergun U, Inan L, Yesiltepe E (2006) Cardiopulmonary and metabolic functions, aerobic capacity, fatigue and quality of life in patients with multiple sclerosis. Acta Neurol Scand 114:261–267
- 184. Wens I, Eijnde BO, Hansen D (2016) Muscular, cardiac, ventilatory and metabolic dysfunction in patients with multiple sclerosis: Implications for screening, clinical care and endurance and resistance exercise therapy, a scoping review. J Neurol Sci 367:107–121
- 185. Asano M, Duquette P, Andersen R, Lapierre Y, Mayo NE (2013) Exercise barriers and preferences among women and men with multiple sclerosis. Disabil Rehabil 35:353–361
- Guthrie TC, Nelson DA (1995) Influence of temperature changes on multiple sclerosis: critical review of mechanisms and research potential. J Neurol Sci 129:1–8

- White A, Wilson T, Davis S, Petajan J (2000) Effect of precooling on physical performance in multiple sclerosis. Mult Scler J 6:176–180
- Motl RW, McAuley E, Snook EM (2005) Physical activity and multiple sclerosis: a meta-analysis. Mult Scler J 11:459–463
- Stroud N, Minahan C, Sabapathy S (2009) The perceived benefits and barriers to exercise participation in persons with multiple sclerosis. Disabil Rehabil 31:2216–2222
- 190. Turner AP, Kivlahan DR, Haselkorn JK (2009) Exercise and quality of life among people with multiple sclerosis: looking beyond physical functioning to mental health and participation in life. Arch Phys Med Rehabil 90:420–428
- 191. Hedström AK, Olsson T, Alfredsson L (2012) High body mass index before age 20 is associated with increased risk for multiple sclerosis in both men and women. Mult Scler J 18:1334–1336
- 192. Gianfrancesco MA, Barcellos LF (2016) Obesity and multiple sclerosis susceptibility: a review. J Neurol Neuromed 1:1
- 193. Langer-Gould A, Brara SM, Beaber BE, Koebnick C (2013) Childhood obesity and risk of pediatric multiple sclerosis and clinically isolated syndrome. Neurology 80:548–552
- 194. Munger KL, Bentzen J, Laursen B, Stenager E, Koch-Henriksen N, Sørensen TI, Baker JL (2013) Childhood body mass index and multiple sclerosis risk: a long-term cohort study. Mult Scler J 19:1323–1329
- 195. Pilutti LA, Motl RW (2019) Body composition and disability in people with multiple sclerosis: a dual-energy x-ray absorptiometry study. Mult scler relat disord 29:41–47
- Dixon AE, Peters U (2018) The effect of obesity on lung function. Expert Rev Respir Med 12:755–767
- Parameswaran K, Todd DC, Soth M (2006) Altered respiratory physiology in obesity. Can Respir J 13:203–210
- Naimark A, Cherniack R (1960) Compliance of the respiratory system and its components in health and obesity. J Appl Physiol 15:377–382
- 199. Pelosi P, Croci M, Ravagnan I, Tredici S, Pedoto A, Lissoni A, Gattinoni L (1998) The effects of body mass on lung volumes, respiratory mechanics, and gas exchange during general anesthesia. Anesth Analg 87:654–660
- Sharp J, Henry J, Sweany S, Meadows W, Pietras R (1964) The total work of breathing in normal and obese men. J Clin Investig 43:728–739
- Behazin N, Jones SB, Cohen RI, Loring SH (2010) Respiratory restriction and elevated pleural and esophageal pressures in morbid obesity. J Appl Physiol 108:212–218
- Sugerman H, Windsor A, Bessos M, Wolfe L (1997) Intraabdominal pressure, sagittal abdominal diameter and obesity comorbidity. J Intern Med 241:71–79
- 203. Chlif M, Keochkerian D, Choquet D, Vaidie A, Ahmaidi S (2009) Effects of obesity on breathing pattern, ventilatory neural drive and mechanics. Respir Physiol Neurobiol 168:198–202
- Collet F, Mallart A, Bervar J, Bautin N, Matran R, Pattou F, Romon M, Perez T (2007) Physiologic correlates of dyspnea in patients with morbid obesity. Int J Obes 31:700–706
- Jones RL, Nzekwu M-MU (2006) The effects of body mass index on lung volumes. Chest 130:827–833
- Ladosky W, Botelho M, Albuquerque J Jr (2001) Chest mechanics in morbidly obese non-hypoventilated patients. Respir Med 95:281–286
- 207. Sampson MG, Grassino AE (1983) Load compensation in obese patients during quiet tidal breathing. J Appl Physiol 55:1269–1276
- Schachter L, Salome C, Peat J, Woolcock A (2001) Obesity is a risk for asthma and wheeze but not airway hyperresponsiveness. Thorax 56:4–8

- 209. Sin DD, Jones RL, Man SP (2002) Obesity is a risk factor for dyspnea but not for airflow obstruction. Arch Intern Med 162:1477-1481
- Zerah F, Harf A, Perlemuter L, Lorino H, Lorino A-M, Atlan G (1993) Effects of obesity on respiratory resistance. Chest 103:1470–1476
- Zavorsky G, Hoffman S (2008) Pulmonary gas exchange in the morbidly obese. Obes Rev 9:326–339
- Holley H, Milic-Emili J, Becklake M, Bates D (1967) Regional distribution of pulmonary ventilation and perfusion in obesity. J Clin Investig 46:475–481
- Martinez-Santibañez G, Nien-Kai Lumeng C (2014) Macrophages and the regulation of adipose tissue remodeling. Annu Rev Nutr 34:57–76
- 214. Alessi M-C, Bastelica D, Morange P, Berthet B, Leduc I, Verdier M, Geel O, Juhan-Vague I (2000) Plasminogen activator inhibitor 1, transforming growth factor-beta1, and BMI are closely associated in human adipose tissue during morbid obesity. Diabetes 49:1374–1380
- Crewe C, An YA, Scherer PE (2017) The ominous triad of adipose tissue dysfunction: inflammation, fibrosis, and impaired angiogenesis. J Clin Investig 127:74–82
- 216. Di Gregorio GB, Yao-Borengasser A, Rasouli N, Varma V, Lu T, Miles LM, Ranganathan G, Peterson CA, McGehee RE, Kern PA (2005) Expression of CD68 and macrophage chemoattractant protein-1 genes in human adipose and muscle tissues: association with cytokine expression, insulin resistance, and reduction by pioglitazone. Diabetes 54:2305–2313
- 217. Mohamed-Ali V, Pinkney J, Coppack S (1998) Adipose tissue as an endocrine and paracrine organ. Int J Obes 22:1145–1158
- 218. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW (2003) Obesity is associated with macrophage accumulation in adipose tissue. J Clin Investig 112:1796–1808
- 219. Summer R, Walsh K, Medoff BD (2011) Obesity and pulmonary arterial hypertension: is adiponectin the molecular link between these conditions? Pulmonary circulation 1:440–447
- 220. Hsu P-S, Wu C-S, Chang J-F, Lin W-N (2015) Leptin promotes cPLA2 gene expression through activation of the MAPK/NF-κB/ p300 cascade. Int J Mol Sci 16:27640–27658
- 221. Woo J, Koziol-White C, Panettieri R Jr, Jude J (2021) TGF-β: The missing link in obesity-associated airway diseases? Curr Res Pharmacol Drug Discover 2:100016
- 222. Gui X, Chen H, Cai H, Sun L, Gu L (2018) Leptin promotes pulmonary fibrosis development by inhibiting autophagy via PI3K/Akt/mTOR pathway. Biochem Biophys Res Commun 498:660–666
- 223. de Boer WI, van Schadewijk A, Sont JK, Sharma HS, Stolk J, Hiemstra PS, van Krieken JHJ (1998) Transforming growth factor β1 and recruitment of macrophages and mast cells in airways in chronic obstructive pulmonary disease. Am J respir crit care med 158:1951–19577
- 224. Fong CY, Pang L, Holland E, Knox AJ (2000) TGF-β1 stimulates IL-8 release, COX-2 expression, and PGE2release in human airway smooth muscle cells. Am J Physiol-Lung Cell Mol Physiol 279:L201–L207
- 225. Kelley J, Kovacs EJ, Nicholson K, Fabisiak JP (1991) Transforming growth factor-β production by lung macrophages and fibroblasts. Chest 99:85S-86S
- 226. Lee K-Y, Ho S-C, Lin H-C, Lin S-M, Liu C-Y, Huang C-D, Wang C-H, Chung KF, Kuo H-P (2006) Neutrophil-derived elastase induces TGF-β1 secretion in human airway smooth muscle via NF-κB pathway. Am J Respir Cell Mol Biol 35:407–414
- 227. Minshall EM, Leung DY, Martin RJ, Song YL, Cameron L, Ernst P, Hamid Q (1997) Eosinophil-associated TGF-β1 mRNA

expression and airways fibrosis in bronchial asthma. Am J Respir Cell Mol Biol 17:326-333

