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OPEN Neuronal adenosine A_{2A} receptors signal ergogenic effects of caffeine

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Caffeine is one of the most used ergogenic aid for physical exercise and sports. However, its mechanism of action is still controversial. The adenosinergic hypothesis is promising due to the pharmacology of caffeine, a nonselective antagonist of adenosine A₁ and A_{2A} receptors. We now investigated A_{2A}R as a possible ergogenic mechanism through pharmacological and genetic inactivation. Forty-two adult females (20.0 ± 0.2 g) and 40 male mice (23.9 ± 0.4 g) from a global and forebrain A_{2A}R knockout (KO) colony ran an incremental exercise test with indirect calorimetry (VO₂ and RER). We administered caffeine (15 mg/kg, i.p., nonselective) and SCH 58261 (1 mg/kg, i.p., selective A_{2A}R antagonist) 15 min before the open field and exercise tests. We also evaluated the estrous cycle and infrared temperature immediately at the end of the exercise test. Caffeine and SCH 58621 were psychostimulant. Moreover, Caffeine and SCH 58621 were ergogenic, that is, they increased VO₂max, running power, and critical power, showing that A_{2A}R antagonism is ergogenic. Furthermore, the ergogenic effects of caffeine were abrogated in global and forebrain A_{2A}R KO mice, showing that the antagonism of $A_{2A}R$ in forebrain neurons is responsible for the ergogenic action of caffeine. Furthermore, caffeine modified the exercising metabolism in an A_{2A}R-dependent manner, and A_{2A}R was paramount for exercise thermoregulation.

The natural plant alkaloid caffeine (1,3,7-trimethylxantine) is one of the most common ergogenic substances for physical activity practitioners and athletes¹⁻¹⁰. Caffeine increases endurance^{1,8-12}, intermittent^{7,13,14} and $resistance ^{4,15}\ exercise\ in\ humans.\ In\ rodents,\ its\ ergogenic\ effects\ are\ conserved\ because\ caffeine\ increases\ run-product of the conserved of the conserved$ ning time on the treadmill at constant 16,17 and accelerated speeds 18,19. Sports sciences promote nonselective phosphodiesterase (PDE) inhibition^{7,8} and increased calcium mobilization^{2,7,8} as mechanisms for these ergogenic effects. However, the primary pharmacological effect of caffeine is the nonselective antagonism of adenosine A₁ and A_{2A} receptors $(A_1R, A_{2A}R)^{20-23}$.

Adenosine can act as an inhibitory modulator of the Central Nervous System (CNS) associated with tiredness and drowsiness²⁴⁻²⁹. During exercise, circulating ADP/AMP/adenosine levels increase due to ATP hydrolysis^{30,31}. However, there is still no substantial evidence on the role of adenosine in exercise-induced fatigue. It is just known that the nonselective A₁R and A_{2A}R agonist 5'-(N-ethylcarboxamido)adenosine (NECA), injected into the rat brain, abolishes the ergogenic effects of caffeine¹⁶.

Since there is increasing evidence that the adenosine modulation system critically controls allostasis²⁹ and $A_{2A}R$ have a crucial role in the ability of caffeine to normalize brain function³⁰, we hypothesized that caffeine decreases fatigue during exercise through antagonism of $A_{2A}R$ in the CNS. We combined the use of pharmacology (SCH 58261 and caffeine) and transgenic mice with tissue-selective deletion of $A_{2A}R$, to test this hypothesis in an incremental running test with indirect calorimetry (or ergospirometry). A_{2A}R knockout (KO) mice allow assessing if the ergogenic effect of caffeine persists in the absence of A2AR; the use of SCH 58261, the current reference for $A_{2A}R$ antagonists^{32,33}, allows directly assessing the ergogenic role of $A_{2A}R$. SCH 58261 has excellent selectivity and affinity for $A_{2A}R^{32,33}$, and affords motor benefits in animal models of Parkinson's disease as does caffeine, supporting the recent FDA approval of the $A_{2A}R$ antagonist Istradefylline for PD treatment³³. Our goal is to assess the ergogenicity of $A_{2A}R$ using the pharmacological and genetic tools described above.

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Methods

Animals and A_{2A}R KO colony. We used 40 male $(23.9\pm0.4 \text{ g}, 8-10 \text{ weeks old})$ and 42 female mice $(20.0\pm0.2 \text{ g}, 8-10 \text{ weeks old})$ from our global-A_{2A}R (A_{2A}R KO) and forebrain-A_{2A}R KO (fb-A_{2A}R KO) inbred colony^{34,35} and wild type littermates. The sample size for ANOVA comparison had $\alpha = 0.05$ and $\beta = 0.8$.