- Jeffery PK (2001) Remodeling in asthma and chronic obstructive lung disease. Am J Respir Crit Care Med 164:S28–S38
- 229. e-Lacerda RR, Anhe GF, Page CP, Vasquez RY (2020) Sex differences in the influence of obesity on a murine model of allergic lung inflammation. Clin Exp Allergy 50:256–266
- 230. Grande JP, Melder DC, Zinsmeister AR (1997) Modulation of collagen gene expression by cytokines: stimulatory effect of transforming growth factor-β1, with divergent effects of epidermal growth factor and tumor necrosis factor-α on collagen type I and collagen type IV. J Lab Clin Med 130:476–486
- 231. Panettieri RA Jr, Tan EM, Ciocca V, Luttmann MA, Leonard TB, Hay DW (1998) Effects of LTD4 on human airway smooth muscle cell proliferation, matrix expression, and contraction in vitro: differential sensitivity to cysteinyl leukotriene receptor antagonists. Am J Respir Cell Mol Biol 19:453–461
- 232. Magdy Beshbishy A, Hetta HF, Hussein DE, Saati AA, Uba C, Rivero-Perez N, Zaragoza-Bastida A, Shah MA, Behl T, Batiha GES (2020) Factors associated with increased morbidity and mortality of obese and overweight COVID-19 patients. Biology 9:280
- 233. Damouche A, Lazure T, Avettand-Fènoël V, Huot N, Dejucq-Rainsford N, Satie A-P, Mélard A, David L, Gommet C, Ghosn J (2015) Adipose tissue is a neglected viral reservoir and an inflammatory site during chronic HIV and SIV infection. PLoS Pathog 11:e1005153
- 234. Jia X, Yin C, Lu S, Chen Y, Liu Q, Bai J and Lu Y (2020) Two things about COVID-19 might need attention. 2020020315. https://doi.org/10.20944/preprints202002.0315.v1
- 235. Yang J, Zheng Y, Gou X, Pu K, Chen Z, Guo Q, Ji R, Wang H, Wang Y, Zhou Y (2020) Prevalence of comorbidities and its effects in patients infected with SARS-CoV-2: a systematic review and meta-analysis. Int J Infect Dis 94:91–95
- 236. Ortiz ME, Thurman A, Pezzulo AA, Leidinger MR, Klesney-Tait JA, Karp PH, Tan P, Wohlford-Lenane C, McCray PB Jr, Meyerholz DK (2020) Heterogeneous expression of the SARS-Coronavirus-2 receptor ACE2 in the human respiratory tract. EBioMedicine 60:102976
- 237. Guo J, Huang Z, Lin L, Lv J (2020) Coronavirus disease 2019 (COVID-19) and cardiovascular disease: a viewpoint on the potential influence of angiotensin-converting enzyme inhibitors/ angiotensin receptor blockers on onset and severity of severe acute respiratory syndrome coronavirus 2 infection. J Am Heart Assoc 9:e016219
- Turner AJ (2015) ACE2 cell biology, regulation, and physiological functions. Prot Arm Ren Angiotensin Syst (RAS). https://doi. org/10.1016/B978-0-12-801364-9.00025-0
- Harmer D, Gilbert M, Borman R, Clark KL (2002) Quantitative mRNA expression profiling of ACE 2, a novel homologue of angiotensin converting enzyme. FEBS Lett 532:107–110
- 240. Razazian N, Almasi V, Afshari D, Bostani A, Moradian N, Farahvashi M (2018) Serum angiotensin-converting enzyme in patients suffering from multiple sclerosis and healthy controls: a pilot study. Neurophysiology 50:348–350
- Sumners C, Horiuchi M, Widdop RE, McCarthy C, Unger T, Steckelings UM (2013) Protective arms of the renin–angiotensin-system in neurological disease. Clin Exp Pharmacol Physiol 40:580–588
- 242. Rice GI, Thomas DA, Grant PJ, Turner AJ, Hooper NM (2004) Evaluation of angiotensin-converting enzyme (ACE), its homologue ACE2 and neprilysin in angiotensin peptide metabolism. Biochem J 383:45–51
- Stegbauer J, Lee D-H, Seubert S, Ellrichmann G, Manzel A, Kvakan H, Muller DN, Gaupp S, Rump LC, Gold R (2009) Role of

- 244. Santos RAS, Sampaio WO, Alzamora AC, Motta-Santos D, Alenina N, Bader M, Campagnole-Santos MJ (2017) The ACE2/ angiotensin-(1–7)/MAS axis of the renin-angiotensin system: focus on angiotensin-(1–7). Physiol Rev. https://doi.org/10.1152/ physrev.00023.2016
- Kalra J, Prakash A, Kumar P, Majeed ABA (2015) Cerebroprotective effects of RAS inhibitors: beyond their cardio-renal actions. J Ren-Angiotensin-Aldosterone Syst. https://doi.org/10. 1177/1470320315583582
- 246. Tikellis C, Thomas M (2012) Angiotensin-converting enzyme 2 (ACE2) is a key modulator of the renin angiotensin system in health and disease. Int J pept. https://doi.org/10.1155/2012/ 256294
- 247. Issa H, Eid AH, Berry B, Takhviji V, Khosravi A, Mantash S, Nehme R, Hallal R, Karaki H, Dhayni K (2021) Combination of angiotensin (1–7) agonists and convalescent plasma as a new strategy to overcome angiotensin converting enzyme 2 (ACE2) inhibition for the treatment of COVID-19. Front Med 8:278
- 248. Dimitrijevic I, Edvinsson M-L, Chen Q, Malmsjö M, Kimblad P-O, Edvinsson L (2009) Increased expression of vascular endothelin type B and angiotensin type 1 receptors in patients with ischemic heart disease. BMC Cardiovasc Disord 9:1–11
- 249. Roig E, Perez-Villa F, Morales M, Jimenez W, Orus J, Heras M, Sanz G (2000) Clinical implications of increased plasma angiotensin II despite ACE inhibitor therapy in patients with congestive heart failure. Eur Heart J 21:53–57
- 250. Gomes-Santos IL, Fernandes T, Couto GK, Ferreira-Filho JCA, Salemi VMC, Fernandes FB, Casarini DE, Brum PC, Rossoni LV, de Oliveira EM (2014) Effects of exercise training on circulating and skeletal muscle renin-angiotensin system in chronic heart failure rats. PLoS ONE 9:e98012
- 251. Wang K, Gheblawi M, Oudit GY (2020) Angiotensin converting enzyme 2: a double-edged sword. Circulation 142:426–428
- 252. Imai Y, Kuba K, Rao S, Huan Y, Guo F, Guan B, Yang P, Sarao R, Wada T, Leong-Poi H (2005) Angiotensin-converting enzyme 2 protects from severe acute lung failure. Nature 436:112–116
- 253. Leung JM, Yang CX, Tam A, Shaipanich T, Hackett T-L, Singhera GK, Dorscheid DR, Sin DD (2020) ACE-2 expression in the small airway epithelia of smokers and COPD patients: implications for COVID-19. Eur Respir J. https://doi.org/10.1183/13993 003.00688-2020
- 254. Anguiano L, Riera M, Pascual J, Soler M (2017) Circulating ACE2 in cardiovascular and kidney diseases. Curr Med Chem 24:3231–3241
- 255. Liu Y, Yang Y, Zhang C, Huang F, Wang F, Yuan J, Wang Z, Li J, Li J, Feng C (2020) Clinical and biochemical indexes from 2019nCoV infected patients linked to viral loads and lung injury. Sci China Life Sci 63:364–374
- 256. Rivellese F, Prediletto E (2020) ACE2 at the centre of COVID-19 from paucisymptomatic infections to severe pneumonia. Autoimmun Rev 19:102536
- 257. Magalhaes GS, Barroso LC, Reis AC, Rodrigues-Machado MG, Gregório JF, Motta-Santos D, Oliveira AC, Perez DA, Barcelos LS, Teixeira MM (2018) Angiotensin-(1–7) promotes resolution of eosinophilic inflammation in an experimental model of asthma. Front Immunol 9:58
- 258. Li Y, Zeng Z, Cao Y, Liu Y, Ping F, Liang M, Xue Y, Xi C, Zhou M, Jiang W (2016) Angiotensin-converting enzyme 2 prevents lipopolysaccharide-induced rat acute lung injury via suppressing the ERK1/2 and NF-κB signaling pathways. Sci Rep 6:1–14
- 259. Magalhães G, Rodrigues-Machado M, Motta-Santos D, Silva A, Caliari M, Prata L, Abreu S, Rocco P, Barcelos L, Santos R (2015) A ngiotensin-(1–7) attenuates airway remodelling and

1553

hyperresponsiveness in a model of chronic allergic lung inflammation. Br J Pharmacol 172:2330–2342

- 260. Kostenis E, Milligan G, Christopoulos A, Sanchez-Ferrer CF, Heringer-Walther S, Sexton PM, Gembardt F, Kellett E, Martini L, Vanderheyden P (2005) G-protein–coupled receptor Mas is a physiological antagonist of the angiotensin II type 1 receptor. Circulation 111:1806–1813
- Imai Y, Kuba K, Penninger JM (2007) Angiotensin-converting enzyme 2 in acute respiratory distress syndrome. Cell Mol Life Sci 64:2006–2012
- Fourrier F, Chopi C, Wallaert B, Mazurier C, Mangalahoyi J, Durocher A (1985) Compared evolution of plasma fibronectin and angiotensin-converting enzyme levels in septic ARDS. Chest 87:191–195
- 263. Abassi ZA, Skorecki K, Heyman SN, Kinaneh S, Armaly Z (2020) Covid-19 infection and mortality: a physiologist's perspective enlightening clinical features and plausible interventional strategies. Am J Physiol-Lung Cell Mol Physiol 318:L1020–L1022
- 264. Zhang H, Penninger JM, Li Y, Zhong N, Slutsky AS (2020) Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target. Intensive Care Med 46:586–590
- Batlle D, Wysocki J, Satchell K (2020) Soluble angiotensin-converting enzyme 2: a potential approach for coronavirus infection therapy? Clin Sci 134:543–545
- 266. Danser AJ, Epstein M, Batlle D (2020) Renin-angiotensin system blockers and the COVID-19 pandemic: at present there is no evidence to abandon renin-angiotensin system blockers. Hypertension 75:1382–1385
- 267. Magalhaes GS, Rodrigues-Machado MdG, Motta-Santos D, Campagnole-Santos MJ, Santos RAS (2020) Activation of Ang-(1–7)/Mas receptor is a possible strategy to treat coronavirus (SARS-CoV-2) infection. Front Physiol 11:730
- 268. Gwathmey TM, Pendergrass KD, Reid SD, Rose JC, Diz DI, Chappell MC (2010) Angiotensin-(1–7)-angiotensin-converting enzyme 2 attenuates reactive oxygen species formation to angiotensin II within the cell nucleus. Hypertension 55:166–171
- 269. Rodrigues Prestes TR, Rocha NP, Miranda AS, Teixeira AL, Simoes-e-Silva AC (2017) The anti-inflammatory potential of ACE2/angiotensin-(1–7)/mas receptor axis: evidence from basic and clinical research. Curr Drug Targets 18:1301–1313
- 270. Wösten-van Asperen RM, Lutter R, Specht PA, Moll GN, van Woensel JB, van der Loos CM, van Goor H, Kamilic J, Florquin S, Bos AP (2011) Acute respiratory distress syndrome leads to reduced ratio of ACE/ACE2 activities and is prevented by angiotensin-(1–7) or an angiotensin II receptor antagonist. J Pathol 225:618–627
- 271. Chen Q, Yang Y, Huang Y, Pan C, Liu L, Qiu H (2013) Angiotensin-(1–7) attenuates lung fibrosis by way of mas receptor in acute lung injury. J Surg Res 185(2):740–747
- 272. Seys LJ, Widagdo W, Verhamme FM, Kleinjan A, Janssens W, Joos GF, Bracke KR, Haagmans BL, Brusselle GG (2018) DPP4, the middle east respiratory syndrome coronavirus receptor, is upregulated in lungs of smokers and chronic obstructive pulmonary disease patients. Clin Infect Dis 66:45–53
- 273. McKinley MJ, Albiston AL, Allen AM, Mathai M, May C, McAllen RM, Oldfield BJ, Mendelsohn F, Chai SY (2003) The brain renin–angiotensin system: location and physiological roles. Int J Biochem Cell Biol 35:901–918
- 274. de Kloet AD, Liu M, Rodríguez V, Krause EG, Sumners C (2015) Role of neurons and glia in the CNS actions of the renin-angiotensin system in cardiovascular control. Am J Physiol-Regulatory Integr Comparative Physiol 309:R444–R458

- Constantinescu CS, Goodman DB, Grossman RI, Mannon LJ, Cohen JA (1997) Serum angiotensin-converting enzyme in multiple sclerosis. Arch Neurol 54:1012–1015
- 276. Lund B, Stone R, Levy A, Lee S, Amundson E, Kashani N, Rodgers K, Kelland E (2019) Reduced disease severity following therapeutic treatment with angiotensin 1–7 in a mouse model of multiple sclerosis. Neurobiol Dis 127:87–100
- 277. Oliveira-Lima OC, Pinto MC, Duchene J, Qadri F, Souza LL, Alenina N, Bader M, Santos RA, Carvalho-Tavares J (2015) Mas receptor deficiency exacerbates lipopolysaccharide-induced cerebral and systemic inflammation in mice. Immunobiology 220:1311–1321
- 278. Stone RE, Liu S, Levy AM, Kashani N, Louie SG, Rodgers KE, Kelland EE, Lund BT (2019) Activation of the protective arm of the renin angiotensin system in demyelinating disease. J Neuroimmune Pharmacol. https://doi.org/10.1007/s11481-019-09894-7
- 279. Platten M, Youssef S, Hur EM, Ho PP, Han MH, Lanz TV, Phillips LK, Goldstein MJ, Bhat R, Raine CS (2009) Blocking angiotensin-converting enzyme induces potent regulatory T cells and modulates TH1-and TH17-mediated autoimmunity. Proc Natl Acad Sci 106:14948–14953
- 280. Hammer A, Yang G, Friedrich J, Kovacs A, Lee D-H, Grave K, Jörg S, Alenina N, Grosch J, Winkler J (2016) Role of the receptor mas in macrophage-mediated inflammation in vivo. Proc Natl Acad Sci 113:14109–14114
- 281. Zhao Y, Qin Y, Liu T, Hao D (2015) Chronic nerve injuryinduced Mas receptor expression in dorsal root ganglion neurons alleviates neuropathic pain. Exp Ther Med 10:2384–2388
- Salehi S, Reddy S, Gholamrezanezhad A (2020) Long-term pulmonary consequences of coronavirus disease 2019 (COVID-19): what we know and what to expect. J Thorac Imaging 35:W87–W89
- 283. Xie L, Liu Y, Xiao Y, Tian Q, Fan B, Zhao H, Chen W (2005) Follow-up study on pulmonary function and lung radiographic changes in rehabilitating severe acute respiratory syndrome patients after discharge. Chest 127:2119–2124
- 284. Chen J, Wu J, Hao S, Yang M, Lu X, Chen X, Li L (2017) Long term outcomes in survivors of epidemic influenza A (H7N9) virus infection. Sci Rep 7:1–8
- 285. Salehi S, Abedi A, Balakrishnan S, Gholamrezanezhad A (2020) Coronavirus disease 2019 (COVID-19): a systematic review of imaging findings in 919 patients. Am J Roentgenol 215:87–93
- 286. Antonio GE, Wong K, Hui DS, Wu A, Lee N, Yuen EH, Leung C, Rainer TH, Cameron P, Chung SS (2003) Thin-section CT in patients with severe acute respiratory syndrome following hospital discharge: preliminary experience. Radiology 228:810–815
- 287. Rietberg MB, Veerbeek JM, Gosselink R, Kwakkel G, van Wegen EE (2017) Respiratory muscle training for multiple sclerosis. Cochrane Database Syst Rev. https://doi.org/10.1002/14651858. CD009424.pub2
- Göçmen R (2018) The relevance of neuroimaging findings to physical disability in multiple sclerosis. Arch Neuropsychiatry 55:S31
- Lee K, Rincon F (2012) Pulmonary complications in patients with severe brain injury. Crit Care Res Pract. https://doi.org/10. 1155/2012/207247
- 290. Gosselink R, Kovacs L, Ketelaer P, Carton H, Decramer M (2000) Respiratory muscle weakness and respiratory muscle training in severely disabled multiple sclerosis patients. Arch Phys Med Rehabil 81:747–751
- Gosselink R, Kovacs L, Decramer M (1999) Respiratory muscle involvement in multiple sclerosis. Eur Respir J 13:449–454
- Smeltzer SC, Levietes MH, Cook SD (1996) Expiratory training in multiple sclerosis. Arch Phys Med Rehabil 77:909–912