The inactivation of exon 2 of $A_{2A}R$ in a near congenic (N6) C57BL/6 genetic background was the method of generating $A_{2A}R$ KO mice^{36,37}. We also have good experience with treadmill running in this strain^{38–40}. $A_{2A}R$ KO mice and wild type littermates were matched for sex and age for each experiment. The Cre-loxP strategy, crossing floxed $A_{2A}R$ mice with mice expressing CRE under the forebrain-selective promoter CAM-kinase 2, allows generating fb- $A_{2A}R$ KO mice, as previously described^{34,41}. We used global $A_{2A}R$ KO females and fb- $A_{2A}R$ KO males due to the characteristic of our colony.

Mice were housed in collective cages in HEPA-filtered ventilated racks (n = 3-5) under a controlled environment (12 h light–dark cycle, lights on at 7 AM, and room temperature of 21 ± 1 °C) with ad libitum access to food and water. Housing and handling were performed according to European Union guidelines (2010/63). The Ethical Committee of the Center for Neuroscience and Cell Biology (University of Coimbra) approved the study.

Drugs. 7-(2-phenylethyl)-5-amino-2-(2-furyl)-pyrazolo-[4,3-e]-1,2,4-triazolo[1,5-c]pyrimidine (SCH 58261) was solubilized in 10% dimethyl sulfoxide (DMSO) in 0.9% NaCl – saline. Caffeine was dissolved in saline. SCH 58261 and caffeine were freshly prepared and administered intraperitoneally (volume of 10 mL/kg body mass). Caffeine, DMSO, and NaCl were obtained from Sigma-Aldrich and SCH 58261 from Tocris. The doses used of SCH 58261 (1 mg/kg) and caffeine (15 mg/kg) were based on our previous experience in the use of these compounds^{42,43} and pilot studies.

Experimental design. Fig.S1 shows the experimental design. The habituation of handling, injections (0.9% NaCl, i.p.), and moving treadmill (15 cm/s) occurred in the first three days of the experiment. The animals were treated with SCH 58261 (1 mg/kg, i.p.) and caffeine (15 mg/kg, i.p.) on days 4 and 5, 15 min before testing in the open field (4th day) and ergospirometry (5th day). The experiments took place between 9 AM and 5 PM, within the light phase of the mouse dark/light cycle, in a sound-attenuated and temperature/humidity controlled room $(20.3\pm0.6\,^{\circ}\text{C}$ and $62.8\pm0.4\%\,\text{H}_2\text{O})$ under low-intensity light ($\approx 10\,\text{lx}$). The open field apparatus and the treadmill were cleaned with 10% ethanol between individual experiments. The allocation for the experimental groups was random. For each test, the experimental unit was an individual animal.

Open field. Mice explored an unaccustomed open field (38×38 cm) for 15 min. Locomotion was analyzed using an ANY-Maze video tracking system (Stoelting Co.).

Ergospirometry. Mice were accustomed to a single-lane treadmill (Panlab LE8710, Harvard apparatus) at speed 15 cm/s (10 min, slope 5°, 0.2 mA) with a 24 h interval between each habituation session (Fig. S1). The incremental running protocol started at 15 cm/s, with an increment of 5 cm/s every 2 min at 5° inclination⁴⁰. The exercise lasted until running exhaustion, defined by the inability of the animal to leave the electrical grid for 5 s^{40,44}.

Oxygen uptake (VO₂) and carbon dioxide production (VCO₂) were estimated in a metabolic chamber (Gas Analyzer ML206, $23 \times 5 \times 5$ cm, AD Instruments, Harvard) coupled to the treadmill. The animals remained in the chamber for 15 min before exercise testing. Atmospheric air (\approx 21% O₂, \approx 0.03% CO₂) was renewed at a rate 120 mL/min, using the same sampling rate for the LASER oxygen sensor (Oxigraf X2004, resolution 0.01%) and infrared carbon dioxide sensor (Servomex Model 15050, resolution 0.1%).

We estimated the running and critical power output for a treadmill based on a standard conversion of the vertical work, body weight, and running speed^{40,45,46}. Running power is the sum (Σ) of all stages of the exercise test, and critical power is the running work performed above VO₂max.