- 293. Tantucci C, Massucci M, Piperno R, Betti L, Grassi V, Sorbini CA (1994) Control of breathing and respiratory muscle strength in patients with multiple sclerosis. Chest 105:1163–1170
- 294. Li J, He X, Yuan Y, Zhang W, Li X, Zhang Y, Li S, Guan C, Gao Z, Dong G (2021) Meta-analysis investigating the relationship between clinical features, outcomes, and severity of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) pneumonia. Am J Infect Control 49:82–89
- 295. Kasraeian M, Zare M, Vafaei H, Asadi N, Faraji A, Bazrafshan K, Roozmeh S (2020) COVID-19 pneumonia and pregnancy; a systematic review and meta-analysis. J Maternal-Fetal & Neonatal Med. https://doi.org/10.1080/14767058.2020.1763952
- 296. Marks BL (2002) Physiologic responses to exercise in older women. Topics Geriatric Rehabil 18:9–20
- 297. Fink JE, Schoenfeld BJ, Kikuchi N, Nakazato K (2017) Acute and long-term responses to different rest intervals in low-load resistance training. Int J Sports Med 38:118–124
- 298. Jakhotia K, Jain N, Retharekar S, Shimpi A, Rairikar S, Sancheti P (2014) Effect of inspiratory muscle training (IMT) on aerobic performance in young healthy sedentary individuals. J Med Thesis 2:21–25
- 299. Okrzymowska P, Kurzaj M, Seidel W, Rożek-Piechura K (2019) Eight weeks of inspiratory muscle training improves pulmonary function in disabled swimmers—a randomized trial. Int J Environ Res Public Health 16:1747
- 300. Bruce CR, Thrush AB, Mertz VA, Bezaire V, Chabowski A, Heigenhauser GJ, Dyck DJ (2006) Endurance training in obese humans improves glucose tolerance and mitochondrial fatty acid oxidation and alters muscle lipid content. Am J Physiol-Endocrinol Metabol 291:E99–E107
- 301. Wilson JM, Loenneke JP, Jo E, Wilson GJ, Zourdos MC, Kim J-S (2012) The effects of endurance, strength, and power training on muscle fiber type shifting. J Strength Conditioning Res 26:1724–1729
- Pette D, Staron RS (2001) Transitions of muscle fiber phenotypic profiles. Histochem Cell Biol 115:359–372
- Holloszy JO (1976) Adaptations of muscular tissue to training. Prog Cardiovasc Dis 18:445–458
- Fitts RH, Booth F, Winder W, Holloszy J (1975) Skeletal muscle respiratory capacity, endurance, and glycogen utilization. Am J Physiol-Leg Content 228:1029–1033
- 305. Brooks G, Fahey T, White T (1996) Physiologic responses and long-term adaptations to exercise. In: Brooks GA (ed) Exercise physiology: human bioenergetics and its applications, 2nd edn. Mayfield Publishing Company, Mountain View (CA), pp 61–77
- 306. Lau HMC, Ng GYF, Jones AYM, Lee EWC, Siu EHK, Hui DSC (2005) A randomised controlled trial of the effectiveness of an exercise training program in patients recovering from severe acute respiratory syndrome. Aust J Physiother 51:213–219
- 307. Li X, Yu R, Wang P, Wang A, Huang H (2021) Effects of exercise training on cardiopulmonary function and quality of life in elderly patients with pulmonary fibrosis: a meta-analysis. Int J Environ Res Public Health 18:7643
- 308. King TE Jr, Bradford WZ, Castro-Bernardini S, Fagan EA, Glaspole I, Glassberg MK, Gorina E, Hopkins PM, Kardatzke D, Lancaster L (2014) A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis. N Engl J Med 370:2083–2092
- 309. Swigris J, Kuschner W, Jacobs S, Wilson S, Gould M (2005) Health-related quality of life in patients with idiopathic pulmonary fibrosis: a systematic review. Thorax 60:588–594
- Swigris JJ, Brown KK, Make BJ, Wamboldt FS (2008) Pulmonary rehabilitation in idiopathic pulmonary fibrosis: a call for continued investigation. Respir Med 102:1675–1680
- 311. HajGhanbari B, Yamabayashi C, Buna TR, Coelho JD, Freedman KD, Morton TA, Palmer SA, Toy MA, Walsh C, Sheel AW (2013) Effects of respiratory muscle training on performance

in athletes: a systematic review with meta-analyses. J Strength Conditioning Res 27:1643–1663

- Illi SK, Held U, Frank I, Spengler CM (2012) Effect of respiratory muscle training on exercise performance in healthy individuals. Sports Med 42:707–724
- 313. Ray AD, Udhoji S, Mashtare TL, Fisher NM (2013) A combined inspiratory and expiratory muscle training program improves respiratory muscle strength and fatigue in multiple sclerosis. Arch Phys Med Rehabil 94:1964–1970
- 314. Chiara T, Martin AD, Davenport PW, Bolser DC (2006) Expiratory muscle strength training in persons with multiple sclerosis having mild to moderate disability: effect on maximal expiratory pressure, pulmonary function, and maximal voluntary cough. Arch Phys Med Rehabil 87:468–473
- 315. Huang MH, Fry D, Doyle L, Burnham A, Houston N, Shea K, Smith H, Wiske L, Goode J, Khitrik E (2020) Effects of inspiratory muscle training in advanced multiple sclerosis. Mult Scler and Relat Disord 37:101492
- 316. Silverman EP, Miller S, Zhang Y, Hoffman-Ruddy B, Yeager J, Daly JJ (2017) Effects of expiratory muscle strength training on maximal respiratory pressure and swallow-related quality of life in individuals with multiple sclerosis. Mult Scler J-Exp Translational Clin 3:2055217317710829
- 317. Martin-Sanchez C, Calvo-Arenillas JI, Barbero-Iglesias FJ, Fonseca E, Sanchez-Santos JM, Martin-Nogueras AM (2020) Effects of 12-week inspiratory muscle training with low resistance in patients with multiple sclerosis: a non-randomised, double-blind, controlled trial. Mult Scler Relat Disord 46:102574
- 318. Ferreira JB, Plentz RDM, Stein C, Casali KR, Arena R, Dal Lago P (2013) Inspiratory muscle training reduces blood pressure and sympathetic activity in hypertensive patients: a randomized controlled trial. Int J Cardiol 166:61–67
- 319. Parshall MB, Schwartzstein RM, Adams L, Banzett RB, Manning HL, Bourbeau J, Calverley PM, Gift AG, Harver A, Lareau SC (2012) An official American thoracic society statement: update on the mechanisms, assessment, and management of dyspnea. Am J Respir Crit Care Med 185:435–452
- 320. Caruso F, Arena R, Phillips S, Bonjorno J Jr, Mendes R, Arakelian V, Bassi D, Nogi C, Borghi-Silva A (2015) Resistance exercise training improves heart rate variability and muscle performance: a randomized controlled trial in coronary artery disease patients. Eur J Phys Rehabil Med 51:281–289
- 321. Murad K, Brubaker PH, Fitzgerald DM, Morgan TM, Goff DC Jr, Soliman EZ, Eggebeen JD, Kitzman DW (2012) Exercise training improves heart rate variability in older patients with heart failure: a randomized, controlled, single-blinded trial. Congest Heart Fail 18:192–197
- 322. Abasıyanık Z, Ertekin Ö, Kahraman T, Yigit P, Özakbaş S (2020) The effects of clinical pilates training on walking, balance, fall risk, respiratory, and cognitive functions in persons with multiple sclerosis: a randomized controlled trial. Explore 16:12–20
- 323. Salgado BC, Jones M, Ilgun S, McCord G, Loper-Powers M, van Houten P (2013) Effects of a 4-month ananda yoga program on physical and mental health outcomes for persons with multiple sclerosis. Int J Yoga Ther 23:27–38
- 324. Abasıyanık Z, Yiğit P, Özdoğar AT, Kahraman T, Ertekin Ö, Özakbaş S (2021) A comparative study of the effects of yoga and clinical Pilates training on walking, cognition, respiratory functions, and quality of life in persons with multiple sclerosis: a quasi-experimental study. Explore 17:424–429
- 325. Sable M, Vaidya S, Sable S (2012) Short communication comparative study of lung functions in swimmers and runners. Indian J Physiol Pharmacol 56(1):100–104