Vaginal cytology. We evaluated the estrous cycle immediately after the exercise test, through 4–5 consecutive vaginal lavages (with 40–50 μ L of distilled H₂O) then mounted on gelatinized slides (76×26 mm)^{47,48}. These procedures lasted no more than 3–5 min, and there were no significant time delays between behavioral experiments and fluid collection for vaginal cytology.

The vaginal smear was desiccated at room temperature and covered with 0.1% crystal violet for 1 min, then twice washed with 1 mL $\rm H_2O$ and desiccated at room temperature^{47,48}. The slides were mounted with Eukitt medium (Sigma-Aldrich) and evaluated under an optical microscope at 1x, 5x, and 20x (Zeiss Axio Imager 2). We evaluated three cell types for determining the estrous cycle: nucleated epithelial cells, cornified epithelial cells, and leukocytes. Cellular prevalence defined proestrus (nucleated), estrus (cornified), metestrus (all types in the same proportion), and diestrus (leukocytes)^{47,48}.

Thermal imaging. An infrared (IR) camera (FLiR C2, emissivity 0.95, FLiR Systems) placed overtop (25 cm height) of a plastic tube (25 cm diameter) was used to acquire a static dorsal thermal image^{40,49,50}. IR images were taken immediately before and after exercise tests, namely at resting and recovery (Fig. 1H), respectively. IR images were analyzed with FLiR Tools software (Flir, Boston).

Statistics. Data are presented as mean ± SEM in graphs built using the GraphPad Prism version 5.00 for Windows, GraphPad Software, San Diego California USA, www.graphpad.com.

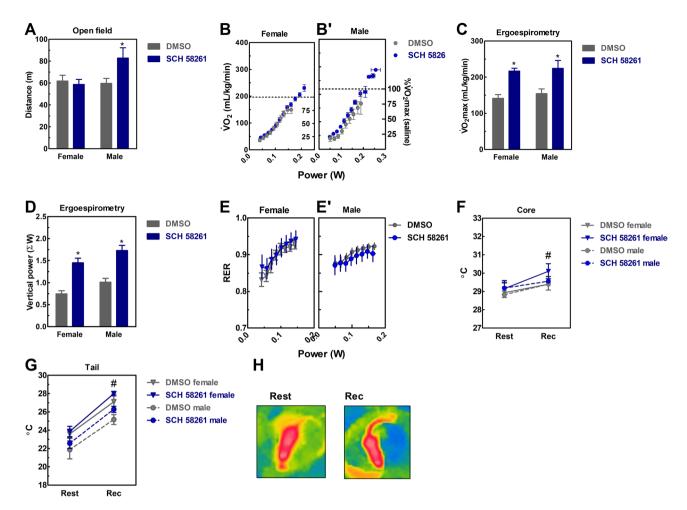


Figure 1. Effects of SCH 58261 (1 mg/kg, i.p.) on locomotion (**A**), ergospirometry (**B**–**E**), and thermoregulation (**F**–**H**) of wild type male and female mice. (**A**) SCH 58261 was psychostimulant only in males. (**B**) The dotted line represents the VO₂max of the DMSO group. Ergospirometry increased VO₂ (**B**), running power (**B**), and metabolic rate (**E**) until the animals reached fatigue. SCH 58261 was ergogenic in both sexes, as it increased VO₂max (**C**) and running power (**D**). The animals presented exercise-induced core and tail hyperthermia (**H**), which was not 58261 modified by SCH (**F**–**G**). Sex was a significant factor in decreasing maximum responses to VO₂ (**C**) and running power (**D**), and increasing core and tail temperature in females. Data are presented as mean \pm SEM. N = 8–9 animals/group for 12 independent experiments. *P<0.05 Vs. DMSO (Two-way ANOVA followed by Newman-Keuls post hoc test). #P<0.05 Vs. rest (Repeated measures ANOVA followed by Bonferroni post hoc test). DMSO dimethyl sulfoxide. Rec recovery. RER Respiratory Exchange Ratio. VO_2 oxygen consumption.

Statistical analyzes were performed according to an intention-to-treat principle using StatSoft, Inc. (2007). STATISTICA (data analysis software system), version 8.0. www.statsoft.com. ANOVA two-way was used to evaluate open field, VO_2 max, running power, and resting and recovery temperature, followed by Newman-Keuls post hoc test. The evolution of submaximal VO_2 , running power, respiratory exchange ratio (RER), and heating were evaluated by ANOVA for repeated measures followed by Bonferroni post hoc test. The differences were considered significant when P < 0.05.

Effect sizes (Cohen's partial eta-square η^2) were calculated for between-group changes in mean differences for VO₂max, running power, and temperature, where a Cohen's η^2 was used for ANOVA, defined as 0.01 small, 0.09 medium, and 0.25 large.

Results

SCH 58261: pharmacological inactivation of $A_{2A}R$ is ergogenic. SCH 58261 was psychostimulant for males, but not for females, since SCH 58261 only increased male locomotion in the open field ($F_{1,39} = 4.5$, $\eta^2 = 0.1$, $\beta = 0.54$, 95% CI 58.8–72.1, P < 0.05, Fig. 1A).

The running power of females ($F_{7,77}$ = 221, P < 0.05, Fig. 1B) and males ($F_{7,84}$ = 183, P < 0.05, Fig. 1B') increased at each stage of the exercise test. Submaximal VO₂ also increased to the maximum (VO₂max, dotted line) of females ($F_{8,77}$ = 168, P < 0.05, Fig. 1B) and males ($F_{7,84}$ = 14.3, P < 0.05, Fig. 1B'). Female ($F_{8,70}$ = 180, P < 0.05,

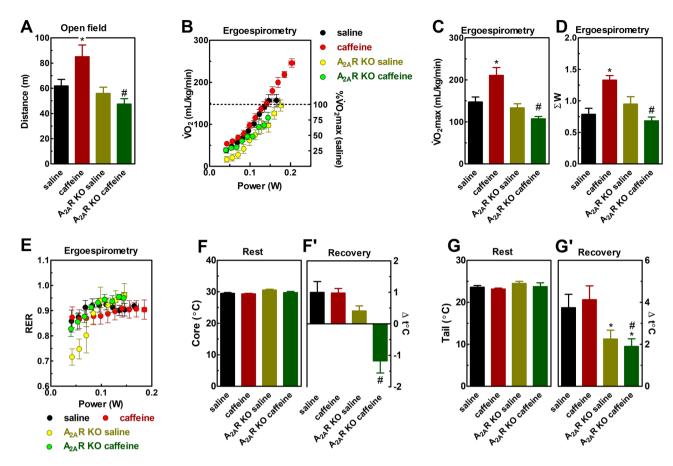


Figure 2. Effects of caffeine (15 mg/kg, i.p.) in wild type and global $A_{2A}R$ KO mice on locomotion (**A**), ergospirometry (**B**-**E**), and thermoregulation (**F**-**G**) of female mice. (**A**) Caffeine displayed a psychostimulant effect in the open field in wild type mice, but not in $A_{2A}R$ KO mice. Ergospirometry increased VO_2 (**B**), running power (**B**), and metabolic rate (**E**) until the animals reached fatigue. The dotted line represents the VO_2 max of the wild type-saline group (**B**). Caffeine increased VO_2 max (**C**) and running power (**D**) of wild type, but not $A_{2A}R$ KO mice. Exercise test induced hyperthermia, which was not affected by caffeine in wild type mice, whereas caffeine caused a hypothermic response in $A_{2A}R$ KO mice (**F** and **G**). Genotype was a significant factor for VO_2 max (**C**), running power (**D**), resting (**F**), and recovery (**F**' and **G**') temperatures. Data are described as mean \pm SEM. N = 8 - 9 animals/group for 12 independent experiments. *P < 0.05 vs. saline and #P < 0.05 vs. caffeine (Two-way ANOVA followed by Newman-Keuls post hoc test). $A_{2A}R$ —adenosine A_{2A} receptor. KO—knockout. Rec recovery. RER Respiratory Exchange Ratio. VO_2 oxygen consumption.

Fig.S2A) and male ($F_{8,70} = 164$, P < 0.05, Fig.S2B) submaximal VCO₂ kinetics was similar to VO₂. SCH 58261 had no effect on these submaximal values.

We demonstrated for the first time that SCH 58261 is ergogenic since SCH 58261 increased VO₂max $(F_{1,36}=27.7, \eta^2=0.44, \beta=0.99, 95\%$ CI 0.16–0.2, P<0.5, Fig. 1C) and running power $(F_{1,35}=55, \eta^2=0.61, \beta=1.0, 95\%$ CI 1.0–1.3, P<0.05, Fig. 1D) in both sexes.