- 326. Vašíčková J, Neumannová K, Svozil Z (2017) The effect of respiratory muscle training on fin-swimmers' performance. J Sports Sci Med 16:521
- 327. Enright S, Chatham K, Ionescu AA, Unnithan VB, Shale DJ (2004) Inspiratory muscle training improves lung function and exercise capacity in adults with cystic fibrosis. Chest 126:405–411
- 328. Kang S-W, Bach JR (2000) Maximum insufflation capacity: vital capacity and cough flows in neuromuscular disease. Am J Phys Med Rehabil 79:222–227
- 329. Mackała K, Kurzaj M, Okrzymowska P, Stodółka J, Coh M, Rożek-Piechura K (2020) The effect of respiratory muscle training on the pulmonary function, lung ventilation, and endurance performance of young soccer players. Int J Environ Res Public Health 17:234
- Mickleborough TD, Stager JM, Chatham K, Lindley MR, Ionescu AA (2008) Pulmonary adaptations to swim and inspiratory muscle training. Eur J Appl Physiol 103:635–646
- Marini JJ, Gattinoni L (2020) Management of COVID-19 respiratory distress. JAMA 323:2329–2330
- 332. Sheel AW, Derchak PA, Morgan BJ, Pegelow DF, Jacques AJ, Dempsey JA (2001) Fatiguing inspiratory muscle work causes reflex reduction in resting leg blood flow in humans. J Physiol 537:277–289
- 333. Romer LM, Lovering AT, Haverkamp HC, Pegelow DF, Dempsey JA (2006) Effect of inspiratory muscle work on peripheral fatigue of locomotor muscles in healthy humans. J Physiol 571:425–439
- 334. Enright SJ, Unnithan VB, Heward C, Withnall L, Davies DH (2006) Effect of high-intensity inspiratory muscle training on lung volumes, diaphragm thickness, and exercise capacity in subjects who are healthy. Phys Ther 86:345–354
- 335. Downey AE, Chenoweth LM, Townsend DK, Ranum JD, Ferguson CS, Harms CA (2007) Effects of inspiratory muscle training on exercise responses in normoxia and hypoxia. Respir Physiol Neurobiol 156:137–146
- 336. Romer LM, McConnell AK, Jones DA (2002) Inspiratory muscle fatigue in trained cyclists: effects of inspiratory muscle training. Med Sci Sports Exerc 34:785–792
- 337. Walsh NP, Gleeson M, Shephard RJ, Gleeson M, Woods JA, Bishop N, Fleshner M, Green C, Pedersen BK, Hoffman-Goete L (2011) Position statement part one: immune function and exercise. Exerc Immunol Rev 17:6–63
- 338. van de Weert-van PB, Arets HGM, van der Ent CK, Beekman JM (2013) Infection, inflammation and exercise in cystic fibrosis. Respir Res 14:1–10
- Mathur N, Pedersen BK (2008) Exercise as a mean to control low-grade systemic inflammation. Mediators inflamm. https:// doi.org/10.1155/2008/109502
- 340. Gleeson M, Bishop NC, Stensel DJ, Lindley MR, Mastana SS, Nimmo MA (2011) The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease. Nat Rev Immunol 11:607–615
- Brandt C, Pedersen BK (2010) The role of exercise-induced myokines in muscle homeostasis and the defense against chronic diseases. J Biomed Biotechnol. https://doi.org/10.1155/2010/ 520258
- 342. Pedersen BK, Steensberg A, Fischer C, Keller C, Ostrowski K, Schjerling P (2001) Exercise and cytokines with particular focus on muscle derived IL-6. Exerc Immunol Rev 7:18–31
- 343. Coelho Junior HJ, Gambassi BB, Diniz TA, Fernandes IMdC, Caperuto ÉC, Uchida MC, Lira FS, Rodrigues B (2016) Inflammatory mechanisms associated with skeletal muscle sequelae after stroke: role of physical exercise. Mediators Inflamm. https:// doi.org/10.1155/2016/3957958

- 344. Steensberg A, Fischer CP, Keller C, Møller K, Pedersen BK (2003) IL-6 enhances plasma IL-1ra, IL-10, and cortisol in humans. Am J Physiol-Endocrinol Metabol 285:E433–E437
- 345. Barton BE (1997) IL-6: insights into novel biological activities. Clin Immunol Immunopathol 85:16–20
- Maynard CL, Weaver CT (2008) Diversity in the contribution of interleukin-10 to T-cell-mediated immune regulation. Immunol Rev 226:219–233
- 347. Gonçalves CTR, Gonçalves CGR, de Almeida FM, dos Santos FDTQ, dos Santos Durão ACC, dos Santos FA, da Silva LFF, Marcourakis T, Castro-Faria-Neto HC, de Paula VR (2012) Protective effects of aerobic exercise on acute lung injury induced by LPS in mice. Crit Care 16:1–11
- 348. Nandi D, Mishra MK, Basu A, Bishayi B (2010) Protective effects of interleukin-6 in lipopolysaccharide (LPS)-induced experimental endotoxemia are linked to alteration in hepatic anti-oxidant enzymes and endogenous cytokines. Immunobiology 215:443–451
- Wolters PJ, Wray C, Sutherland RE, Kim SS, Koff J, Mao Y, Frank JA (2009) Neutrophil-derived IL-6 limits alveolar barrier disruption in experimental ventilator-induced lung injury. J Immunol 182:8056–8062
- Venihaki M, Dikkes P, Carrigan A, Karalis KP (2001) Corticotropin-releasing hormone regulates IL-6 expression during inflammation. J Clin Investig 108:1159–1166
- 351. Ropelle ER, Flores MB, Cintra DE, Rocha GZ, Pauli JR, Morari J, de Souza CT, Moraes JC, Prada PO, Guadagnini D (2010) IL-6 and IL-10 anti-inflammatory activity links exercise to hypothalamic insulin and leptin sensitivity through IKKβ and ER stress inhibition. PLoS Biol 8:e1000465
- 352. Royall JA, Berkow RL, Beckman JS, Cunningham MK, Matalon S, Freeman BA (1989) Tumor necrosis factor and interleukin 1 alpha increase vascular endothelial permeability. Am J Physiol-Lung Cell Mol Physiol 257:L399–L410
- 353. Linke A, Adams V, Schulze PC, Erbs S, Gielen S, Fiehn E, Möbius-Winkler S, Schubert A, Schuler G, Hambrecht R (2005) Antioxidative effects of exercise training in patients with chronic heart failure: increase in radical scavenger enzyme activity in skeletal muscle. Circulation 111:1763–1770
- 354. Menegali BT, Nesi RT, Souza PS, Silva LA, Silveira PC, Valença SS, Pinho RA (2009) The effects of physical exercise on the cigarette smoke-induced pulmonary oxidative response. Pulm Pharmacol Ther 22:567–573
- 355. Nemmar A, Al-Salam S, Yuvaraju P, Beegam S, Ali BH (2018) Exercise training mitigates water pipe smoke exposure-induced pulmonary impairment via inhibiting NF-κB and activating Nrf2 signalling pathways. Oxidative Med Cell Longevity. https://doi.org/10.1155/2018/7459612
- Pavlov VA, Tracey KJ (2005) The cholinergic anti-inflammatory pathway. Brain Behav Immun 19:493–499
- 357. Pavlov VA, Tracey KJ (2012) The vagus nerve and the inflammatory reflex—linking immunity and metabolism. Nat Rev Endocrinol 8:743–754
- 358. Skinner NA, MacIsaac C, Hamilton JA, Visvanathan K (2005) Regulation of toll-like receptor (TLR) 2 and TLR4 on CD14dimCD16+ monocytes in response to sepsis-related antigens. Clin Exp Immunol 141:270–278
- 359. Simpson RJ, McFarlin BK, McSporran C, Spielmann G, Hartaigh B, Guy K (2009) Toll-like receptor expression on classic and pro-inflammatory blood monocytes after acute exercise in humans. Brain Behavior Immunity 23:232–239
- 360. Yeh S, Chuang H, Lin L, Hsiao C, Eng H (2006) Regular tai chi chuan exercise enhances functional mobility and CD4CD25 regulatory T cells. Br J Sports Med 40:239–243
- 361. Yeh S, Chuang H, Lin L, Hsiao C, Wang P, Liu R, Yang K (2009) Regular tai chi chuan exercise improves T cell helper

function of patients with type 2 diabetes mellitus with an increase in T-bet transcription factor and IL-12 production. Br J Sports Med 43:845–850