SCH 58261 had no effect on increasing RER of females ($F_{7,70}$ = 6.9, P < 0.5, η^2 = 0.43, β = 0.99, Fig. 1E) and males ($F_{7,84}$ = 9.4, η^2 = 0.57, β = 0.99, P < 0.5, Fig. 1E'). Exercise test raised the animals' core ($F_{1,26}$ = 5.5, η^2 = 0.17, β = 0.62, 95% CI 28.7–29.39, P < 0.05, Fig. 1F) and tail temperature ($F_{1,22}$ = 81, η^2 = 0.78, β = 0.99, 95% CI 24.2–25.6, P < 0.05, Fig. 1G), with no effect of SCH 58261. Figure 1 shows the heating of the mouse's tail in post-exercise recovery (rec) in relation to rest. Three females at estrus (Fig.S3C) were excluded from temperature experiments due to large exercise-induced tail hyperthermia⁴⁰. The previous results refer to females in diestrus (Fig.S3A), proestrus (Fig.S3B), and metestrus (Fig.S3D).

Caffeine is not ergogenic in global A_{2A}R knockouts. Caffeine was psychostimulant based on its ability to increase locomotion in the wild type mice ($F_{1,36} = 5.8$, $\eta^2 = 0.13$, $\beta = 0.64$, 95% CI 55.9–71.6, P < 0.05, Fig. 2A), an effect not seen in A_{2A}R KO mice.

Figure 2B shows the progressive increase in submaximal VO₂ ($F_{7,196}$ = 255, P<0.05), VCO₂ ($F_{7,196}$ = 189, P<0.05, Fig.S2C) and running power ($F_{7,210}$ =6,243, P<0.05) at speeds 35–50 cm/s, with less VO₂ for $A_{2A}R$ KO mice. Caffeine was ergogenic but only in mice expressing $A_{2A}R$. Caffeine improved VO₂max ($F_{1,33}$ =12.6, η^2 =0.28, β =0.93, 95 CI 0.12–0.17, P<0.05, Fig. 2C) and running power ($F_{1,32}$ =22.3, η^2 =0.4, β =0.99, 95% CI 0.84–1.09, P<0.05, Fig. 2D) of wild type mice. The increase in critical power was 43.1 ± 7.1% concerning controls. $A_{2A}R$ KO did not display the ergogenic effects of caffeine.

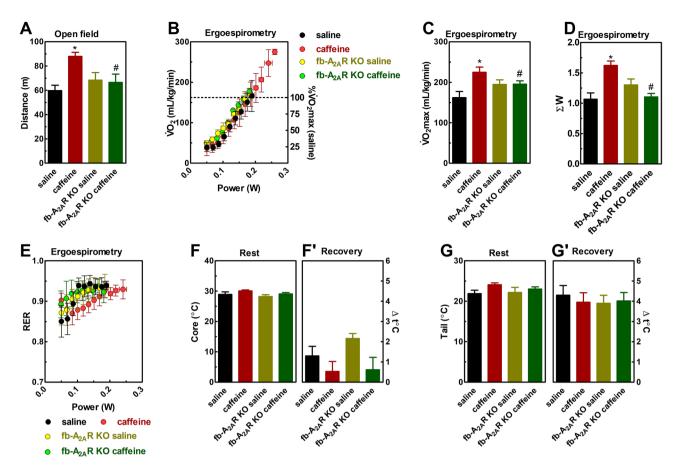


Figure 3. Effects of caffeine (15 mg/kg, i.p.) in wild type and forebrain $A_{2A}R$ KO mice on motor behavior (**A**), ergospirometry (**B**–**E**), and thermoregulation (**F**, **G**) of male mice. (**A**) Caffeine displayed a psychostimulant effect in the open field in wild type mice, but not in forebrain- $A_{2A}R$ KO mice. Ergospirometry increased VO₂ (**B**), running power (**B**), and metabolic rate (**E**) until the animals reached fatigue. The dotted line represents the VO₂max of the wild type-saline group (**B**). Caffeine increased VO₂max (**C**) and running power (**D**) of wild type, but not forebrain $A_{2A}R$ KO mice. Resting core (**F**) and tail (**G**) temperature and tail heating (**G**') were similar between groups. Data are presented as mean \pm SEM. N = 8–9 animals/group for 12 independent experiments. * $P < 0.05 \ vs.$ saline and * $P < 0.05 \ vs.$ caffeine (Two-way ANOVA followed by Newman-Keuls post hoc test). $A_{2A}R$ adenosine A_{2A} receptor, fb forebrain, KO knockout, Rec recovery, RER respiratory exchange ratio. VO₂ oxygen consumption.