- 362. Mee-Inta O, Zhao Z-W, Kuo Y-M (2019) Physical exercise inhibits inflammation and microglial activation. Cells 8:691
- 363. Pekny M, Pekna M (2014) Astrocyte reactivity and reactive astrogliosis: costs and benefits. Physiol Rev 94:1077–1098
- Arbour N, Day R, Newcombe J, Talbot PJ (2000) Neuroinvasion by human respiratory coronaviruses. J Virol 74:8913–8921
- 365. Ge Y (2006) Multiple sclerosis: the role of MR imaging. Am J Neuroradiol 27:1165–1176
- 366. Shi Y, Liu T, Nieman DC, Cui Y, Li F, Yang L, Shi H, Chen P (2020) Aerobic exercise attenuates acute lung injury through NET inhibition. Front Immunol 11:409
- 367. Dwyer TJ, Elkins MR, Bye PT (2011) The role of exercise in maintaining health in cystic fibrosis. Curr Opin Pulm Med 17:455–460
- 368. Nikolaizik W, Simon H, Iseli P, Blaser K, Schoni M (2000) Effect of 3 weeks' rehabilitation on neutrophil surface antigens and lung function in cystic fibrosis. Eur Respir J 15:942–948
- 369. Feter N, Freitas M, Gonzales N, Umpierre D, Cardoso R, Rombaldi A (2018) Effects of physical exercise on myelin sheath regeneration: a systematic review and meta-analysis. Sci Sports 33:8–21
- 370. Jensen SK, Yong VW (2016) Activity-dependent and experience-driven myelination provide new directions for the management of multiple sclerosis. Trends Neurosci 39:356–365
- 371. Rafalski VA, Ho PP, Brett JO, Ucar D, Dugas JC, Pollina EA, Chow LM, Ibrahim A, Baker SJ, Barres BA (2013) Expansion of oligodendrocyte progenitor cells following SIRT1 inactivation in the adult brain. Nat Cell Biol 15:614–624
- 372. St-Pierre J, Drori S, Uldry M, Silvaggi JM, Rhee J, Jäger S, Handschin C, Zheng K, Lin J, Yang W (2006) Suppression of reactive oxygen species and neurodegeneration by the PGC-1 transcriptional coactivators. Cell 127:397–408
- 373. Camacho A, Huang JK, Delint-Ramirez I, Yew Tan C, Fuller M, Lelliott CJ, Vidal-Puig A, Franklin RJ (2013) Peroxisome proliferator-activated receptor gamma-coactivator-1 alpha coordinates sphingolipid metabolism, lipid raft composition and myelin protein synthesis. Eur J Neurosci 38:2672–2683
- Greer JM, Lees MB (2002) Myelin proteolipid protein—the first 50 years. Int J Biochem Cell Biol 34:211–215
- 375. Michel K, Zhao T, Karl M, Lewis K, Fyffe-Maricich SL (2015) Translational control of myelin basic protein expression by ERK2 MAP kinase regulates timely remyelination in the adult brain. J Neurosci 35:7850–7865
- 376. Kohman RA, DeYoung EK, Bhattacharya TK, Peterson LN, Rhodes JS (2012) Wheel running attenuates microglia proliferation and increases expression of a proneurogenic phenotype in the hippocampus of aged mice. Brain Behav Immun 26:803–810
- 377. Vukovic J, Colditz MJ, Blackmore DG, Ruitenberg MJ, Bartlett PF (2012) Microglia modulate hippocampal neural precursor activity in response to exercise and aging. J Neurosci 32:6435–6443
- 378. Walsh NP, Oliver SJ (2016) Exercise, immune function and respiratory infection: an update on the influence of training and environmental stress. Immunol Cell Biol 94:132–139
- 379. Matthews CE, Ockene IS, Freedson PS, Rosal MC, Merriam PA, Hebert JR (2002) Moderate to vigorous physical activity and risk of upper-respiratory tract infection. Med Sci Sports Exerc 34:1242–1248
- 380. Klentrou P, Cieslak T, MacNeil M, Vintinner A, Plyley M (2002) Effect of moderate exercise on salivary immunoglobulin A and infection risk in humans. Eur J Appl Physiol 87:153–158
- 381. Shimizu K, Kimura F, Akimoto T, Akama T, Otsuki T, Nishijima T, Kuno S, Kono I (2007) Effects of exercise, age and gender

on salivary secretory immunoglobulin A in elderly individuals. Exerc Immunol Rev 13:55–66

- Lowder T, Padgett DA, Woods JA (2006) Moderate exercise early after influenza virus infection reduces the Th1 inflammatory response in lungs of mice. Exerc Immunol Rev 12:97–111
- Martin SA, Pence BD, Woods JA (2009) Exercise and respiratory tract viral infections. Exerc Sport Sci Rev 37:157
- Nieman DC, Henson DA, Austin MD, Brown VA (2005) Immune response to a 30-minute walk. Med Sci Sports Exerc 37:57–62
- Shephard RJ (1998) Immune changes induced by exercise in an adverse environment. Can J Physiol Pharmacol 76:539–546
- Dhabhar FS (2014) Effects of stress on immune function: the good, the bad, and the beautiful. Immunol Res 58:193–210
- Pedersen BK (2017) Anti-inflammatory effects of exercise: role in diabetes and cardiovascular disease. Eur J Clin Invest 47:600–611
- 388. Bigley AB, Rezvani K, Chew C, Sekine T, Pistillo M, Crucian B, Bollard CM, Simpson RJ (2014) Acute exercise preferentially redeploys NK-cells with a highly-differentiated phenotype and augments cytotoxicity against lymphoma and multiple myeloma target cells. Brain Behav Immun 39:160–171
- Adams GR, Zaldivar FP, Nance DM, Kodesh E, Radom-Aizik S, Cooper DM (2011) Exercise and leukocyte interchange among central circulation, lung, spleen, and muscle. Brain Behav Immun 25:658–666
- 390. Campbell JP, Turner JE (2018) Debunking the myth of exerciseinduced immune suppression: redefining the impact of exercise on immunological health across the lifespan. Front Immunol 9:648
- 391. Chubak J, McTiernan A, Sorensen B, Wener MH, Yasui Y, Velasquez M, Wood B, Rajan KB, Wetmore CM, Potter JD (2006) Moderate-intensity exercise reduces the incidence of colds among postmenopausal women. Am J med 119:937–942
- 392. Barrett B, Hayney MS, Muller D, Rakel D, Brown R, Zgierska AE, Barlow S, Hayer S, Barnet JH, Torres ER (2018) Meditation or exercise for preventing acute respiratory infection (MEPARI-2): a randomized controlled trial. PLoS ONE 13:e0197778
- 393. Durigon TS, MacKenzie B, Oliveira-Junior MC, Santos-Dias A, De Angelis K, Malfitano C, Palma RK, Guerra JM, Damaceno-Rodrigues NR, Caldini EG (2018) Aerobic exercise protects from Pseudomonas aeruginosa-induced pneumonia in elderly mice. J Innate Immun 10:279–290
- 394. Olivo CR, Miyaji EN, Oliveira MLS, Almeida FM, Lourenço JD, Abreu RM, Arantes PM, Lopes FD, Martins MA (2014) Aerobic exercise attenuates pulmonary inflammation induced by Streptococcus pneumoniae. J Appl Physiol 117:998–1007
- Williams PT (2014) Dose-response relationship between exercise and respiratory disease mortality. Med Sci Sports Exerc 46:711
- 396. Nieman DC, Wentz LM (2019) The compelling link between physical activity and the body's defense system. J Sport Health Sci 8:201–217
- 397. Yang J, Petitjean SJ, Koehler M, Zhang Q, Dumitru AC, Chen W, Derclaye S, Vincent SP, Soumillion P, Alsteens D (2020) Molecular interaction and inhibition of SARS-CoV-2 binding to the ACE2 receptor. Nat Commun 11:1–10
- 398. Guimarães GG, Santos SH, Oliveira ML, Pimenta-Velloso EP, Motta DF, Martins AS, Alenina N, Bader M, Santos RA, Campagnole-Santos MJ (2012) Exercise induces renin–angiotensin system unbalance and high collagen expression in the heart of mas-deficient mice. Peptides 38:54–61
- 399. Prata LO, Rodrigues CR, Martins JM, Vasconcelos PC, Oliveira FMS, Ferreira AJ, Rodrigues-Machado MdG, Caliari MV (2017) ACE2 activator associated with physical exercise potentiates the reduction of pulmonary fibrosis. Exp Biol Med 242:8–21