Caffeine slowed the progression of RER in the wild type mice ($F_{7,133}=3.5$, $\eta^2=0.15$, $\beta=0.96$, P<0.05, Fig. 2E). Resting core (Fig. 2F) and tail (Fig. 2G) temperatures were similar between groups. Exercise increased the core ($F_{1,24}=0.16$, $\eta^2=0.99$, $\beta=0.99$, 95% CI 29.5–30.2, P<0.05, Fig. 2F') and tail ($F_{1,25}=82$, $\eta^2=0.73$, $\beta=0.99$, 95% CI 26.2–27.6, P<0.05, Fig. 2G') temperature of wild type animals. Caffeine did not change the exercise-induced core and tail heating, which was lower in the $A_{2A}R$ KO mice. Core temperature even dropped in caffeine-treated $A_{2A}R$ KO mice, as expected from the participation of A_1R , also targeted by caffeine, on the control of body temperature⁵¹.

Knocking out neuronal $A_{2A}R$ abrogates the ergogenic effects of caffeine. The psychostimulant effect of caffeine was operated by $A_{2A}R$ since caffeine increased the locomotion of wild type males in the open field $(F_{1,34}=8.6, \eta^2=0.11, \beta=0.55, 95\%$ CI 65.5–78.3, P<0.05, Fig. 3A), but not in fb- $A_{2A}R$ KO mice.

Submaximal VO₂ (F_{7,147} = 329, P < 0.05, Fig. 3B) and VCO₂ (F_{7,154} = 359, P < 0.05, Fig.S2D) increased during the exercise test without caffeine and genotype effects. Caffeine was ergogenic but only in mice expressing neuronal A_{2A}R. Caffeine increased VO₂max (F_{1,31} = 5.7, η^2 = 0.16, β = 0.64, 95% CI 0.17–0.21, P < 0.05, Fig. 3C) and running power (F_{1,29} = 4.4, η^2 = 0.13, β = 0.98, 95% CI 1.16–1.39, P < 0.05, Fig. 3D) of wild type animals. The increase in critical power was 31.9 ± 4.7% concerning controls. Most importantly, caffeine was not ergogenic in fb-A_{2A}R KO mice.

The increase in RER during the exercise test was lower in animals treated with caffeine ($F_{7,119} = 3.6$, $\eta^2 = 0.17$, $\beta = 0.97$, P < 0.05, Fig. 3E), wild type, and fb- $A_{2A}R$ KO. Resting and recovery core temperatures were similar in all groups (Figs. 3F and Fig. 3G). The exercise test did not change the core temperature (Fig. 3F'). However, exercise heated the mice's tail in a similar way between groups ($F_{1,22} = 102$, $\eta^2 = 0.69$, $\beta = 0.99$, 95% CI 24.9–26.4, P < 0.05, Fig. 3G').

Discussion

Neuronal $A_{2A}R$ **antagonism is ergogenic.** Caffeine increases exercise performance in rodents ^{16,17,19,26,40} and humans ^{1,4,8–15,24,28,51,52}. Our results show the key role of $A_{2A}R$ in the ergogenic effects of caffeine using pharmacological and genetic tools. Thus, the potent and selective $A_{2A}R$ antagonist SCH 58261 displayed an ergogenic effect similar to that of caffeine, and the ergogenic effect of caffeine was abrogated in $A_{2A}R$ KO mice.

SCH 58261 and caffeine improved VO_2 max, running and critical power of wild type mice. These results are in line with the improved running time observed in caffeine-treated rats 16,26,53 and mice 19 . Further evidence for the ergogenic effect of caffeine is based on its ability to increase muscle power and endurance output in rodents $^{54-58}$. For the first time, we demonstrated that the selective antagonism of $A_{2A}R$ is ergogenic. Also, for the first time, we demonstrated that the genetic inactivation of $A_{2A}R$ impaired the ergogenic effects of caffeine. Tissue-specific $A_{2A}R$ KO selectively in forebrain neurons further allowed showing that these ergogenic effects of caffeine are due to the antagonism of $A_{2A}R$ in forebrain neurons. Thus, we suggest that caffeine decreases central fatigue during exercise. Moreover, caffeine decreased RER in the submaximal stages of the exercise test, an effect also abrogated in $A_{2A}R$ KO mice. However, exercise-induced core and tail hyperthermia were similar among animals treated with SCH 58261 or caffeine, except for $A_{2A}R$ KO mice, suggesting possible $A_{1}R$ - $A_{2A}R$ -mediated interactions 56,57 in the temperature control 51 .

Selective A_{2A}R antagonism is psychostimulant in males, not females. We assessed the baseline motor behavior due to the motor nature of the running test, without any motor impairment found related to the different genotypes and treatments. Thus, the observed differences were not due to impaired animals' motor behavior. We also assessed the psychostimulant effects of caffeine and SCH 58261³⁴. Notably, the effects of caffeine were abrogated in A_{2A}R KO mice, and SCH 58261 did not modify locomotion in female mice. These results corroborate the robust evidence showing the psychostimulant effects of caffeine in male rodents⁵⁸. However, little is known about the role of sexual dimorphism in adenosine signaling^{59–63}. The absence of a psychostimulating effect of SCH 58261 in females is on step ahead, in notable agreement with the reported ability of the anxiolytic effect of SCH 58261 in males^{59–61} but not in females⁶⁰. However, these differences did not disturb the ergogenic effects of SCH 58261 on females. Future studies will better understand sex differences in adenosine signaling, which was not the aim of this study.

The neuropharmacology of the ergogenic effects of SCH 58261 and caffeine. Adenosine is a potent purine that modulates CNS signaling and functions from its main A_1R and $A_{2A}R^{21,23,29,62}$. Here, caffeine (nonspecific A₁R and A_{2A}R antagonist) and SCH 58261 (selective A_{2A}R antagonist) similarly increased the VO₂max, running power, and critical power of exercising male and female mice. Most importantly, these ergogenic effects were abrogated by the selective deletion of $A_{2A}R$ in forebrain neurons, which indicates the key role of CNS A_{2A}R as an ergogenic mechanism. The basal nuclei, namely the striatum, is the brain region with the highest density of $A_{2A}R^{34,35,37,63}$, which prompts the hypothesis that the $A_{2A}R$ antagonism in the basal ganglia might mediate the ergogenic effect of SCH 58261. In resting and running rodents, caffeine intake can result in a concentration of caffeine of 50 μ M in the brain ^{19,64}. This concentration is close to the EC₅₀ of caffeine (40 μ M) to antagonize A₁R and A_{2A}R in the CNS²³. Since caffeine was not ergogenic in fb-A_{2A}R KO mice, it is concluded that forebrain A_{2A}R signal the ergogenic effects of caffeine. This provides a direct demonstration of the involvement of neuronal A2AR in the ergogenic effects of caffeine, as suggested by two previous reports showing that NECA prevented the ergogenic effects of caffeine in rats¹⁶ and, conversely, that systemic caffeine reversed the poor running performance of NECA-treated rats²⁴. Although nonselective, the pharmacological use of NECA demonstrated that adenosine receptors are crucial for the ergogenic effects of caffeine 16,26,53. We now identified A_{2A}R, specifically located in forebrain neurons, as responsible for this ergogenicity of caffeine.

The neurological effects of caffeine highlight its action on the CNS. Caffeine decreases the rate of perceived exertion^{4–6,9,65}, pain^{4,6,64,66,67}, central and mental fatigue during exercise^{24,28,68,69}, indicating that caffeine attenuates mental fatigue during exercise. Caffeine also improves performance expectations⁷⁰, cognitive and executive functions^{65,71–73}, and vigor⁷⁴ in exercising subjects. The contribution of the CNS-mediated effects on exercise-induced fatigue conceptualizes central fatigue^{24,28,51,52,68,75}. Caffeine reduces saccadic eyes fatigue^{24,28},also, the cortical silence of fatigued ankle muscles^{51,52}. Moreover, caffeine increases spinal excitability⁷⁶ and cortical motor area potentiation⁶⁸ after exhausting exercise.