- 400. Magalhães DM, Nunes-Silva A, Rocha GC, Vaz LN, de Faria MHS, Vieira ELM, Rocha NP, e Silva ACS (2020) Two protocols of aerobic exercise modulate the counter-regulatory axis of the renin-angiotensin system. Heliyon 6:e03208
- 401. Frantz EDC, Prodel E, Braz ID, Giori IG, Bargut TCL, Magliano DAC, Nobrega ACL (2018) Modulation of the renin–angiotensin system in white adipose tissue and skeletal muscle: focus on exercise training. Clin Sci 132:1487–1507
- 402. Nunes-Silva A, Rocha GC, Magalhaes DM, Vaz LN, de Salviano Faria MH, Simoes e Silva AC (2017) Physical exercise and ACE2-angiotensin-(1–7)-mas receptor axis of the renin angiotensin system. Protein Peptide Lett 24:809–816
- 403. Tyrankiewicz U, Olkowicz M, Berkowicz P, Jablonska M, Smolenski RT, Zoladz JA, Chlopicki S (2021) Physical activity and inhibition of ACE additively modulate ACE/ACE-2 balance in heart failure in mice. Front Pharmacol 12:1207
- 404. Alves CR, Fernandes T, Lemos JR Jr, Magalhaes FdC, Trombetta IC, Alves GB, Mota GdFAd, Dias RG, Pereira AC, Krieger JE (2018) Aerobic exercise training differentially affects ACE C-and N-domain activities in humans: interactions with ACE I/D polymorphism and association with vascular reactivity. J Ren-Angiotensin-Aldosterone Syst. https://doi.org/10.1177/14703 20318761725
- 405. Santos R, Ferreira AJ, Verano-Braga T, Bader M (2013) Angiotensin-converting enzyme 2, angiotensin-(1–7) and Mas: new players of the renin-angiotensin system. J Endocrinol 216:R1–R17
- 406. Silva SD Jr, Jara ZP, Peres R, Lima LS, Scavone C, Montezano AC, Touyz RM, Casarini DE, Michelini LC (2017) Temporal changes in cardiac oxidative stress, inflammation and remodeling induced by exercise in hypertension: role for local angiotensin II reduction. PLoS ONE 12:e0189535
- 407. Prasannarong M, Santos FR, Henriksen EJ (2012) ANG-(1–7) reduces ANG II-induced insulin resistance by enhancing Akt phosphorylation via a Mas receptor-dependent mechanism in rat skeletal muscle. Biochem Biophys Res Commun 426:369–373
- 408. Muñoz MC, Giani JF, Burghi V, Mayer MA, Carranza A, Taira CA, Dominici FP (2012) The Mas receptor mediates modulation of insulin signaling by angiotensin-(1–7). Regul Pept 177:1–11
- 409. Marshall RP, Gohlke P, Chambers RC, Howell DC, Bottoms SE, Unger T, McAnulty RJ, Laurent GJ (2004) Angiotensin II and the fibroproliferative response to acute lung injury. Am J Physiol-Lung Cell Mol Physiol 286:L156–L164
- Evangelista FS (2020) Physical exercise and the renin angiotensin system: prospects in the COVID-19. Front Physiol 11:1282
- 411. Sandoval J, Del Valle-Mondragón L, Masso F, Zayas N, Pulido T, Teijeiro R, Gonzalez-Pacheco H, Olmedo-Ocampo R, Sisniega C, Paez-Arenas A (2020) Angiotensin converting enzyme 2 and angiotensin (1–7) axis in pulmonary arterial hypertension. Eur Respir J. https://doi.org/10.1183/13993003.02416-2019
- 412. Heffernan KS, Jae SY (2020) Exercise as medicine for COVID-19: An ACE in the hole? Med Hypotheses 142:109835
- Shoelson SE, Herrero L, Naaz A (2007) Obesity, inflammation, and insulin resistance. Gastroenterology 132:2169–2180
- 414. Cho J, Lee I, Kim D, Koh Y, Kong J, Lee S, Kang H (2014) Effect of aerobic exercise training on non-alcoholic fatty liver disease induced by a high fat diet in C57BL/6 mice. J Exerc Nutr Biochem 18:339
- 415. Murawska-Cialowicz E, Wojna J, Zuwala-Jagiello J (2015) Crossfit training changes brain-derived neurotrophic factor and irisin levels at rest, after wingate and progressive tests, and improves aerobic capacity and body composition of young physically active men and women. J Physiol Pharmacol 66:811–821
- 416. Barfield J, Anderson A (2014) Effect of crossfit<sup>™</sup> on healthrelated physical fitness: a pilot study. J Sport Human Perform 2:23–28

- 417. Brisebois MF, Rigby BR, Nichols DL (2018) Physiological and fitness adaptations after eight weeks of high-intensity functional training in physically inactive adults. Sports 6:146
- 418. Cocks M, Shaw CS, Shepherd SO, Fisher JP, Ranasinghe AM, Barker TA, Tipton KD, Wagenmakers AJ (2013) Sprint interval and endurance training are equally effective in increasing muscle microvascular density and eNOS content in sedentary males. J Physiol 591:641–656
- 419. Shepherd SO, Cocks M, Tipton K, Ranasinghe AM, Barker TA, Burniston JG, Wagenmakers AJ, Shaw CS (2013) Sprint interval and traditional endurance training increase net intramuscular triglyceride breakdown and expression of perilipin 2 and 5. J Physiol 591:657–675
- 420. Egan B, Carson BP, Garcia-Roves PM, Chibalin AV, Sarsfield FM, Barron N, McCaffrey N, Moyna NM, Zierath JR, O'Gorman DJ (2010) Exercise intensity-dependent regulation of peroxisome proliferator-activated receptor  $\gamma$  coactivator-1 $\alpha$  mRNA abundance is associated with differential activation of upstream signalling kinases in human skeletal muscle. J Physiol 588:1779–1790
- 421. Wojtaszewski JF, Nielsen P, Hansen BF, Richter EA, Kiens B (2000) Isoform-specific and exercise intensity-dependent activation of 5'-AMP-activated protein kinase in human skeletal muscle. J Physiol 528:221–226
- 422. Wende AR, Schaeffer PJ, Parker GJ, Zechner C, Han D-H, Chen MM, Hancock CR, Lehman JJ, Huss JM, McClain DA (2007) A role for the transcriptional coactivator PGC-1α in muscle refueling. J Biol Chem 282:36642–36651
- 423. Wu Z, Puigserver P, Andersson U, Zhang C, Adelmant G, Mootha V, Troy A, Cinti S, Lowell B, Scarpulla RC (1999) Mechanisms controlling mitochondrial biogenesis and respiration through the thermogenic coactivator PGC-1. Cell 98:115–124
- 424. Yoshimura S, Nakashima S, Tomiga Y, Kawakami S, Uehara Y, Higaki Y (2018) Short-and long-term effects of high-fat diet feeding and voluntary exercise on hepatic lipid metabolism in mice. Biochem Biophys Res Commun 507:291–296
- 425. Zechner R, Kienesberger PC, Haemmerle G, Zimmermann R, Lass A (2009) Adipose triglyceride lipase and the lipolytic catabolism of cellular fat stores. J Lipid Res 50:3–21
- 426. Marinho R, Ropelle ER, Cintra DE, De Souza CT, Da Silva ASR, Bertoli FC, Colantonio E, D'Almeida V, Pauli JR (2012) Endurance exercise training increases APPL1 expression and improves insulin signaling in the hepatic tissue of diet-induced obese mice, independently of weight loss. J Cell Physiol 227:2917–2926
- 427. Watanabe M, Houten SM, Wang L, Moschetta A, Mangelsdorf DJ, Heyman RA, Moore DD, Auwerx J (2004) Bile acids lower triglyceride levels via a pathway involving FXR, SHP, and SREBP-1c. J Clin Investig 113:1408–1418
- 428. Motahari Rad M, Bijeh N, Attarzadeh Hosseini SR, Raouf Saeb A (2020) The effect of two concurrent exercise modalities on serum concentrations of FGF21, irisin, follistatin, and myostatin in men with type 2 diabetes mellitus. Arch Physiol Biochem. https://doi.org/10.1080/13813455.2020.1829649
- 429. Czarkowska-Paczek B, Zendzian-Piotrowska M, Bartlomiejczyk I, Przybylski J, Gorski J (2009) The effect of acute and prolonged endurance exercise on transforming growth factor-beta1 generation in rat skeletal and heart muscle. J physiol pharmacol 60:157–162
- 430. Hulmi JJ, Oliveira BM, Silvennoinen M, Hoogaars WM, Pasternack A, Kainulainen H, Ritvos O (2013) Exercise restores decreased physical activity levels and increases markers of autophagy and oxidative capacity in myostatin/activin-blocked mdx mice. Am J Physiol-Endocrinol Metabol 305:E171–E182
- 431. Silva R, Bueno P, Avó L, Nonaka K, Selistre-Araújo H, Leal A (2014) Effect of physical training on liver expression of activin