Caffeine decreases RER in mice expressing peripheral $A_{2A}R$. Caffeine decreases the RER in submaximal exercise in humans^{1,77,78} and rats⁷⁹. For the first time, we provide evidence that this metabolic effect involves a modification of the $A_{2A}R$ function. In the past, the inhibition of phosphodiesterase and increased intracellular calcium mobilization^{2,7,8,80} were the proposed mechanisms. However, these proposals are inconsistent with pharmacological data: caffeine has an EC_{50} for adenosine antagonism of 40 μ M, 1,000 mM for phosphodiesterase inhibition, and 3,000 mM for Ca^{2+} -triggered muscle contraction²³. Higher caffeine concentrations cause toxicity (above 200 μ M) and lethality (above 500 μ M)²³. Thus, biological effects for caffeine must be in the range below 100 μ M. We have previously shown that caffeine reaches a plasma peak of 10 μ M after caffeine intake (6 mg/kg) in running mice¹⁹. The metabolic effects of caffeine during exercise are currently associated with increased activity of the autonomic nervous system (ANS)^{1,19,77,78}, including high blood adrenaline and lactate levels, tachycardia and increased blood pressure. However, we must recognize the limitations of lungbased RER measures and their effects on metabolism, due to the possible artifacts such as hyperventilation and disturbances in the acid–base balance.

Adenosine receptors are crucial for exercise-induced hyperthermia. The temperature changes observed were dependent on sex and genotype. The exercise test improved VO_2 , an index of heat production²⁶, but the core temperature increased only in females. The tail temperature, an index of heat loss²⁶, increased in both sexes. These results are in line with previous results from our group⁴⁰. Caffeine and SCH 58261 did not modify these thermal responses to the exercise test. In males, tail heating of fb- $A_{2A}R$ KO mice was also similar to that of the wild type mice. However, the thermal response of the global $A_{2A}R$ KO females was different, indicating a peripheral role of these receptors, known to regulate body temperature⁵¹.

NECA (nonspecific A_1R and $\hat{A}_{2A}R$ agonist) causes core hypothermia in rats and rabbits^{26,76}, an effect inhibited by 8-phenyltheophylline (a potent and selective antagonist for A₁R and A_{2A}R that crosses the blood-brain barrier, BBB)⁷⁶, but unaffected by 8-(p-sulfophenyl)theophylline (another potent adenosine receptor antagonist with little BBB penetration)⁷⁶ indicating a centrally-mediated effect. In the case of A_{2A}R, its role in regulating body temperature is controversial⁸¹. The selective A_{2A}R agonist 2-p-(2-carboxyethyl)-phenethylamino-5'-Nethylcarboxamidoadenosine-hydrochloride (CGS 21680) induces hyperthermia in rats⁸² and mice⁸³. We now show that SCH 58261 does not change resting and recovery temperature. This evidence suggests that the peripheral activation of A_{2A}R can induce hypothermia in rodents, but this mechanism does not seem to be physiologically engaged as gauged by the lack of effects of the pharmacological or genetic blockade of A2AR. The previous data are from animals at rest—animals with CNS A2AR deletion present normal hyperthermia response during exercise. However, global A_{2A}R KO displays a decreased response, even hypothermia, when treated with caffeine. Thus, A_{2A}R seems to undergo a gain of function in the periphery during exercise. This data reinforces the wellknown role of A_1R in hypothermia⁸¹. Circulating adenosine levels increase during exercise^{30,31}, and global $A_{2A}R$ KO imbalance appears to increase the A₁R role, signaling hypothermia even after exercise. These results are limited to the use of infrared temperature, as we have not measured rectal temperature due to interference (vaginal manipulation) in the evaluation of the estrous cycle of females. Also, we kept the same methodology in males.

Conclusion. In summary, we have now demonstrated that $A_{2A}R$ antagonism is a mechanism of action for ergogenicity, as SCH 58261 was ergogenic. Furthermore, we showed that the antagonism of forebrain $A_{2A}R$ was the mechanism underlying the ergogenic effect of caffeine since caffeine was not ergogenic in fb- $A_{2A}R$ KO. The use of selective $A_{2A}R$ KO in forebrain neurons further reinforces the ergogenic role of caffeine in decreasing central fatigue, with possible involvement of decreased perceived exertion, pain, and mental fatigue in humans. Despite methodological limitations, our data further suggest that caffeine modified the exercising metabolism in an $A_{2A}R$ -dependent manner and that $A_{2A}R$ is essential for exercise thermoregulation.

Data availability

The datasets generated and analyzed during the current study are available from the corresponding author on reasonable request.

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Author contributions

A.S.A.Jr designed and performed the experiments, prepared the figures, and wrote the manuscript. A.E.S performed the experiments. P.M.C. designed the experiments and wrote the manuscript. R.A.C. designed the experiments and wrote the manuscript. All authors revised the manuscript.

Competing interests

RAC is a scientific consultant for the Institute for Scientific Information on Coffee (ISIC). All other authors declare no conflict of interest.

Additional information

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