A and follistatin in a nonalcoholic fatty liver disease model in rats. Braz J Med Biol Res 47:746–752

- 432. Pervin S, Reddy ST, Singh R (2021) Novel roles of follistatin/ myostatin in transforming growth factor-β signaling and adipose browning: Potential for therapeutic intervention in obesity related metabolic disorders. Front Endocrinol 12:653179
- 433. Hakala K, Stenius-Aarniala B, Sovija A (2000) Effects of weight loss on peak flow variability, airways obstruction, and lung volumes in obese patients with asthma. Chest 118:1315–1321
- 434. Aaron SD, Fergusson D, Dent R, Chen Y, Vandemheen KL, Dales RE (2004) Effect of weight reduction on respiratory function and airway reactivity in obese women. Chest 125:2046–2052
- 435. Pakhale S, Baron J, Dent R, Vandemheen K, Aaron SD (2015) Effects of weight loss on airway responsiveness in obese adults with asthma: does weight loss lead to reversibility of asthma? Chest 147:1582–1590
- 436. Peters U, Hernandez P, Dechman G, Ellsmere J, Maksym G (2016) Early detection of changes in lung mechanics with oscillometry following bariatric surgery in severe obesity. Appl Physiol Nutr Metab 41:538–547
- 437. Thomas P, Cowen E, Hulands G, Milledge J (1989) Respiratory function in the morbidly obese before and after weight loss. Thorax 44:382–386
- 438. Womack CJ, Harris DL, Katzel LI, Hagberg JM, Bleecker ER, Goldberg AP (2000) Weight loss, not aerobic exercise, improves pulmonary function in older obese men. J Gerontol A 55:M453–M457
- 439. Chapman DG, Irvin CG, Kaminsky DA, Forgione PM, Bates JH, Dixon AE (2014) Influence of distinct asthma phenotypes on lung function following weight loss in the obese. Respirology 19:1170–1177
- 440. Dixon AE, Pratley RE, Forgione PM, Kaminsky DA, Whittaker-Leclair LA, Griffes LA, Garudathri J, Raymond D, Poynter ME, Bunn JY (2011) Effects of obesity and bariatric surgery on airway hyperresponsiveness, asthma control, and inflammation. J Allergy Clin Immunol 128:508–515
- 441. Razi O, Tartibian B, Teixeira AM, Zamani N, Govindasamy K, Suzuki K, Laher I, Zouhal H (2022) Thermal dysregulation in patients with multiple sclerosis during SARS-CoV-2 infection. The potential therapeutic role of exercise. Mult scler relat disord. https://doi.org/10.1016/j.msard.2022.103557
- 442. Périard JD, Travers GJ, Racinais S, Sawka MN (2016) Cardiovascular adaptations supporting human exercise-heat acclimation. Auton Neurosci 196:52–62
- 443. Nadel E (1988) Temperature regulation and prolonged exercise. Prolonged exercise.
- 444. Takeda R, Okazaki K (2018) Body temperature regulation during exercise and hyperthermia in diabetics. In: Ahmed RG (ed) Diabetes and its complications. London
- 445. Werner J (1993) Temperature regulation during exercise: an overview. Exercise, heat thermoregul. https://doi.org/10. 1055/s-2007-971967
- 446. Geor R, McCutcheon L (1996) Influence of training on exerciseassociated heat tolerance in thoroughbred horses. J Sports Sci 14:349–349
- 447. Rowell LB (2011) Cardiovascular adjustments to thermal stress. In: Rowell LB (ed) Comprehensive physiology. Wiley, pp 967–1023
- 448. Moien-Afshari F, Ghosh S, Khazaei M, Kieffer T, Brownsey R, Laher I (2008) Exercise restores endothelial function independently of weight loss or hyperglycaemic status in db/db mice. Diabetologia 51:1327–1337
- 449. Woodman CR, Thompson MA, Turk JR, Laughlin MH (2005) Endurance exercise training improves endothelium-dependent relaxation in brachial arteries from hypercholesterolemic male pigs. J Appl Physiol 99:1412–1421

- 451. Zhang Y, Li X, Pitzer AL, Chen Y, Wang L, Li P-L (2015) Coronary endothelial dysfunction induced by nucleotide oligomerization domain-like receptor protein with pyrin domain containing 3 inflammasome activation during hypercholesterolemia: beyond inflammation. Antioxid Redox Signal 22:1084–1096
- 452. Ellsworth ML, Sprague RS (2012) Regulation of blood flow distribution in skeletal muscle: role of erythrocyte-released ATP. J Physiol 590:4985–4991
- 453. Hellsten Y, Maclean D, Gr R, Saltin B, Bangsbo J (1998) Adenosine concentrations in the interstitium of resting and contracting human skeletal muscle. Circulation 98:6–8
- 454. Singel DJ, Stamler JS (2005) Chemical physiology of blood flow regulation by red blood cells: the role of nitric oxide and S-nitrosohemoglobin. Annu Rev Physiol 67:99
- 455. Burnstock G, Arnett TR, Orriss IR (2013) Purinergic signalling in the musculoskeletal system. Purinergic Signal 9:541–572
- 456. Frandsen U, Bangsbo J, Langberg H, Saltin B, Hellsten Y (2000) Inhibition of nitric oxide synthesis by systemic NG-monomethyl-L-arginine administration in humans: effects on interstitial adenosine, prostacyclin and potassium concentrations in resting and contracting skeletal muscle. J Vasc Res 37:297–302

- 457. Hellsten Y, Nyberg M, Jensen L, Mortensen S (2012) Vasodilator interactions in skeletal muscle blood flow regulation. J Physiol 590:6297–6305
- 458. Huang A, Sun D, Koller A (2000) Shear stress-induced release of prostaglandin H2 in arterioles of hypertensive rats. Hypertension 35:925–930
- 459. Mortensen SP, González-Alonso J, Bune LT, Saltin B, Pilegaard H, Hellsten Y (2009) ATP-induced vasodilation and purinergic receptors in the human leg: roles of nitric oxide, prostaglandins, and adenosine. Am J Physiol-Regulatory, Integr Comparative Physiol 296:R1140–R1148
- 460. Mortensen SP, Nyberg M, Thaning P, Saltin B, Hellsten Y (2009) Adenosine contributes to blood flow regulation in the exercising human leg by increasing prostaglandin and nitric oxide formation. Hypertension 53:993–999
- Lorenzo S, Minson CT (2010) Heat acclimation improves cutaneous vascular function and sweating in trained cyclists. J Appl Physiol 109:1736–1743

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.